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(54) Title: KINASES AND PHOSPHATASES

(57) Abstract: The invention provides human kinases and phosphatases (KAP) and polnucleotides which identify and encode KAP. The invention also provides expresson vectors, host cells, antibodies, agonists, and antagonists. The invention also provides methods for diagnosing, treating, or preventing disorders associated with aberrant expression of KAP.

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KINASES AND PHOSPHATASES

TECHNICAL FIELD

This invention relates to nucleic acid and amino acid sequences of kinases and phosphatases and to the use of these sequences in the diagnosis, treatment, and prevention of cardiovascular diseases, immune system disorders, neurological disorders, disorders affecting growth and development, lipid disorders, cell proliferative disorders, and cancers, and in the assessment of the effects of exogenous compounds on the expression of nucleic acid and amino acid sequences of kinases and phosphatases.

BACKGROUND OF THE INVENTION

Reversible protein phosphorylation is the ubiquitous strategy used to control many of the intracellular events in eukaryotic cells. It is estimated that more than ten percent of proteins active in a typical mammalian cell are phosphorylated. Kinases catalyze the transfer of high-energy phosphate groups from adenosine triphosphate (ATP) to target proteins on the hydroxyamino acid residues serine, threonine, or tyrosine. Phosphatases, in contrast, remove these phosphate groups. Extracellular signals including hormones, neurotransmitters, and growth and differentiation factors can activate kinases, which can occur as cell surface receptors or as the activator of the final effector protein, as well as other locations along the signal transduction pathway. Cascades of kinases occur, as well as kinases sensitive to second messenger molecules. This system allows for the amplification of weak signals (low abundance growth factor molecules, for example), as well as the synthesis of many weak signals into an all-or-nothing response. Phosphatases, then, are essential in determining the extent of phosphorylation in the cell and, together with kinases, regulate key cellular processes such as metabolic enzyme activity, proliferation, cell growth and differentiation, cell adhesion, and cell cycle progression.

KINASES

Kinases comprise the largest known enzyme superfamily and vary widely in their target molecules. Kinases catalyze the transfer of high energy phosphate groups from a phosphate donor to a phosphate acceptor. Nucleotides usually serve as the phosphate donor in these reactions, with most kinases utilizing adenosine triphosphate (ATP). The phosphate acceptor can be any of a variety of molecules, including nucleosides, nucleotides, lipids, carbohydrates, and proteins. Proteins are phosphorylated on hydroxyamino acids. Addition of a phosphate group alters the local charge on the acceptor molecule, causing internal conformational changes and potentially influencing intermolecular contacts. Reversible protein phosphorylation is the primary method for

regulating protein activity in eukaryotic cells. In general, proteins are activated by phosphorylation in response to extracellular signals such as hormones, neurotransmitters, and growth and differentiation factors. The activated proteins initiate the cell's intracellular response by way of intracellular signaling pathways and second messenger molecules such as cyclic nucleotides,

5 calcium-calmodulin, inositol, and various mitogens, that regulate protein phosphorylation.

Kinases are involved in all aspects of a cell's function, from basic metabolic processes, such as glycolysis, to cell-cycle regulation, differentiation, and communication with the extracellular environment through signal transduction cascades. Inappropriate phosphorylation of proteins in cells has been linked to changes in cell cycle progression and cell differentiation. Changes in the cell cycle have been linked to induction of apoptosis or cancer. Changes in cell differentiation have been linked to diseases and disorders of the reproductive system, immune system, and skeletal muscle.

There are two classes of protein kinases. One class, protein tyrosine kinases (PTKs), phosphorylates tyrosine residues, and the other class, protein serine/threonine kinases (STKs), 15 phosphorylates serine and threonine residues. Some PTKs and STKs possess structural characteristics of both families and have dual specificity for both tyrosine and serine/threonine residues. Almost all kinases contain a conserved 250-300 amino acid catalytic domain containing specific residues and sequence motifs characteristic of the kinase family. The protein kinase catalytic domain can be further divided into 11 subdomains. N-terminal subdomains I-IV fold into a 20 two-lobed structure which binds and orients the ATP donor molecule, and subdomain V spans the two lobes. C-terminal subdomains VI-XI bind the protein substrate and transfer the gamma phosphate from ATP to the hydroxyl group of a tyrosine, serine, or threonine residue. Each of the 11 subdomains contains specific catalytic residues or amino acid motifs characteristic of that subdomain. For example, subdomain I contains an 8-amino acid glycine-rich ATP binding 25 consensus motif, subdomain II contains a critical lysine residue required for maximal catalytic activity, and subdomains VI through IX comprise the highly conserved catalytic core. PTKs and STKs also contain distinct sequence motifs in subdomains VI and VIII which may confer hydroxyamino acid specificity.

In addition, kinases may also be classified by additional amino acid sequences, generally

between 5 and 100 residues, which either flank or occur within the kinase domain. These additional
amino acid sequences regulate kinase activity and determine substrate specificity. (Reviewed in
Hardie, G. and S. Hanks (1995) The Protein Kinase Facts Book, Vol I, pp. 17-20 Academic Press.

San Diego CA.). In particular, two protein kinase signature sequences have been identified in the
kinase domain, the first containing an active site lysine residue involved in ATP binding, and the
second containing an aspartate residue important for catalytic activity. If a protein analyzed

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includes the two protein kinase signatures, the probability of that protein being a protein kinase is close to 100% (PROSITE: PDOC00100, November 1995).

Protein Tyrosine Kinases

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Protein tyrosine kinases (PTKs) may be classified as either transmembrane, receptor PTKs 5 or nontransmembrane, nonreceptor PTK proteins. Transmembrane tyrosine kinases function as receptors for most growth factors. Growth factors bind to the receptor tyrosine kinase (RTK), which causes the receptor to phosphorylate itself (autophosphorylation) and specific intracellular second messenger proteins. Growth factors (GF) that associate with receptor PTKs include epidermal GF, platelet-derived GF, fibroblast GF, hepatocyte GF, insulin and insulin-like GFs, 10 nerve GF, vascular endothelial GF, and macrophage colony stimulating factor.

Nontransmembrane, nonreceptor PTKs lack transmembrane regions and, instead, form signaling complexes with the cytosolic domains of plasma membrane receptors. Receptors that function through non-receptor PTKs include those for cytokines and hormones (growth hormone and prolactin), and antigen-specific receptors on T and B lymphocytes.

Many PTKs were first identified as oncogene products in cancer cells in which PTK activation was no longer subject to normal cellular controls. In fact, about one third of the known oncogenes encode PTKs. Furthermore, cellular transformation (oncogenesis) is often accompanied by increased tyrosine phosphorylation activity (Charbonneau, H. and N.K. Tonks (1992) Annu. Rev. Cell Biol. 8:463-493). Regulation of PTK activity may therefore be an important strategy in 20 controlling some types of cancer.

Protein Serine/Threonine Kinases

Protein serine/threonine kinases (STKs) are nontransmembrane proteins. A subclass of STKs are known as ERKs (extracellular signal regulated kinases) or MAPs (mitogen-activated protein kinases) and are activated after cell stimulation by a variety of hormones and growth factors. 25 Cell stimulation induces a signaling cascade leading to phosphorylation of MEK (MAP/ERK kinase) which, in turn, activates ERK via serine and threonine phosphorylation. A varied number of proteins represent the downstream effectors for the active ERK and implicate it in the control of cell proliferation and differentiation, as well as regulation of the cytoskeleton. Activation of ERK is normally transient, and cells possess dual specificity phosphatases that are responsible for its down-30 regulation. Also, numerous studies have shown that elevated ERK activity is associated with some cancers. Other STKs include the second messenger dependent protein kinases such as the cyclic-AMP dependent protein kinases (PKA), calcium-calmodulin (CaM) dependent protein kinases, and the mitogen-activated protein kinases (MAP); the cyclin-dependent protein kinases; checkpoint and cell cycle kinases; Numb-associated kinase (Nak); human Fused (hFu); 35 proliferation-related kinases; 5'-AMP-activated protein kinases; and kinases involved in apoptosis.

One member of the ERK family of MAP kinases, ERK 7, is a novel 61-kDa protein that has motif similarities to ERK1 and ERK2, but is not activated by extracellular stimuli as are ERK1 and ERK2 nor by the common activators, c-Jun N-terminal kinase (JNK) and p38 kinase. ERK7 regulates its nuclear localization and inhibition of growth through its C-terminal tail, not through the kinase domain as is typical with other MAP kinases (Abe, M.K. (1999) Mol. Cell. Biol. 19:1301-1312).

The second messenger dependent protein kinases primarily mediate the effects of second messengers such as cyclic AMP (cAMP), cyclic GMP, inositol triphosphate, phosphatidylinositol, 3,4,5-triphosphate, cyclic ADP ribose, arachidonic acid, diacylglycerol and calcium-calmodulin.

- The PKAs are involved in mediating hormone-induced cellular responses and are activated by cAMP produced within the cell in response to hormone stimulation. cAMP is an intracellular mediator of hormone action in all animal cells that have been studied. Hormone-induced cellular responses include thyroid hormone secretion, cortisol secretion, progesterone secretion, glycogen breakdown, bone resorption, and regulation of heart rate and force of heart muscle contraction.
- 15 PKA is found in all animal cells and is thought to account for the effects of cAMP in most of these cells. Altered PKA expression is implicated in a variety of disorders and diseases including cancer, thyroid disorders, diabetes, atherosclerosis, and cardiovascular disease (Isselbacher, K.J. et al. (1994) Harrison's Principles of Internal Medicine, McGraw-Hill, New York NY, pp. 416-431, 1887).
- The casein kinase I (CKI) gene family is another subfamily of serine/threonine protein kinases. This continuously expanding group of kinases have been implicated in the regulation of numerous cytoplasmic and nuclear processes, including cell metabolism and DNA replication and repair. CKI enzymes are present in the membranes, nucleus, cytoplasm and cytoskeleton of eukaryotic cells, and on the mitotic spindles of mammalian cells (Fish, K.J. et al. (1995) J. Biol. Chem. 270:14875-14883).

The CKI family members all have a short amino-terminal domain of 9-76 amino acids, a highly conserved kinase domain of 284 amino acids, and a variable carboxyl-terminal domain that ranges from 24 to over 200 amino acids in length (Cegielska, A. et al. (1998) J. Biol. Chem. 273:1357-1364). The CKI family is comprised of highly related proteins, as seen by the identification of isoforms of casein kinase I from a variety of sources. There are at least five mammalian isoforms, α, β, γ, δ, and ε. Fish et al. identified CKI-epsilon from a human placenta cDNA library. It is a basic protein of 416 amino acids and is closest to CKI-delta. Through recombinant expression, it was determined to phosphorylate known CKI substrates and was inhibited by the CKI-specific inhibitor CKI-7. The human gene for CKI-epsilon was able to rescue yeast with a slow-growth phenotype caused by deletion of the yeast CKI locus, HRR250 (Fish et al.,

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supra).

The mammalian circadian mutation tau was found to be a semidominant autosomal allele of CKI-epsilon that markedly shortens period length of circadian rhythms in Syrian hamsters. The tau locus is encoded by casein kinase I-epsilon, which is also a homolog of the Drosophila circadian 5 gene double-time. Studies of both the wildtype and tau mutant CKI-epsilon enzyme indicated that the mutant enzyme has a noticeable reduction in the maximum velocity and autophosphorylation state. Further, in vitro, CKI-epsilon is able to interact with mammalian PERIOD proteins, while the mutant enzyme is deficient in its ability to phosphorylate PERIOD. Lowrey et al. have proposed that CKI-epsilon plays a major role in delaying the negative feedback signal within the 10 transcription-translation-based autoregulatory loop that composes the core of the circadian mechanism. Therefore the CKI-epsilon enzyme is an ideal target for pharmaceutical compounds influencing circadian rhythms, jet-lag and sleep, in addition to other physiologic and metabolic processes under circadian regulation (Lowrey, P.L. et al. (2000) Science 288:483-491). Calcium-Calmodulin Dependent Protein Kinases

Calcium-calmodulin dependent (CaM) kinases are involved in regulation of smooth muscle 15 contraction, glycogen breakdown (phosphorylase kinase), and neurotransmission (CaM kinase I and CaM kinase II). CaM dependent protein kinases are activated by calmodulin, an intracellular calcium receptor, in response to the concentration of free calcium in the cell. Many CaM kinases are also activated by phosphorylation. Some CaM kinases are also activated by 20 autophosphorylation or by other regulatory kinases. CaM kinase I phosphorylates a variety of substrates including the neurotransmitter-related proteins synapsin I and II, the gene transcription regulator, CREB, and the cystic fibrosis conductance regulator protein, CFTR (Haribabu, B. et al. (1995) EMBO J. 14:3679-3686). CaM kinase II also phosphorylates synapsin at different sites and controls the synthesis of catecholamines in the brain through phosphorylation and activation of 25 tyrosine hydroxylase. CaM kinase II controls the synthesis of catecholamines and seratonin, through phosphorylation/activation of tyrosine hydroxylase and tryptophan hydroxylase, respectively (Fujisawa, H. (1990) BioEssays 12:27-29). The mRNA encoding a calmodulin-binding protein kinase-like protein was found to be enriched in mammalian forebrain. This protein is associated with vesicles in both axons and dendrites and accumulates largely postnatally. The 30 amino acid sequence of this protein is similar to CaM-dependent STKs, and the protein binds calmodulin in the presence of calcium (Godbout, M. et al. (1994) J. Neurosci. 14:1-13).

Homeodomain-interacting protein kinases (HIPKs) are serine/threonine kinases and novel members of the DYRK kinase subfamily (Hofmann, T.G. et al. (2000) Biochimie 82:1123-1127). HIPKs contain a conserved protein kinase domain separated from a domain that interacts with 35 homeoproteins. HIPKs are nuclear kinases, and HIPK2 is highly expressed in neuronal tissue (Kim,

Y.H. et al. (1998) J. Biol. Chem. 273:25875-25879; Wang, Y. et al. (2001) Biochim. Biophys. Acta 1518:168-172). HIPKs act as corepressors for homeodomian transcription factors. This corepressor activity is seen in posttranslational modifications such as ubiquitination and phosphorylation, each of which are important in the regulation of cellular protein function (Kim,
Y.H. et al. (1999) Proc. Natl. Acad. Sci. USA 96:12350-12355).

The human h-warts protein, a homolog of Drosophila warts tumor suppressor gene, maps to chromosome 6q24-25.1. It has a serine/threonine kinase domain and is localized to centrosomes in interphase cells. It is involved in mitosis and functions as a component of the mitotic apparatus (Nishiyama, Y. et al. (1999) FEBS Lett. 459:159-165).

10 Calcium-Calmodulin Dependent Protein Kinases

Calcium-calmodulin dependent (CaM) kinases are involved in regulation of smooth muscle contraction, glycogen breakdown (phosphorylase kinase), and neurotransmission (CaM kinase I and CaM kinase II). CaM dependent protein kinases are activated by calmodulin, an intracellular calcium receptor, in response to the concentration of free calcium in the cell. Many CaM kinases 15 are also activated by phosphorylation. Some CaM kinases are also activated by autophosphorylation or by other regulatory kinases. CaM kinase I phosphorylates a variety of substrates including the neurotransmitter-related proteins synapsin I and II, the gene transcription regulator, CREB, and the cystic fibrosis conductance regulator protein, CFTR (Haribabu, B. et al. (1995) EMBO J. 14:3679-3686). CaM kinase II also phosphorylates synapsin at different sites and 20 controls the synthesis of catecholamines in the brain through phosphorylation and activation of tyrosine hydroxylase. CaM kinase II controls the synthesis of catecholamines and seratonin, through phosphorylation/activation of tyrosine hydroxylase and tryptophan hydroxylase, respectively (Fujisawa, H. (1990) BioEssays 12:27-29). The mRNA encoding a calmodulin-binding protein kinase-like protein was found to be enriched in mammalian forebrain. This protein is 25 associated with vesicles in both axons and dendrites and accumulates largely postnatally. The amino acid sequence of this protein is similar to CaM-dependent STKs, and the protein binds calmodulin in the presence of calcium (Godbout, M. et al. (1994) J. Neurosci. 14:1-13).

Mitogen-Activated Protein Kinases

The mitogen-activated protein kinases (MAP), which mediate signal transduction from the

cell surface to the nucleus via phosphorylation cascades, are another STK family that regulates intracellular signaling pathways. Several subgroups have been identified, and each manifests

different substrate-specificities and responds to distinct extracellular stimuli (Egan, S.E. and R.A.

Weinberg (1993) Nature 365:781-783). There are three kinase modules comprising the MAP kinase cascade: MAPK (MAP), MAPK kinase (MAP2K, MAPKK, or MKK), and MKK kinase (MAP3K,

MAPKKK, OR MEKK) (Wang,X.S. et al (1998) Biochem. Biophys. Res. Commun. 253:33-37).

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The extracellular-regulated kinase (ERK) pathway is activated by growth factors and mitogens, for example, epidermal growth factor (EGF), ultraviolet light, hyperosmolar medium, heat shock, or endotoxic lipopolysaccharide (LPS). The closely related though distinct parallel pathways, the c-Jun N-terminal kinase (JNK), or stress-activated kinase (SAPK) pathway, and the p38 kinase 5 pathway are activated by stress stimuli and proinflammatory cytokines such as tumor necrosis factor (TNF) and interleukin-1 (IL-1). Altered MAP kinase expression is implicated in a variety of disease conditions including cancer, inflammation, immune disorders, and disorders affecting growth and development.. MAP kinase signaling pathways are present in mammalian cells as well as in yeast.

The family of p21-activated protein kinases (PAKs) appear to be present in all organisms 10 that have Cdc42-like GTPases. In mammalian cells, PAKs have been implicated in the activation of mitogen-activated protein kinase cascades. PAK functions also include the dissolution of cytoskeletal stress fibers and reorganization of focal complexes (Manser, E. et al. (1997) Mol. Cell Biol.17(3):1129-1143).

Cyclin-Dependent Protein Kinases

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The cyclin-dependent protein kinases (CDKs) are STKs that control the progression of cells through the cell cycle. The entry and exit of a cell from mitosis are regulated by the synthesis and destruction of a family of activating proteins called cyclins. Cyclins are small regulatory proteins that bind to and activate CDKs, which then phosphorylate and activate selected proteins involved in the mitotic process. CDKs are unique in that they require multiple inputs to become activated. In 20 addition to cyclin binding, CDK activation requires the phosphorylation of a specific threonine residue and the dephosphorylation of a specific tyrosine residue on the CDK.

Another family of STKs associated with the cell cycle are the NIMA (never in mitosis)related kinases (Neks). Both CDKs and Neks are involved in duplication, maturation, and separation of the microtubule organizing center, the centrosome, in animal cells (Fry, A.M. et al. 25 (1998) EMBO J. 17:470-481).

Checkpoint and Cell Cycle Kinases

In the process of cell division, the order and timing of cell cycle transitions are under control of cell cycle checkpoints, which ensure that critical events such as DNA replication and chromosome segregation are carried out with precision. If DNA is damaged, e.g. by radiation, a 30 checkpoint pathway is activated that arrests the cell cycle to provide time for repair. If the damage is extensive, apoptosis is induced. In the absence of such checkpoints, the damaged DNA is inherited by aberrant cells which may cause proliferative disorders such as cancer. Protein kinases play an important role in this process. For example, a specific kinase, checkpoint kinase 1 (Chk1), has been identified in yeast and mammals, and is activated by DNA damage in yeast. Activation of 35 Chk1 leads to the arrest of the cell at the G2/M transition (Sanchez, Y. et al. (1997) Science

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277:1497-1501). Specifically, Chk1 phosphorylates the cell division cycle phosphatase CDC25, inhibiting its normal function which is to dephosphorylate and activate the cyclin-dependent kinase Cdc2. Cdc2 activation controls the entry of cells into mitosis (Peng, C.-Y. et al. (1997) Science 277:1501-1505). Thus, activation of Chk1 prevents the damaged cell from entering mitosis. A 5 deficiency in a checkpoint kinase, such as Chk1, may also contribute to cancer by failure to arrest cells with damaged DNA at other checkpoints such as G2/M.

Proliferation-Related Kinases

Proliferation-related kinase is a serum/cytokine inducible STK that is involved in regulation of the cell cycle and cell proliferation in human megakarocytic cells (Li, B. et al. (1996) J. Biol. 10 Chem. 271:19402-19408). Proliferation-related kinase is related to the polo (derived from Drosophila polo gene) family of STKs implicated in cell division. Proliferation-related kinase is downregulated in lung tumor tissue and may be a proto-oncogene whose deregulated expression in normal tissue leads to oncogenic transformation.

5'-AMP-activated protein kinase

A ligand-activated STK protein kinase is 5'-AMP-activated protein kinase (AMPK) (Gao, 15 G. et al. (1996) J. Biol Chem. 271:8675-8681). Mammalian AMPK is a regulator of fatty acid and sterol synthesis through phosphorylation of the enzymes acetyl-CoA carboxylase and hydroxymethylglutaryl-CoA reductase and mediates responses of these pathways to cellular stresses such as heat shock and depletion of glucose and ATP. AMPK is a heterotrimeric complex 20 comprised of a catalytic alpha subunit and two non-catalytic beta and gamma subunits that are believed to regulate the activity of the alpha subunit. Subunits of AMPK have a much wider distribution in non-lipogenic tissues such as brain, heart, spleen, and lung than expected. This distribution suggests that its role may extend beyond regulation of lipid metabolism alone.

The RET (rearranged during transfection) proto-oncogene encodes a tyrosine kinase 25 receptor involved in both multiple endocrine neoplasia type 2, an inherited cancer syndrome, and Hirschsprung disease, a developmental defect of enteric neurons. RET and its functional ligand, glial cell line-derived neurotrophic factor, play key roles in the development of the human enteric nervous system (Pachnis, V. et al. (1998) Am. J. Physiol. 275:G183-G186).

Kinases in Apoptosis

Apoptosis is a highly regulated signaling pathway leading to cell death that plays a crucial role in tissue development and homeostasis. Deregulation of this process is associated with the pathogenesis of a number of diseases including autoimmune diseases, neurodegenerative disorders, and cancer. Various STKs play key roles in this process. ZIP kinase is an STK containing a C-terminal leucine zipper domain in addition to its N-terminal protein kinase domain. This 35 C-terminal domain appears to mediate homodimerization and activation of the kinase as well as

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interactions with transcription factors such as activating transcription factor, ATF4, a member of the cyclic-AMP responsive element binding protein (ATF/CREB) family of transcriptional factors (Sanjo, H. et al. (1998) J. Biol. Chem. 273:29066-29071). DRAK1 and DRAK2 are STKs that share homology with the death-associated protein kinases (DAP kinases), known to function in interferon-γ induced apoptosis (Sanjo et al., supra). Like ZIP kinase, DAP kinases contain a C-terminal protein-protein interaction domain, in the form of ankyrin repeats, in addition to the N-terminal kinase domain. ZIP, DAP, and DRAK kinases induce morphological changes associated with apoptosis when transfected into NIH3T3 cells (Sanjo et al., supra). However, deletion of either the N-terminal kinase catalytic domain or the C-terminal domain of these proteins abolishes apoptosis activity, indicating that in addition to the kinase activity, activity in the C-terminal domain is also necessary for apoptosis, possibly as an interacting domain with a regulator or a specific substrate.

RICK is another STK recently identified as mediating a specific apoptotic pathway involving the death receptor, CD95 (Inohara, N. et al. (1998) J. Biol. Chem. 273:12296-12300).

15 CD95 is a member of the tumor necrosis factor receptor superfamily and plays a critical role in the regulation and homeostasis of the immune system (Nagata, S. (1997) Cell 88:355-365). The CD95 receptor signaling pathway involves recruitment of various intracellular molecules to a receptor complex following ligand binding. This process includes recruitment of the cysteine protease caspase-8 which, in turn, activates a caspase cascade leading to cell death. RICK is composed of an N-terminal kinase catalytic domain and a C-terminal "caspase-recruitment" domain that interacts with caspase-like domains, indicating that RICK plays a role in the recruitment of caspase-8. This interpretation is supported by the fact that the expression of RICK in human 293T cells promotes activation of caspase-8 and potentiates the induction of apoptosis by various proteins involved in the CD95 apoptosis pathway (Inohara et al., supra).

25 Mitochondrial Protein Kinases

A novel class of eukaryotic kinases, related by sequence to prokaryotic histidine protein kinases, are the mitochondrial protein kinases (MPKs) which seem to have no sequence similarity with other eukaryotic protein kinases. These protein kinases are located exclusively in the mitochondrial matrix space and may have evolved from genes originally present in respiration-dependent bacteria which were endocytosed by primitive eukaryotic cells. MPKs are responsible for phosphorylation and inactivation of the branched-chain alpha-ketoacid dehydrogenase and pyruvate dehydrogenase complexes (Harris, R.A. et al. (1995) Adv. Enzyme Regul. 34:147-162). Five MPKs have been identified. Four members correspond to pyruvate dehydrogenase kinase isozymes, regulating the activity of the pyruvate dehydrogenase complex, which is an important regulatory enzyme at the interface between glycolysis and the citric acid cycle. The fifth member

corresponds to a branched-chain alpha-ketoacid dehydrogenase kinase, important in the regulation of the pathway for the disposal of branched-chain amino acids. (Harris, R.A. et al. (1997) Adv. Enzyme Regul. 37:271-293). Both starvation and the diabetic state are known to result in a great increase in the activity of the pyruvate dehydrogenase kinase in the liver, heart and muscle of the rat. This increase contributes in both disease states to the phosphorylation and inactivation of the pyruvate dehydrogenase complex and conservation of pyruvate and lactate for gluconeogenesis (Harris (1995) supra).

KINASES WITH NON-PROTEIN SUBSTRATES

10 Lipid and Inositol kinases

PINISOCCIO: <!NO

Lipid kinases phosphorylate hydroxyl residues on lipid head groups. A family of kinases involved in phosphorylation of phosphatidylinositol (PI) has been described, each member phosphorylating a specific carbon on the inositol ring (Leevers, S.J. et al. (1999) Curr. Opin. Cell. Biol. 11:219-225). The phosphorylation of phosphatidylinositol is involved in activation of the protein kinase C signaling pathway. The inositol phospholipids (phosphoinositides) intracellular signaling pathway begins with binding of a signaling molecule to a G-protein linked receptor in the plasma membrane. This leads to the phosphorylation of phosphatidylinositol (PI) residues on the inner side of the plasma membrane by inositol kinases, thus converting PI residues to the biphosphate state (PIP₂). PIP₂ is then cleaved into inositol triphosphate (IP₃) and diacylglycerol.

These two products act as mediators for separate signaling pathways. Cellular responses that are mediated by these pathways are glycogen breakdown in the liver in response to vasopressin, smooth muscle contraction in response to acetylcholine, and thrombin-induced platelet aggregation.

PI 3-kinase (PI3K), which phosphorylates the D3 position of PI and its derivatives, has a central role in growth factor signal cascades involved in cell growth, differentiation, and metabolism. PI3K is a heterodimer consisting of an adapter subunit and a catalytic subunit. The adapter subunit acts as a scaffolding protein, interacting with specific tyrosine-phosphorylated proteins, lipid moieties, and other cytosolic factors. When the adapter subunit binds tyrosine phosphorylated targets, such as the insulin responsive substrate (IRS)-1, the catalytic subunit is activated and converts PI (4,5) bisphosphate (PIP₂) to PI (3,4,5) P₃ (PIP₃). PIP₃ then activates a number of other proteins, including PKA, protein kinase B (PKB), protein kinase C (PKC), glycogen synthase kinase (GSK)-3, and p70 ribosomal s6 kinase. PI3K also interacts directly with the cytoskeletal organizing proteins. Rac, rho, and cdc42 (Shepherd, P.R. et al. (1998) Biochem. J. 333:471-490). Animal models for diabetes, such as obese and fat mice, have altered PI3K adapter subunit levels. Specific mutations in the adapter subunit have also been found in an insulin-resistant Danish population, suggesting a role for PI3K in type-2 diabetes (Shepard, supra).

An example of lipid kinase phosphorylation activity is the phosphorylation of D-erythro-sphingosine to the sphingolipid metabolite, sphingosine-1-phosphate (SPP). SPP has emerged as a novel lipid second-messenger with both extracellular and intracellular actions (Kohama, T. et al. (1998) J. Biol. Chem. 273:23722-23728). Extracellularly, SPP is a ligand for the G-protein coupled receptor EDG-1 (endothelial-derived, G-protein coupled receptor). Intracellularly, SPP regulates cell growth, survival, motility, and cytoskeletal changes. SPP levels are regulated by sphingosine kinases that specifically phosphorylate D-erythro-sphingosine to SPP. The importance of sphingosine kinase in cell signaling is indicated by the fact that various stimuli, including platelet-derived growth factor (PDGF), nerve growth factor, and activation of protein kinase C, increase cellular levels of SPP by activation of sphingosine kinase, and the fact that competitive inhibitors of the enzyme selectively inhibit cell proliferation induced by PDGF (Kohama et al., supra).

PKC is also activated by diacylglycerol (DAG). Phorbol esters (PE) are analogs of DAG and tumor promoters that cause a variety of physiological changes when administered to cells and tissues. PE and DAG bind to the N-terminal region of PKC. This region contains one or more copies of a cysteine-rich domain about 50 amino-acid residues long and essential for DAG/PE-binding. Diacylglycerol kinase (DGK), the enzyme that converts DAG into phosphatidate, contains two copies of the DAG/PE-binding domain in its N-terminal section (Azzi, A. et al. (1992) Eur. J. Biochem. 208:547-557).

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Purine Nucleotide Kinases

The purine nucleotide kinases, adenylate kinase (ATP:AMP phosphotransferase, or AdK) and guanylate kinase (ATP:GMP phosphotransferase, or GuK) play a key role in nucleotide

35 metabolism and are crucial to the synthesis and regulation of cellular levels of ATP and GTP,

respectively. These two molecules are precursors in DNA and RNA synthesis in growing cells and provide the primary source of biochemical energy in cells (ATP), and signal transduction pathways (GTP). Inhibition of various steps in the synthesis of these two molecules has been the basis of many antiproliferative drugs for cancer and antiviral therapy (Pillwein, K. et al. (1990) Cancer Res. 50:1576-1579).

AdK is found in almost all cell types and is especially abundant in cells having high rates of ATP synthesis and utilization such as skeletal muscle. In these cells AdK is physically associated with mitochondria and myofibrils, the subcellular structures that are involved in energy production and utilization, respectively. Recent studies have demonstrated a major function for AdK in transferring high energy phosphoryls from metabolic processes generating ATP to cellular components consuming ATP (Zeleznikar, R.J. et al. (1995) J. Biol. Chem. 270:7311-7319). Thus AdK may have a pivotal role in maintaining energy production in cells, particularly those having a high rate of growth or metabolism such as cancer cells, and may provide a target for suppression of its activity in order to treat certain cancers. Alternatively, reduced AdK activity may be a source of various metabolic, muscle-energy disorders that can result in cardiac or respiratory failure and may be treatable by increasing AdK activity.

GuK, in addition to providing a key step in the synthesis of GTP for RNA and DNA synthesis, also fulfills an essential function in signal transduction pathways of cells through the regulation of GDP and GTP. Specifically, GTP binding to membrane associated G proteins 20 mediates the activation of cell receptors, subsequent intracellular activation of adenyl cyclase, and production of the second messenger, cyclic AMP. GDP binding to G proteins inhibits these processes. GDP and GTP levels also control the activity of certain oncogenic proteins such as p21^{ras} known to be involved in control of cell proliferation and oncogenesis (Bos, J.L. (1989) Cancer Res. 49:4682-4689). High ratios of GTP:GDP caused by suppression of GuK cause activation of p21^{ras} and promote oncogenesis. Increasing GuK activity to increase levels of GDP and reduce the GTP:GDP ratio may provide a therapeutic strategy to reverse oncogenesis.

GuK is an important enzyme in the phosphorylation and activation of certain antiviral drugs useful in the treatment of herpes virus infections. These drugs include the guanine homologs acyclovir and buciclovir (Miller, W.H. and R.L. Miller (1980) J. Biol. Chem. 255:7204-7207;

30 Stenberg, K. et al. (1986) J. Biol. Chem. 261:2134-2139). Increasing GuK activity in infected cells may provide a therapeutic strategy for augmenting the effectiveness of these drugs and possibly for reducing the necessary dosages of the drugs.

Pyrimidine Kinases

The pyrimidine kinases are deoxycytidine kinase and thymidine kinase 1 and 2.

35 Deoxycytidine kinase is located in the nucleus, and thymidine kinase 1 and 2 are found in the

cytosol (Johansson, M. et al. (1997) Proc. Natl. Acad. Sci. USA 94:11941-11945). Phosphorylation of deoxyribonucleosides by pyrimidine kinases provides an alternative pathway for <u>de novo</u> synthesis of DNA precursors. The role of pyrimidine kinases, like purine kinases, in phosphorylation is critical to the activation of several chemotherapeutically important nucleoside analogues (Arner E.S. and S. Eriksson (1995) Pharmacol. Ther. 67:155-186).

PHOSPHATASES

Protein phosphatases are generally characterized as either serine/threonine- or tyrosine-specific based on their preferred phospho-amino acid substrate. However, some phosphatases

10 (DSPs, for dual specificity phosphatases) can act on phosphorylated tyrosine, serine, or threonine residues. The protein serine/threonine phosphatases (PSPs) are important regulators of many cAMP-mediated hormone responses in cells. Protein tyrosine phosphatases (PTPs) play a significant role in cell cycle and cell signaling processes. Another family of phosphatases is the acid phosphatase or histidine acid phosphatase (HAP) family whose members hydrolyze phosphate esters at acidic pH conditions.

PSPs are found in the cytosol, nucleus, and mitochondria and in association with cytoskeletal and membranous structures in most tissues, especially the brain. Some PSPs require divalent cations, such as Ca²⁺ or Mn²⁺, for activity. PSPs play important roles in glycogen metabolism, muscle contraction, protein synthesis, T cell function, neuronal activity, oocyte maturation, and hepatic metabolism (reviewed in Cohen, P. (1989) Annu. Rev. Biochem. 58:453-508). PSPs can be separated into two classes. The PPP class includes PP1, PP2A, PP2B/calcineurin, PP4, PP5, PP6, and PP7. Members of this class are composed of a homologous catalytic subunit bearing a very highly conserved signature sequence, coupled with one or more regulatory subunits (PROSITE PDOC00115). Further interactions with scaffold and anchoring molecules determine the intracellular localization of PSPs and substrate specificity. The PPM class consists of several closely related isoforms of PP2C and is evolutionarily unrelated to the PPP class.

PP1 dephosphorylates many of the proteins phosphorylated by cyclic AMP-dependent protein kinase (PKA) and is an important regulator of many cAMP-mediated hormone responses in cells. A number of isoforms have been identified, with the alpha and beta forms being produced by alternative splicing of the same gene. Both ubiquitous and tissue-specific targeting proteins for PP1 have been identified. In the brain, inhibition of PP1 activity by the dopamine and adenosine 3',5'-monophosphate-regulated phosphoprotein of 32kDa (DARPP-32) is necessary for normal dopamine response in neostriatal neurons (reviewed in Price, N.E. and M.C. Mumby (1999) Curr. Opin. Neurobiol. 9:336-342). PP1, along with PP2A, has been shown to limit motility in microvascular endothelial cells, suggesting a role for PSPs in the inhibition of angiogenesis (Gabel, S. et al. (1999)

Otolaryngol. Head Neck Surg. 121:463-468).

PP2A is the main serine/threonine phosphatase. The core PP2A enzyme consists of a single 36 kDa catalytic subunit (C) associated with a 65 kDa scaffold subunit (A), whose role is to recruit additional regulatory subunits (B). Three gene families encoding B subunits are known (PR55, 5 PR61, and PR72), each of which contain multiple isoforms, and additional families may exist (Millward, T.A et al. (1999) Trends Biosci. 24:186-191). These "B-type" subunits are cell type- and tissue-specific and determine the substrate specificity, enzymatic activity, and subcellular localization of the holoenzyme. The PR55 family is highly conserved and bears a conserved motif (PROSITE PDOC00785). PR55 increases PP2A activity toward mitogen-activated protein kinase 10 (MAPK) and MAPK kinase (MEK). PP2A dephosphorylates the MAPK active site, inhibiting the cell's entry into mitosis. Several proteins can compete with PR55 for PP2A core enzyme binding, including the CKII kinase catalytic subunit, polyomavirus middle and small T antigens, and SV40 small t antigen. Viruses may use this mechanism to commandeer PP2A and stimulate progression of the cell through the cell cycle (Pallas, D.C. et al. (1992) J. Virol. 66:886-893). Altered MAP 15 kinase expression is also implicated in a variety of disease conditions including cancer, inflammation, immune disorders, and disorders affecting growth and development. PP2A, in fact, can dephosphorylate and modulate the activities of more than 30 protein kinases in vitro, and other evidence suggests that the same is true in vivo for such kinases as PKB, PKC, the calmodulindependent kinases, ERK family MAP kinases, cyclin-dependent kinases, and the IkB kinases 20 (reviewed in Millward et al., supra). PP2A is itself a substrate for CKI and CKII kinases, and can be stimulated by polycationic macromolecules. A PP2A-like phosphatase is necessary to maintain the G1 phase destruction of mammalian cyclins A and B (Bastians, H. et al. (1999) Mol. Biol. Cell 10:3927-3941). PP2A is a major activity in the brain and is implicated in regulating neurofilament stability and normal neural function, particularly the phosphorylation of the microtubule-associated 25 protein tau. Hyperphosphorylation of tau has been proposed to lead to the neuronal degeneration seen in Alzheimer's disease (reviewed in Price and Mumby, supra).

PP2B, or calcineurin, is a Ca²⁺-activated dimeric phosphatase and is particularly abundant in the brain. It consists of catalytic and regulatory subunits, and is activated by the binding of the calcium/calmodulin complex. Calcineurin is the target of the immunosuppressant drugs

30 cyclosporine and FK506. Along with other cellular factors, these drugs interact with calcineurin and inhibit phosphatase activity. In T cells, this blocks the calcium dependent activation of the NF-AT family of transcription factors, leading to immunosuppression. This family is widely distributed, and it is likely that calcineurin regulates gene expression in other tissues as well. In neurons, calcineurin modulates functions which range from the inhibition of neurotransmitter release to desensitization of postsynaptic NMDA-receptor coupled calcium channels to long term memory

(reviewed in Price and Mumby, supra).

Other members of the PPP class have recently been identified (Cohen, P.T. (1997) Trends Biochem. Sci. 22:245-251). One of them, PP5, contains regulatory domains with tetratricopeptide repeats. It can be activated by polyunsaturated fatty acids and anionic phospholipids in vitro and appears to be involved in a number of signaling pathways, including those controlled by atrial natriuretic peptide or steroid hormones (reviewed in Andreeva, A.V. and M.A. Kutuzov (1999) Cell Signal. 11:555-562).

PP2C is a ~42kDa monomer with broad substrate specificity and is dependent on divalent cations (mainly Mn²+ or Mg²+) for its activity. PP2C proteins share a conserved N-terminal region with an invariant DGH motif, which contains an aspartate residue involved in cation binding (PROSITE PDOC00792). Targeting proteins and mechanisms regulating PP2C activity have not been identified. PP2C has been shown to inhibit the stress-responsive p38 and Jun kinase (JNK) pathways (Takekawa, M. et al. (1998) EMBO J. 17:4744-4752).

In contrast to PSPs, tyrosine-specific phosphatases (PTPs) are generally monomeric proteins

of very diverse size (from 20kDa to greater than 100kDa) and structure that function primarily in the
transduction of signals across the plasma membrane. PTPs are categorized as either soluble
phosphatases or transmembrane receptor proteins that contain a phosphatase domain. All PTPs
share a conserved catalytic domain of about 300 amino acids which contains the active site. The
active site consensus sequence includes a cysteine residue which executes a nucleophilic attack on
the phosphate moiety during catalysis (Neel, B.G. and N.K. Tonks (1997) Curr. Opin. Cell Biol.
9:193-204). Receptor PTPs are made up of an N-terminal extracellular domain of variable length, a
transmembrane region, and a cytoplasmic region that generally contains two copies of the catalytic
domain. Although only the first copy seems to have enzymatic activity, the second copy apparently
affects the substrate specificity of the first. The extracellular domains of some receptor PTPs
contain fibronectin-like repeats, immunoglobulin-like domains, MAM domains (an extracellular
motif likely to have an adhesive function), or carbonic anhydrase-like domains (PROSITE PDOC
00323). This wide variety of structural motifs accounts for the diversity in size and specificity of
PTPs.

PTPs play important roles in biological processes such as cell adhesion, lymphocyte activation, and cell proliferation. PTPs μ and κ are involved in cell-cell contacts, perhaps regulating cadherin/catenin function. A number of PTPs affect cell spreading, focal adhesions, and cell motility, most of them via the integrin/tyrosine kinase signaling pathway (reviewed in Neel and Tonks, supra). CD45 phosphatases regulate signal transduction and lymphocyte activation (Ledbetter, J.A. et al. (1988) Proc. Natl. Acad. Sci. USA 85:8628-8632). Soluble PTPs containing 35 Src-homology-2 domains have been identified (SHPs), suggesting that these molecules might

interact with receptor tyrosine kinases. SHP-1 regulates cytokine receptor signaling by controlling the Janus family PTKs in hematopoietic cells, as well as signaling by the T-cell receptor and c-Kit (reviewed in Neel and Tonks, supra). M-phase inducer phosphatase plays a key role in the induction of mitosis by dephosphorylating and activating the PTK CDC2, leading to cell division 5 (Sadhu, K. et al. (1990) Proc. Natl. Acad. Sci. USA 87:5139-5143). In addition, the genes encoding at least eight PTPs have been mapped to chromosomal regions that are translocated or rearranged in various neoplastic conditions, including lymphoma, small cell lung carcinoma, leukemia, adenocarcinoma, and neuroblastoma (reviewed in Charbonneau, H. and N.K. Tonks (1992) Annu. Rev. Cell Biol. 8:463-493). The PTP enzyme active site comprises the consensus sequence of the 10 MTM1 gene family. The MTM1 gene is responsible for X-linked recessive myotubular myopathy, a congenital muscle disorder that has been linked to Xq28 (Kioschis, P. et al., (1998) Genomics 54:256-266). Many PTKs are encoded by oncogenes, and it is well known that oncogenesis is often accompanied by increased tyrosine phosphorylation activity. It is therefore possible that PTPs may serve to prevent or reverse cell transformation and the growth of various cancers by controlling the 15 levels of tyrosine phosphorylation in cells. This is supported by studies showing that overexpression of PTP can suppress transformation in cells and that specific inhibition of PTP can enhance cell transformation (Charbonneau and Tonks, supra).

Dual specificity phosphatases (DSPs) are structurally more similar to the PTPs than the PSPs. DSPs bear an extended PTP active site motif with an additional 7 amino acid residues. DSPs are primarily associated with cell proliferation and include the cell cycle regulators cdc25A, B, and C. The phosphatases DUSP1 and DUSP2 inactivate the MAPK family members ERK (extracellular signal-regulated kinase), JNK (c-Jun N-terminal kinase), and p38 on both tyrosine and threonine residues (PROSITE PDOC 00323, supra). In the activated state, these kinases have been implicated in neuronal differentiation, proliferation, oncogenic transformation, platelet aggregation, and apoptosis. Thus, DSPs are necessary for proper regulation of these processes (Muda, M. et al. (1996) J. Biol. Chem. 271:27205-27208). The tumor suppressor PTEN is a DSP that also shows lipid phosphatase activity. It seems to negatively regulate interactions with the extracellular matrix and maintains sensitivity to apoptosis. PTEN has been implicated in the prevention of angiogenesis (Giri, D. and M. Ittmann (1999) Hum. Pathol. 30:419-424) and abnormalities in its expression are associated with numerous cancers (reviewed in Tamura, M. et al. (1999) J. Natl. Cancer Inst. 91:1820-1828).

Histidine acid phosphatase (HAP; EXPASY EC 3.1.3.2), also known as acid phosphatase, hydrolyzes a wide spectrum of substrates including alkyl, aryl, and acyl orthophosphate monoesters and phosphorylated proteins at low pH. HAPs share two regions of conserved sequences, each centered around a histidine residue which is involved in catalytic activity. Members of the HAP

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family include lysosomal acid phosphatase (LAP) and prostatic acid phosphatase (PAP), both sensitive to inhibition by L-tartrate (PROSITE PDOC00538).

LAP, an orthophosphoric monoester of the endosomal/lysosomal compartment is a housekeeping gene whose enzymatic activity has been detected in all tissues examined (Geier, C. et 5 al. (1989) Eur. J. Biochem. 183:611-616). LAP-deficient mice have progressive skeletal disorder and an increased disposition toward generalized seizures (Saftig, P. et al. (1997) J. Biol. Chem. 272:18628-18635). LAP-deficient patients were found to have the following clinical features: intermittent vomiting, hypotonia, lethargy, opisthotonos, terminal bleeding, seizures, and death in early infancy (Online Mendelian Inheritance in Man (OMIM) *200950).

PAP, a prostate epithelium-specific differentiation antigen produced by the prostate gland, has been used to diagnose and stage prostate cancer. In prostate carcinomas, the enzymatic activity of PAP was shown to be decreased compared with normal or benign prostate hypertrophy cells (Foti, A. G. et al. (1977) Cancer Res. 37: 4120-4124). Two forms of PAP have been identified, secreted and intracellular. Mature secreted PAP is detected in the seminal fluid and is active as a 15 glycosylated homodimer with a molecular weight of approximately 100-kilodalton. Intracellular PAP is found to exhibit endogenous phosphotyrosyl protein phosphatase activity and is involved in regulating prostate cell growth (Meng, T.C. and Lin, M.F. (1998) J. Biol. Chem. 34: 22096-22104).

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Synaptojanin, a polyphosphoinositide phosphatase, dephosphorylates phosphoinositides at positions 3, 4 and 5 of the inositol ring. Synaptojanin is a major presynaptic protein found at 20 clathrin-coated endocytic intermediates in nerve terminals, and binds the clathrin coat-associated protein, EPS15. This binding is mediated by the C-terminal region of synaptojanin-170, which has 3 Asp-Pro-Phe amino acid repeats. Further, this 3 residue repeat had been found to be the binding site for the EH domains of EPS15 (Haffner, C. et al. (1997) FEBS Lett. 419:175-180). Additionally, synaptojanin may potentially regulate interactions of endocytic proteins with the plasma membrane, 25 and be involved in synaptic vesicle recycling (Brodin, L. et al. (2000) Curr. Opin. Neurobiol. 10:312-320). Studies in mice with a targeted disruption in the synaptojanin 1 gene (Synj1) were shown to support coat formation of endocytic vesicles more effectively than was seen in wild-type mice, suggesting that Synj1 can act as a negative regulator of membrane-coat protein interactions. These findings provide genetic evidence for a crucial role of phosphoinositide metabolism in 30 synaptic vesicle recycling (Cremona, O. et al. (1999) Cell 99:179-188).

The discovery of new kinases and phosphatases, and the polynucleotides encoding them, satisfies a need in the art by providing new compositions which are useful in the diagnosis, prevention, and treatment of cardiovascular diseases, immune system disorders, neurological disorders, disorders affecting growth and development, lipid disorders, cell proliferative disorders, 35 and cancers, and in the assessment of the effects of exogenous compounds on the expression of

nucleic acid and amino acid sequences of kinases and phosphatases.

SUMMARY OF THE INVENTION

The invention features purified polypeptides, kinases and phosphatases, referred to

5 collectively as "KAP" and individually as "KAP-1," "KAP-2," "KAP-3," "KAP-4," "KAP-5,"

"KAP-6," "KAP-7," "KAP-8," "KAP-9," "KAP-10," "KAP-11," "KAP-12," "KAP-13," "KAP
14," "KAP-15," "KAP-16," "KAP-17," "KAP-18," "KAP-19," and "KAP-20." In one aspect, the invention provides an isolated polypeptide selected from the group consisting of a) a polypeptide comprising an amino acid sequence selected from the group consisting of SEQ ID NO:1-20, b) a

10 polypeptide comprising a naturally occurring amino acid sequence at least 90% identical to an amino acid sequence selected from the group consisting of SEQ ID NO:1-20, c) a biologically active fragment of a polypeptide having an amino acid sequence selected from the group consisting of SEQ ID NO:1-20, and d) an immunogenic fragment of a polypeptide having an amino acid sequence selected from the group consisting of SEQ ID NO:1-20. In one alternative, the invention provides

15 an isolated polypeptide comprising the amino acid sequence of SEQ ID NO:1-20.

The invention further provides an isolated polynucleotide encoding a polypeptide selected from the group consisting of a) a polypeptide comprising an amino acid sequence selected from the group consisting of SEQ ID NO:1-20, b) a polypeptide comprising a naturally occurring amino acid sequence at least 90% identical to an amino acid sequence selected from the group consisting of 20 SEQ ID NO:1-20, c) a biologically active fragment of a polypeptide having an amino acid sequence selected from the group consisting of SEQ ID NO:1-20, and d) an immunogenic fragment of a polypeptide having an amino acid sequence selected from the group consisting of SEQ ID NO:1-20. In one alternative, the polynucleotide encodes a polypeptide selected from the group consisting of SEQ ID NO:1-20. In another alternative, the polynucleotide is selected from the group consisting of SEQ ID NO:1-40.

Additionally, the invention provides a recombinant polynucleotide comprising a promoter sequence operably linked to a polynucleotide encoding a polypeptide selected from the group consisting of a) a polypeptide comprising an amino acid sequence selected from the group consisting of SEQ ID NO:1-20, b) a polypeptide comprising a naturally occurring amino acid 30 sequence at least 90% identical to an amino acid sequence selected from the group consisting of SEQ ID NO:1-20, c) a biologically active fragment of a polypeptide having an amino acid sequence selected from the group consisting of SEQ ID NO:1-20, and d) an immunogenic fragment of a polypeptide having an amino acid sequence selected from the group consisting of SEQ ID NO:1-20. In one alternative, the invention provides a cell transformed with the recombinant polynucleotide.

polynucleotide.

The invention also provides a method for producing a polypeptide selected from the group consisting of a) a polypeptide comprising an amino acid sequence selected from the group consisting of SEQ ID NO:1-20, b) a polypeptide comprising a naturally occurring amino acid sequence at least 90% identical to an amino acid sequence selected from the group consisting of SEQ ID NO:1-20, c) a biologically active fragment of a polypeptide having an amino acid sequence selected from the group consisting of SEQ ID NO:1-20, and d) an immunogenic fragment of a polypeptide having an amino acid sequence selected from the group consisting of SEQ ID NO:1-20. The method comprises a) culturing a cell under conditions suitable for expression of the polypeptide, wherein said cell is transformed with a recombinant polynucleotide comprising a promoter sequence operably linked to a polynucleotide encoding the polypeptide, and b) recovering the polypeptide so expressed.

Additionally, the invention provides an isolated antibody which specifically binds to a polypeptide selected from the group consisting of a) a polypeptide comprising an amino acid sequence selected from the group consisting of SEQ ID NO:1-20, b) a polypeptide comprising a naturally occurring amino acid sequence at least 90% identical to an amino acid sequence selected from the group consisting of SEQ ID NO:1-20, c) a biologically active fragment of a polypeptide having an amino acid sequence selected from the group consisting of SEQ ID NO:1-20, and d) an immunogenic fragment of a polypeptide having an amino acid sequence selected from the group 20 consisting of SEQ ID NO:1-20.

The invention further provides an isolated polynucleotide selected from the group consisting of a) a polynucleotide comprising a polynucleotide sequence selected from the group consisting of SEQ ID NO:21-40, b) a polynucleotide comprising a naturally occurring polynucleotide sequence at least 90% identical to a polynucleotide sequence selected from the group consisting of SEQ ID NO:21-40, c) a polynucleotide complementary to the polynucleotide of a), d) a polynucleotide complementary to the polynucleotide of b), and e) an RNA equivalent of a)-d). In one alternative, the polynucleotide comprises at least 60 contiguous nucleotides.

Additionally, the invention provides a method for detecting a target polynucleotide in a sample, said target polynucleotide having a sequence of a polynucleotide selected from the group 30 consisting of a) a polynucleotide comprising a polynucleotide sequence selected from the group consisting of SEQ ID NO:21-40, b) a polynucleotide comprising a naturally occurring polynucleotide sequence at least 90% identical to a polynucleotide sequence selected from the group consisting of SEQ ID NO:21-40, c) a polynucleotide complementary to the polynucleotide of a), d) a polynucleotide complementary to the polynucleotide of b), and e) an RNA equivalent of a)-35 d). The method comprises a) hybridizing the sample with a probe comprising at least 20 contiguous

nucleotides comprising a sequence complementary to said target polynucleotide in the sample, and which probe specifically hybridizes to said target polynucleotide, under conditions whereby a hybridization complex is formed between said probe and said target polynucleotide or fragments thereof, and b) detecting the presence or absence of said hybridization complex, and optionally, if 5 present, the amount thereof. In one alternative, the probe comprises at least 60 contiguous nucleotides.

The invention further provides a method for detecting a target polynucleotide in a sample, said target polynucleotide having a sequence of a polynucleotide selected from the group consisting of a) a polynucleotide comprising a polynucleotide sequence selected from the group consisting of SEQ ID NO:21-40, b) a polynucleotide comprising a naturally occurring polynucleotide sequence at least 90% identical to a polynucleotide sequence selected from the group consisting of SEQ ID NO:21-40, c) a polynucleotide complementary to the polynucleotide of a), d) a polynucleotide complementary to the polynucleotide of b), and e) an RNA equivalent of a)-d). The method comprises a) amplifying said target polynucleotide or fragment thereof using polymerase chain 15 reaction amplification, and b) detecting the presence or absence of said amplified target polynucleotide or fragment thereof, and, optionally, if present, the amount thereof.

The invention further provides a composition comprising an effective amount of a polypeptide selected from the group consisting of a) a polypeptide comprising an amino acid sequence selected from the group consisting of SEQ ID NO:1-20, b) a polypeptide comprising a 20 naturally occurring amino acid sequence at least 90% identical to an amino acid sequence selected from the group consisting of SEQ ID NO:1-20, c) a biologically active fragment of a polypeptide having an amino acid sequence selected from the group consisting of SEQ ID NO:1-20, and d) an immunogenic fragment of a polypeptide having an amino acid sequence selected from the group consisting of SEQ ID NO:1-20, and a pharmaceutically acceptable excipient. In one embodiment, 25 the composition comprises an amino acid sequence selected from the group consisting of SEQ ID NO:1-20. The invention additionally provides a method of treating a disease or condition associated with decreased expression of functional KAP, comprising administering to a patient in need of such treatment the composition.

The invention also provides a method for screening a compound for effectiveness as an 30 agonist of a polypeptide selected from the group consisting of a) a polypeptide comprising an amino acid sequence selected from the group consisting of SEQ ID NO:1-20, b) a polypeptide comprising a naturally occurring amino acid-sequence at least 90% identical to an amino acid-sequence selected from the group consisting of SEQ ID NO:1-20, c) a biologically active fragment of a polypeptide having an amino acid sequence selected from the group consisting of SEQ ID NO:1-20, and d) an 35 immunogenic fragment of a polypeptide having an amino acid sequence selected from the group

consisting of SEQ ID NO:1-20. The method comprises a) exposing a sample comprising the polypeptide to a compound, and b) detecting agonist activity in the sample. In one alternative, the invention provides a composition comprising an agonist compound identified by the method and a pharmaceutically acceptable excipient. In another alternative, the invention provides a method of treating a disease or condition associated with decreased expression of functional KAP, comprising administering to a patient in need of such treatment the composition.

Additionally, the invention provides a method for screening a compound for effectiveness as an antagonist of a polypeptide selected from the group consisting of a) a polypeptide comprising an amino acid sequence selected from the group consisting of SEQ ID NO:1-20, b) a polypeptide comprising a naturally occurring amino acid sequence at least 90% identical to an amino acid sequence selected from the group consisting of SEQ ID NO:1-20, c) a biologically active fragment of a polypeptide having an amino acid sequence selected from the group consisting of SEQ ID NO:1-20, and d) an immunogenic fragment of a polypeptide having an amino acid sequence selected from the group consisting of SEQ ID NO:1-20. The method comprises a) exposing a sample comprising the polypeptide to a compound, and b) detecting antagonist activity in the sample. In one alternative, the invention provides a composition comprising an antagonist compound identified by the method and a pharmaceutically acceptable excipient. In another alternative, the invention provides a method of treating a disease or condition associated with overexpression of functional KAP, comprising administering to a patient in need of such treatment the composition.

20 The invention further provides a method of screening for a compound that specifically binds to a polypeptide selected from the group consisting of a) a polypeptide comprising an amino acid sequence selected from the group consisting of SEQ ID NO:1-20, b) a polypeptide comprising a naturally occurring amino acid sequence at least 90% identical to an amino acid sequence selected from the group consisting of SEQ ID NO:1-20, c) a biologically active fragment of a polypeptide 25 having an amino acid sequence selected from the group consisting of SEQ ID NO:1-20, and d) an immunogenic fragment of a polypeptide having an amino acid sequence selected from the group consisting of SEQ ID NO:1-20. The method comprises a) combining the polypeptide with at least one test compound under suitable conditions, and b) detecting binding of the polypeptide to the test compound, thereby identifying a compound that specifically binds to the polypeptide.

The invention further provides a method of screening for a compound that modulates the activity of a polypeptide selected from the group consisting of a) a polypeptide comprising an amino acid sequence selected from the group consisting of SEQ ID NO:1-20, b) a polypeptide comprising a naturally occurring amino acid sequence at least 90% identical to an amino acid sequence selected from the group consisting of SEQ ID NO:1-20, c) a biologically active fragment of a polypeptide so having an amino acid sequence selected from the group consisting of SEQ ID NO:1-20, and d) an

immunogenic fragment of a polypeptide having an amino acid sequence selected from the group consisting of SEQ ID NO:1-20. The method comprises a) combining the polypeptide with at least one test compound under conditions permissive for the activity of the polypeptide, b) assessing the activity of the polypeptide in the presence of the test compound, and c) comparing the activity of the polypeptide in the polypeptide in the absence of the test compound with the activity of the polypeptide in the presence of the test compound, wherein a change in the activity of the polypeptide in the presence of the test compound is indicative of a compound that modulates the activity of the polypeptide.

The invention further provides a method for screening a compound for effectiveness in altering expression of a target polynucleotide, wherein said target polynucleotide comprises a 10 polynucleotide sequence selected from the group consisting of SEQ ID NO:21-40, the method comprising a) exposing a sample comprising the target polynucleotide to a compound, b) detecting altered expression of the target polynucleotide, and c) comparing the expression of the target polynucleotide in the presence of varying amounts of the compound and in the absence of the compound.

15 The invention further provides a method for assessing toxicity of a test compound, said method comprising a) treating a biological sample containing nucleic acids with the test compound; b) hybridizing the nucleic acids of the treated biological sample with a probe comprising at least 20 contiguous nucleotides of a polynucleotide selected from the group consisting of i) a polynucleotide comprising a polynucleotide sequence selected from the group consisting of SEO ID NO:21-40, ii) 20 a polynucleotide comprising a naturally occurring polynucleotide sequence at least 90% identical to a polynucleotide sequence selected from the group consisting of SEO ID NO:21-40, iii) a polynucleotide having a sequence complementary to i), iv) a polynucleotide complementary to the polynucleotide of ii), and v) an RNA equivalent of i)-iv). Hybridization occurs under conditions whereby a specific hybridization complex is formed between said probe and a target polynucleotide 25 in the biological sample, said target polynucleotide selected from the group consisting of i) a polynucleotide comprising a polynucleotide sequence selected from the group consisting of SEO ID NO:21-40, ii) a polynucleotide comprising a naturally occurring polynucleotide sequence at least 90% identical to a polynucleotide sequence selected from the group consisting of SEQ ID NO:21-40, iii) a polynucleotide complementary to the polynucleotide of i), iv) a polynucleotide 30 complementary to the polynucleotide of ii), and v) an RNA equivalent of i)-iv). Alternatively, the target polynucleotide comprises a fragment of a polynucleotide sequence selected from the group consisting of i)-v) above; c) quantifying the amount of hybridization complex; and d) comparing the amount of hybridization complex in the treated biological sample with the amount of hybridization complex in an untreated biological sample, wherein a difference in the amount of hybridization

35 complex in the treated biological sample is indicative of toxicity of the test compound.

BRIEF DESCRIPTION OF THE TABLES

Table 1 summarizes the nomenclature for the full length polynucleotide and polypeptide sequences of the present invention.

Table 2 shows the GenBank identification number and annotation of the nearest GenBank homolog for polypeptides of the invention. The probability scores for the matches between each polypeptide and its homolog(s) are also shown.

Table 3 shows structural features of polypeptide sequences of the invention, including predicted motifs and domains, along with the methods, algorithms, and searchable databases used 10 for analysis of the polypeptides.

Table 4 lists the cDNA and/or genomic DNA fragments which were used to assemble polynucleotide sequences of the invention, along with selected fragments of the polynucleotide sequences.

Table 5 shows the representative cDNA library for polynucleotides of the invention.

Table 6 provides an appendix which describes the tissues and vectors used for construction of the cDNA libraries shown in Table 5.

Table 7 shows the tools, programs, and algorithms used to analyze the polynucleotides and polypeptides of the invention, along with applicable descriptions, references, and threshold parameters.

20

DESCRIPTION OF THE INVENTION

Before the present proteins, nucleotide sequences, and methods are described, it is understood that this invention is not limited to the particular machines, materials and methods described, as these may vary. It is also to be understood that the terminology used herein is for the purpose of describing particular embodiments only, and is not intended to limit the scope of the present invention which will be limited only by the appended claims.

It must be noted that as used herein and in the appended claims, the singular forms "a," "an," and "the" include plural reference unless the context clearly dictates otherwise. Thus, for example, a reference to "a host cell" includes a plurality of such host cells, and a reference to "an 30 antibody" is a reference to one or more antibodies and equivalents thereof known to those skilled in the art, and so forth.

Unless defined otherwise, all technical and scientific terms used herein have the same meanings as commonly understood by one of ordinary skill in the art to which this invention belongs. Although any machines, materials, and methods similar or equivalent to those described 35 herein can be used to practice or test the present invention, the preferred machines, materials and

methods are now described. All publications mentioned herein are cited for the purpose of describing and disclosing the cell lines, protocols, reagents and vectors which are reported in the publications and which might be used in connection with the invention. Nothing herein is to be construed as an admission that the invention is not entitled to antedate such disclosure by virtue of 5 prior invention.

DEFINITIONS

BNS/DOCID-

"KAP" refers to the amino acid sequences of substantially purified KAP obtained from any species, particularly a mammalian species, including bovine, ovine, porcine, murine, equine, and human, and from any source, whether natural, synthetic, semi-synthetic, or recombinant.

The term "agonist" refers to a molecule which intensifies or mimics the biological activity of KAP. Agonists may include proteins, nucleic acids, carbohydrates, small molecules, or any other compound or composition which modulates the activity of KAP either by directly interacting with KAP or by acting on components of the biological pathway in which KAP participates.

An "allelic variant" is an alternative form of the gene encoding KAP. Allelic variants may 15 result from at least one mutation in the nucleic acid sequence and may result in altered mRNAs or in polypeptides whose structure or function may or may not be altered. A gene may have none, one, or many allelic variants of its naturally occurring form. Common mutational changes which give rise to allelic variants are generally ascribed to natural deletions, additions, or substitutions of nucleotides. Each of these types of changes may occur alone, or in combination with the others, one 20 or more times in a given sequence.

"Altered" nucleic acid sequences encoding KAP include those sequences with deletions, insertions, or substitutions of different nucleotides, resulting in a polypeptide the same as KAP or a polypeptide with at least one functional characteristic of KAP. Included within this definition are polymorphisms which may or may not be readily detectable using a particular oligonucleotide probe of the polynucleotide encoding KAP, and improper or unexpected hybridization to allelic variants, with a locus other than the normal chromosomal locus for the polynucleotide sequence encoding KAP. The encoded protein may also be "altered," and may contain deletions, insertions, or substitutions of amino acid residues which produce a silent change and result in a functionally equivalent KAP. Deliberate amino acid substitutions may be made on the basis of similarity in 30 polarity, charge, solubility, hydrophobicity, hydrophilicity, and/or the amphipathic nature of the residues, as long as the biological or immunological activity of KAP is retained. For example, negatively charged amino acids may include aspartic acid and glutamic acid, and positively charged amino acids may include lysine and arginine. Amino acids with uncharged polar side chains having similar hydrophilicity values may include: asparagine and glutamine; and serine and threonine.

isoleucine, and valine; glycine and alanine; and phenylalanine and tyrosine.

The terms "amino acid" and "amino acid sequence" refer to an oligopeptide, peptide, polypeptide, or protein sequence, or a fragment of any of these, and to naturally occurring or synthetic molecules. Where "amino acid sequence" is recited to refer to a sequence of a naturally occurring protein molecule, "amino acid sequence" and like terms are not meant to limit the amino acid sequence to the complete native amino acid sequence associated with the recited protein molecule.

"Amplification" relates to the production of additional copies of a nucleic acid sequence.

Amplification is generally carried out using polymerase chain reaction (PCR) technologies well

10 known in the art.

The term "antagonist" refers to a molecule which inhibits or attenuates the biological activity of KAP. Antagonists may include proteins such as antibodies, nucleic acids, carbohydrates, small molecules, or any other compound or composition which modulates the activity of KAP either by directly interacting with KAP or by acting on components of the biological pathway in which 15 KAP participates.

The term "antibody" refers to intact immunoglobulin molecules as well as to fragments thereof, such as Fab, F(ab')₂, and Fv fragments, which are capable of binding an epitopic determinant. Antibodies that bind KAP polypeptides can be prepared using intact polypeptides or using fragments containing small peptides of interest as the immunizing antigen. The polypeptide or oligopeptide used to immunize an animal (e.g., a mouse, a rat, or a rabbit) can be derived from the translation of RNA, or synthesized chemically, and can be conjugated to a carrier protein if desired. Commonly used carriers that are chemically coupled to peptides include bovine serum albumin, thyroglobulin, and keyhole limpet hemocyanin (KLH). The coupled peptide is then used to immunize the animal.

The term "antigenic determinant" refers to that region of a molecule (i.e., an epitope) that makes contact with a particular antibody. When a protein or a fragment of a protein is used to immunize a host animal, numerous regions of the protein may induce the production of antibodies which bind specifically to antigenic determinants (particular regions or three-dimensional structures on the protein). An antigenic determinant may compete with the intact antigen (i.e., the immunogen 30 used to elicit the immune response) for binding to an antibody.

The term "aptamer" refers to a nucleic acid or oligonucleotide molecule that binds to a specific molecular target. Aptamers are derived from an <u>in vitro</u> evolutionary process (e.g., SELEX (Systematic Evolution of Ligands by EXponential Enrichment), described in U.S. Patent No. 5,270,163), which selects for target-specific aptamer sequences from large combinatorial libraries.

35 Aptamer compositions may be double-stranded or single-stranded, and may include

deoxyribonucleotides, ribonucleotides, nucleotide derivatives, or other nucleotide-like molecules. The nucleotide components of an aptamer may have modified sugar groups (e.g., the 2'-OH group of a ribonucleotide may be replaced by 2'-F or 2'-NH₂), which may improve a desired property, e.g., resistance to nucleases or longer lifetime in blood. Aptamers may be conjugated to other molecules, 5 e.g., a high molecular weight carrier to slow clearance of the aptamer from the circulatory system. Aptamers may be specifically cross-linked to their cognate ligands, e.g., by photo-activation of a cross-linker. (See, e.g., Brody, E.N. and L. Gold (2000) J. Biotechnol. 74:5-13.)

The term "intramer" refers to an aptamer which is expressed in vivo. For example, a vaccinia virus-based RNA expression system has been used to express specific RNA aptamers at 10 high levels in the cytoplasm of leukocytes (Blind, M. et al. (1999) Proc. Natl Acad. Sci. USA 96:3606-3610).

The term "spiegelmer" refers to an aptamer which includes L-DNA, L-RNA, or other left-handed nucleotide derivatives or nucleotide-like molecules. Aptamers containing left-handed nucleotides are resistant to degradation by naturally occurring enzymes, which normally act on 15 substrates containing right-handed nucleotides.

The term "antisense" refers to any composition capable of base-pairing with the "sense" (coding) strand of a specific nucleic acid sequence. Antisense compositions may include DNA; RNA; peptide nucleic acid (PNA); oligonucleotides having modified backbone linkages such as phosphorothioates, methylphosphonates, or benzylphosphonates; oligonucleotides having modified 20 sugar groups such as 2'-methoxyethyl sugars or 2'-methoxyethoxy sugars; or oligonucleotides having modified bases such as 5-methyl cytosine, 2'-deoxyuracil, or 7-deaza-2'-deoxyguanosine. Antisense molecules may be produced by any method including chemical synthesis or transcription. Once introduced into a cell, the complementary antisense molecule base-pairs with a naturally occurring nucleic acid sequence produced by the cell to form duplexes which block either transcription or 25 translation. The designation "negative" or "minus" can refer to the antisense strand, and the designation "positive" or "plus" can refer to the sense strand of a reference DNA molecule.

The term "biologically active" refers to a protein having structural, regulatory, or biochemical functions of a naturally occurring molecule. Likewise, "immunologically active" or "immunogenic" refers to the capability of the natural, recombinant, or synthetic KAP, or of any 30 oligopeptide thereof, to induce a specific immune response in appropriate animals or cells and to bind with specific antibodies.

"Complementary" describes the relationship between two single-stranded nucleic acid sequences that anneal by base-pairing. For example, 5'-AGT-3' pairs with its complement, 3'-TCA-5'.

A "composition comprising a given polynucleotide sequence" and a "composition

comprising a given amino acid sequence" refer broadly to any composition containing the given polynucleotide or amino acid sequence. The composition may comprise a dry formulation or an aqueous solution. Compositions comprising polynucleotide sequences encoding KAP or fragments of KAP may be employed as hybridization probes. The probes may be stored in freeze-dried form and may be associated with a stabilizing agent such as a carbohydrate. In hybridizations, the probe may be deployed in an aqueous solution containing salts (e.g., NaCl), detergents (e.g., sodium dodecyl sulfate; SDS), and other components (e.g., Denhardt's solution, dry milk, salmon sperm DNA, etc.).

"Consensus sequence" refers to a nucleic acid sequence which has been subjected to 10 repeated DNA sequence analysis to resolve uncalled bases, extended using the XL-PCR kit (Applied Biosystems, Foster City CA) in the 5' and/or the 3' direction, and resequenced, or which has been assembled from one or more overlapping cDNA, EST, or genomic DNA fragments using a computer program for fragment assembly, such as the GELVIEW fragment assembly system (GCG, Madison WI) or Phrap (University of Washington, Seattle WA). Some sequences have been both 15 extended and assembled to produce the consensus sequence.

"Conservative amino acid substitutions" are those substitutions that are predicted to least interfere with the properties of the original protein, i.e., the structure and especially the function of the protein is conserved and not significantly changed by such substitutions. The table below shows amino acids which may be substituted for an original amino acid in a protein and which are 20 regarded as conservative amino acid substitutions.

20 108		
	Original Residue	Conservative Substitution
-	Ala	Gly, Ser
	Arg	His, Lys
	Asn	Asp, Gln, His
25	Asp	Asn, Glu
23	Cys	Ala, Ser
	Gln	Asn, Glu, His
	Glu	Asp, Gln, His
	Gly	Ala
30	His	Asn, Arg, Gln, Glu
	Пе	Leu, Val
	Leu	Ile, Val
	Lys	Arg, Gln, Glu
	Met	Leu, Ile
35	Phe	His, Met, Leu, Trp, Tyr
	Ser	Cys, Thr
	Thr	Ser, Val
	Trp	Phe, Tyr
	Tyr	His, Phe, Trp
40	Val	Ile, Leu, Thr
. •		

Conservative amino acid substitutions generally maintain (a) the structure of the

polypeptide backbone in the area of the substitution, for example, as a beta sheet or alpha helical conformation, (b) the charge or hydrophobicity of the molecule at the site of the substitution, and/or (c) the bulk of the side chain.

A "deletion" refers to a change in the amino acid or nucleotide sequence that results in the 5 absence of one or more amino acid residues or nucleotides.

The term "derivative" refers to a chemically modified polynucleotide or polypeptide.

Chemical modifications of a polynucleotide can include, for example, replacement of hydrogen by an alkyl, acyl, hydroxyl, or amino group. A derivative polynucleotide encodes a polypeptide which retains at least one biological or immunological function of the natural molecule. A derivative 10 polypeptide is one modified by glycosylation, pegylation, or any similar process that retains at least one biological or immunological function of the polypeptide from which it was derived.

A "detectable label" refers to a reporter molecule or enzyme that is capable of generating a measurable signal and is covalently or noncovalently joined to a polynucleotide or polypeptide.

"Differential expression" refers to increased or upregulated; or decreased, downregulated,
15 or absent gene or protein expression, determined by comparing at least two different samples. Such
comparisons may be carried out between, for example, a treated and an untreated sample, or a
diseased and a normal sample.

"Exon shuffling" refers to the recombination of different coding regions (exons). Since an exon may represent a structural or functional domain of the encoded protein, new proteins may be 20 assembled through the novel reassortment of stable substructures, thus allowing acceleration of the evolution of new protein functions.

A "fragment" is a unique portion of KAP or the polynucleotide encoding KAP which is identical in sequence to but shorter in length than the parent sequence. A fragment may comprise up to the entire length of the defined sequence, minus one nucleotide/amino acid residue. For 25 example, a fragment may comprise from 5 to 1000 contiguous nucleotides or amino acid residues. A fragment used as a probe, primer, antigen, therapeutic molecule, or for other purposes, may be at least 5, 10, 15, 16, 20, 25, 30, 40, 50, 60, 75, 100, 150, 250 or at least 500 contiguous nucleotides or amino acid residues in length. Fragments may be preferentially selected from certain regions of a molecule. For example, a polypeptide fragment may comprise a certain length of contiguous amino 30 acids selected from the first 250 or 500 amino acids (or first 25% or 50%) of a polypeptide as shown in a certain defined sequence. Clearly these lengths are exemplary, and any length that is supported by the specification, including the Sequence Listing, tables, and figures, may be encompassed by the present embodiments.

A fragment of SEQ ID NO:21-40 comprises a region of unique polynucleotide sequence 35 that specifically identifies SEQ ID NO:21-40, for example, as distinct from any other sequence in

the genome from which the fragment was obtained. A fragment of SEQ ID NO:21-40 is useful, for example, in hybridization and amplification technologies and in analogous methods that distinguish SEQ ID NO:21-40 from related polynucleotide sequences. The precise length of a fragment of SEQ ID NO:21-40 and the region of SEQ ID NO:21-40 to which the fragment corresponds are routinely 5 determinable by one of ordinary skill in the art based on the intended purpose for the fragment.

A fragment of SEQ ID NO:1-20 is encoded by a fragment of SEQ ID NO:21-40. A fragment of SEQ ID NO:1-20 comprises a region of unique amino acid sequence that specifically identifies SEQ ID NO:1-20. For example, a fragment of SEQ ID NO:1-20 is useful as an immunogenic peptide for the development of antibodies that specifically recognize SEQ ID NO:1-10 20. The precise length of a fragment of SEQ ID NO:1-20 and the region of SEQ ID NO:1-20 to which the fragment corresponds are routinely determinable by one of ordinary skill in the art based on the intended purpose for the fragment.

A "full length" polynucleotide sequence is one containing at least a translation initiation codon (e.g., methionine) followed by an open reading frame and a translation termination codon. A 15 "full length" polynucleotide sequence encodes a "full length" polypeptide sequence.

"Homology" refers to sequence similarity or, interchangeably, sequence identity, between two or more polynucleotide sequences or two or more polypeptide sequences.

The terms "percent identity" and "% identity," as applied to polynucleotide sequences, refer to the percentage of residue matches between at least two polynucleotide sequences aligned using a 20 standardized algorithm. Such an algorithm may insert, in a standardized and reproducible way, gaps in the sequences being compared in order to optimize alignment between two sequences, and therefore achieve a more meaningful comparison of the two sequences.

Percent identity between polynucleotide sequences may be determined using the default parameters of the CLUSTAL V algorithm as incorporated into the MEGALIGN version 3.12e sequence alignment program. This program is part of the LASERGENE software package, a suite of molecular biological analysis programs (DNASTAR, Madison WI). CLUSTAL V is described in Higgins, D.G. and P.M. Sharp (1989) CABIOS 5:151-153 and in Higgins, D.G. et al. (1992) CABIOS 8:189-191. For pairwise alignments of polynucleotide sequences, the default parameters are set as follows: Ktuple=2, gap penalty=5, window=4, and "diagonals saved"=4. The "weighted" 30 residue weight table is selected as the default. Percent identity is reported by CLUSTAL V as the "percent similarity" between aligned polynucleotide sequences.

Alternatively, a suite of commonly used and freely available sequence comparison algorithms is provided by the National Center for Biotechnology Information (NCBI) Basic Local Alignment Search Tool (BLAST) (Altschul, S.F. et al. (1990) J. Mol. Biol. 215:403-410), which is available from several sources, including the NCBI, Bethesda, MD, and on the Internet at

http://www.ncbi.nlm.nih.gov/BLAST/. The BLAST software suite includes various sequence analysis programs including "blastn," that is used to align a known polynucleotide sequence with other polynucleotide sequences from a variety of databases. Also available is a tool called "BLAST 2 Sequences" that is used for direct pairwise comparison of two nucleotide sequences. "BLAST 2

5 Sequences" can be accessed and used interactively at http://www.ncbi.nlm.nih.gov/gorf/bl2.html.

The "BLAST 2 Sequences" tool can be used for both blastn and blastp (discussed below). BLAST programs are commonly used with gap and other parameters set to default settings. For example, to compare two nucleotide sequences, one may use blastn with the "BLAST 2 Sequences" tool

Version 2.0.12 (April-21-2000) set at default parameters. Such default parameters may be, for 10 example:

Matrix: BLOSUM62

Reward for match: 1

Penalty for mismatch: -2

Open Gap: 5 and Extension Gap: 2 penalties

15 *Gap x drop-off: 50*

Expect: 10 Word Size: 11

Filter: on

Percent identity may be measured over the length of an entire defined sequence, for 20 example, as defined by a particular SEQ ID number, or may be measured over a shorter length, for example, over the length of a fragment taken from a larger, defined sequence, for instance, a fragment of at least 20, at least 30, at least 40, at least 50, at least 70, at least 100, or at least 200 contiguous nucleotides. Such lengths are exemplary only, and it is understood that any fragment length supported by the sequences shown herein, in the tables, figures, or Sequence Listing, may be 25 used to describe a length over which percentage identity may be measured.

Nucleic acid sequences that do not show a high degree of identity may nevertheless encode similar amino acid sequences due to the degeneracy of the genetic code. It is understood that changes in a nucleic acid sequence can be made using this degeneracy to produce multiple nucleic acid sequences that all encode substantially the same protein.

The phrases "percent identity" and "% identity," as applied to polypeptide sequences, refer to the percentage of residue matches between at least two polypeptide sequences aligned using a standardized algorithm. Methods of polypeptide sequence alignment are well-known. Some alignment methods take into account conservative amino acid substitutions. Such conservative substitutions, explained in more detail above, generally preserve the charge and hydrophobicity at 35 the site of substitution, thus preserving the structure (and therefore function) of the polypeptide.

Percent identity between polypeptide sequences may be determined using the default parameters of the CLUSTAL V algorithm as incorporated into the MEGALIGN version 3.12e sequence alignment program (described and referenced above). For pairwise alignments of polypeptide sequences using CLUSTAL V, the default parameters are set as follows: Ktuple=1, gap 5 penalty=3, window=5, and "diagonals saved"=5. The PAM250 matrix is selected as the default residue weight table. As with polynucleotide alignments, the percent identity is reported by CLUSTAL V as the "percent similarity" between aligned polypeptide sequence pairs.

Alternatively the NCBI BLAST software suite may be used. For example, for a pairwise comparison of two polypeptide sequences, one may use the "BLAST 2 Sequences" tool Version 10 2.0.12 (April-21-2000) with blastp set at default parameters. Such default parameters may be, for example:

Matrix: BLOSUM62

Open Gap: 11 and Extension Gap: 1 penalties

Gap x drop-off: 50

15 Expect: 10

Word Size: 3

Filter: on

Percent identity may be measured over the length of an entire defined polypeptide sequence, for example, as defined by a particular SEQ ID number, or may be measured over a shorter length, 20 for example, over the length of a fragment taken from a larger, defined polypeptide sequence, for instance, a fragment of at least 15, at least 20, at least 30, at least 40, at least 50, at least 70 or at least 150 contiguous residues. Such lengths are exemplary only, and it is understood that any fragment length supported by the sequences shown herein, in the tables, figures or Sequence Listing, may be used to describe a length over which percentage identity may be measured.

25 "Human artificial chromosomes" (HACs) are linear microchromosomes which may contain DNA sequences of about 6 kb to 10 Mb in size and which contain all of the elements required for chromosome replication, segregation and maintenance.

The term "humanized antibody" refers to an antibody molecule in which the amino acid sequence in the non-antigen binding regions has been altered so that the antibody more closely 30 resembles a human antibody, and still retains its original binding ability.

"Hybridization" refers to the process by which a polynucleotide strand anneals with a complementary strand through base pairing under defined hybridization conditions. Specific hybridization is an indication that two nucleic acid sequences share a high degree of complementarity. Specific hybridization complexes form under permissive annealing conditions and remain hybridized after the "washing" step(s). The washing step(s) is particularly important in

determining the stringency of the hybridization process, with more stringent conditions allowing less non-specific binding, i.e., binding between pairs of nucleic acid strands that are not perfectly matched. Permissive conditions for annealing of nucleic acid sequences are routinely determinable by one of ordinary skill in the art and may be consistent among hybridization experiments, whereas 5 wash conditions may be varied among experiments to achieve the desired stringency, and therefore hybridization specificity. Permissive annealing conditions occur, for example, at 68°C in the presence of about 6 x SSC, about 1% (w/v) SDS, and about 100 μ g/ml sheared, denatured salmon sperm DNA.

Generally, stringency of hybridization is expressed, in part, with reference to the 10 temperature under which the wash step is carried out. Such wash temperatures are typically selected to be about 5°C to 20°C lower than the thermal melting point (T_m) for the specific sequence at a defined ionic strength and pH. The T_m is the temperature (under defined ionic strength and pH) at which 50% of the target sequence hybridizes to a perfectly matched probe. An equation for calculating T_m and conditions for nucleic acid hybridization are well known and can be found in 15 Sambrook, J. et al. (1989) Molecular Cloning: A Laboratory Manual, 2nd ed., vol. 1-3, Cold Spring Harbor Press, Plainview NY; specifically see volume 2, chapter 9.

High stringency conditions for hybridization between polynucleotides of the present invention include wash conditions of 68°C in the presence of about 0.2 x SSC and about 0.1% SDS, for 1 hour. Alternatively, temperatures of about 65°C, 60°C, 55°C, or 42°C may be used. SSC 20 concentration may be varied from about 0.1 to 2 x SSC, with SDS being present at about 0.1%. Typically, blocking reagents are used to block non-specific hybridization. Such blocking reagents include, for instance, sheared and denatured salmon sperm DNA at about 100-200 μg/ml. Organic solvent, such as formamide at a concentration of about 35-50% v/v, may also be used under particular circumstances, such as for RNA:DNA hybridizations. Useful variations on these wash 25 conditions will be readily apparent to those of ordinary skill in the art. Hybridization, particularly under high stringency conditions, may be suggestive of evolutionary similarity between the nucleotides. Such similarity is strongly indicative of a similar role for the nucleotides and their encoded polypeptides.

The term "hybridization complex" refers to a complex formed between two nucleic acid 30 sequences by virtue of the formation of hydrogen bonds between complementary bases. A hybridization complex may be formed in solution (e.g., C_0 t or R_0 t analysis) or formed between one nucleic acid sequence present in solution and another nucleic acid sequence immobilized on a solid support (e.g., paper, membranes, filters, chips, pins or glass slides, or any other appropriate substrate to which cells or their nucleic acids have been fixed).

35 The words "insertion" and "addition" refer to changes in an amino acid or nucleotide

sequence resulting in the addition of one or more amino acid residues or nucleotides, respectively.

"Immune response" can refer to conditions associated with inflammation, trauma, immune disorders, or infectious or genetic disease, etc. These conditions can be characterized by expression of various factors, e.g., cytokines, chemokines, and other signaling molecules, which may affect 5 cellular and systemic defense systems.

An "immunogenic fragment" is a polypeptide or oligopeptide fragment of KAP which is capable of eliciting an immune response when introduced into a living organism, for example, a mammal. The term "immunogenic fragment" also includes any polypeptide or oligopeptide fragment of KAP which is useful in any of the antibody production methods disclosed herein or 10 known in the art.

The term "microarray" refers to an arrangement of a plurality of polynucleotides, polypeptides, or other chemical compounds on a substrate.

The terms "element" and "array element" refer to a polynucleotide, polypeptide, or other chemical compound having a unique and defined position on a microarray.

The term "modulate" refers to a change in the activity of KAP. For example, modulation may cause an increase or a decrease in protein activity, binding characteristics, or any other biological, functional, or immunological properties of KAP.

The phrases "nucleic acid" and "nucleic acid sequence" refer to a nucleotide, oligonucleotide, polynucleotide, or any fragment thereof. These phrases also refer to DNA or RNA 20 of genomic or synthetic origin which may be single-stranded or double-stranded and may represent the sense or the antisense strand, to peptide nucleic acid (PNA), or to any DNA-like or RNA-like material.

"Operably linked" refers to the situation in which a first nucleic acid sequence is placed in a functional relationship with a second nucleic acid sequence. For instance, a promoter is operably linked to a coding sequence if the promoter affects the transcription or expression of the coding sequence. Operably linked DNA sequences may be in close proximity or contiguous and, where necessary to join two protein coding regions, in the same reading frame.

"Peptide nucleic acid" (PNA) refers to an antisense molecule or anti-gene agent which comprises an oligonucleotide of at least about 5 nucleotides in length linked to a peptide backbone 30 of amino acid residues ending in lysine. The terminal lysine confers solubility to the composition. PNAs preferentially bind complementary single stranded DNA or RNA and stop transcript elongation, and may be pegylated to extend their lifespan in the cell.

"Post-translational modification" of an KAP may involve lipidation, glycosylation, phosphorylation, acetylation, racemization, proteolytic cleavage, and other modifications known in 35 the art. These processes may occur synthetically or biochemically. Biochemical modifications will

vary by cell type depending on the enzymatic milieu of KAP.

"Probe" refers to nucleic acid sequences encoding KAP, their complements, or fragments thereof, which are used to detect identical, allelic or related nucleic acid sequences. Probes are isolated oligonucleotides or polynucleotides attached to a detectable label or reporter molecule.

5 Typical labels include radioactive isotopes, ligands, chemiluminescent agents, and enzymes.

"Primers" are short nucleic acids, usually DNA oligonucleotides, which may be annealed to a target polynucleotide by complementary base-pairing. The primer may then be extended along the target

DNA strand by a DNA polymerase enzyme. Primer pairs can be used for amplification (and

identification) of a nucleic acid sequence, e.g., by the polymerase chain reaction (PCR).

Probes and primers as used in the present invention typically comprise at least 15 contiguous nucleotides of a known sequence. In order to enhance specificity, longer probes and primers may also be employed, such as probes and primers that comprise at least 20, 25, 30, 40, 50, 60, 70, 80, 90, 100, or at least 150 consecutive nucleotides of the disclosed nucleic acid sequences. Probes and primers may be considerably longer than these examples, and it is understood that any 15 length supported by the specification, including the tables, figures, and Sequence Listing, may be used.

Methods for preparing and using probes and primers are described in the references, for example Sambrook, J. et al. (1989) Molecular Cloning: A Laboratory Manual, 2nd ed., vol. 1-3, Cold Spring Harbor Press, Plainview NY; Ausubel, F.M. et al. (1987) Current Protocols in Molecular 20 Biology, Greene Publ. Assoc. & Wiley-Intersciences, New York NY; Innis, M. et al. (1990) PCR Protocols, A Guide to Methods and Applications, Academic Press, San Diego CA. PCR primer pairs can be derived from a known sequence, for example, by using computer programs intended for that purpose such as Primer (Version 0.5, 1991, Whitehead Institute for Biomedical Research, Cambridge MA).

Oligonucleotides for use as primers are selected using software known in the art for such purpose. For example, OLIGO 4.06 software is useful for the selection of PCR primer pairs of up to 100 nucleotides each, and for the analysis of oligonucleotides and larger polynucleotides of up to 5,000 nucleotides from an input polynucleotide sequence of up to 32 kilobases. Similar primer selection programs have incorporated additional features for expanded capabilities. For example, 30 the PrimOU primer selection program (available to the public from the Genome Center at University of Texas South West Medical Center, Dallas TX) is capable of choosing specific primers from megabase sequences and is thus useful for designing primers on a genome-wide scope. The Primer3 primer selection program (available to the public from the Whitehead Institute/MIT Center for Genome Research, Cambridge MA) allows the user to input a "mispriming library," in which sequences to avoid as primer binding sites are user-specified. Primer3 is useful, in particular, for

the selection of oligonucleotides for microarrays. (The source code for the latter two primer selection programs may also be obtained from their respective sources and modified to meet the user's specific needs.) The PrimeGen program (available to the public from the UK Human Genome Mapping Project Resource Centre, Cambridge UK) designs primers based on multiple sequence alignments, thereby allowing selection of primers that hybridize to either the most conserved or least conserved regions of aligned nucleic acid sequences. Hence, this program is useful for identification of both unique and conserved oligonucleotides and polynucleotide fragments. The oligonucleotides and polynucleotide fragments identified by any of the above selection methods are useful in hybridization technologies, for example, as PCR or sequencing 10 primers, microarray elements, or specific probes to identify fully or partially complementary polynucleotides in a sample of nucleic acids. Methods of oligonucleotide selection are not limited to those described above.

A "recombinant nucleic acid" is a sequence that is not naturally occurring or has a sequence that is made by an artificial combination of two or more otherwise separated segments of sequence.

15 This artificial combination is often accomplished by chemical synthesis or, more commonly, by the artificial manipulation of isolated segments of nucleic acids, e.g., by genetic engineering techniques such as those described in Sambrook, supra. The term recombinant includes nucleic acids that have been altered solely by addition, substitution, or deletion of a portion of the nucleic acid. Frequently, a recombinant nucleic acid may include a nucleic acid sequence operably linked to a promoter sequence. Such a recombinant nucleic acid may be part of a vector that is used, for example, to transform a cell.

Alternatively, such recombinant nucleic acids may be part of a viral vector, e.g., based on a vaccinia virus, that could be use to vaccinate a mammal wherein the recombinant nucleic acid is expressed, inducing a protective immunological response in the mammal.

A "regulatory element" refers to a nucleic acid sequence usually derived from untranslated regions of a gene and includes enhancers, promoters, introns, and 5' and 3' untranslated regions (UTRs). Regulatory elements interact with host or viral proteins which control transcription, translation, or RNA stability.

"Reporter molecules" are chemical or biochemical moieties used for labeling a nucleic acid, 30 amino acid, or antibody. Reporter molecules include radionuclides; enzymes; fluorescent, chemiluminescent, or chromogenic agents; substrates; cofactors; inhibitors; magnetic particles; and other moieties known in the art.

An "RNA equivalent," in reference to a DNA sequence, is composed of the same linear sequence of nucleotides as the reference DNA sequence with the exception that all occurrences of the nitrogenous base thymine are replaced with uracil, and the sugar backbone is composed of

ribose instead of deoxyribose.

The term "sample" is used in its broadest sense. A sample suspected of containing KAP, nucleic acids encoding KAP, or fragments thereof may comprise a bodily fluid; an extract from a cell, chromosome, organelle, or membrane isolated from a cell; a cell; genomic DNA, RNA, or 5 cDNA, in solution or bound to a substrate; a tissue; a tissue print; etc.

The terms "specific binding" and "specifically binding" refer to that interaction between a protein or peptide and an agonist, an antibody, an antagonist, a small molecule, or any natural or synthetic binding composition. The interaction is dependent upon the presence of a particular structure of the protein, e.g., the antigenic determinant or epitope, recognized by the binding 10 molecule. For example, if an antibody is specific for epitope "A," the presence of a polypeptide comprising the epitope A, or the presence of free unlabeled A, in a reaction containing free labeled A and the antibody will reduce the amount of labeled A that binds to the antibody.

The term "substantially purified" refers to nucleic acid or amino acid sequences that are removed from their natural environment and are isolated or separated, and are at least 60% free, 15 preferably at least 75% free, and most preferably at least 90% free from other components with which they are naturally associated.

A "substitution" refers to the replacement of one or more amino acid residues or nucleotides by different amino acid residues or nucleotides, respectively.

"Substrate" refers to any suitable rigid or semi-rigid support including membranes, filters, 20 chips, slides, wafers, fibers, magnetic or nonmagnetic beads, gels, tubing, plates, polymers, microparticles and capillaries. The substrate can have a variety of surface forms, such as wells, trenches, pins, channels and pores, to which polynucleotides or polypeptides are bound.

A "transcript image" or "expression profile" refers to the collective pattern of gene expression by a particular cell type or tissue under given conditions at a given time.

- 25 "Transformation" describes a process by which exogenous DNA is introduced into a recipient cell. Transformation may occur under natural or artificial conditions according to various methods well known in the art, and may rely on any known method for the insertion of foreign nucleic acid sequences into a prokaryotic or eukaryotic host cell. The method for transformation is selected based on the type of host cell being transformed and may include, but is not limited to,
- 30 bacteriophage or viral infection, electroporation, heat shock, lipofection, and particle bombardment.

 The term "transformed cells" includes stably transformed cells in which the inserted DNA is

 capable of replication either as an autonomously replicating plasmid or as part of the host

 chromosome, as well as transiently transformed cells which express the inserted DNA or RNA for
 limited periods of time.
- 35 A "transgenic organism," as used herein, is any organism, including but not limited to

animals and plants, in which one or more of the cells of the organism contains heterologous nucleic acid introduced by way of human intervention, such as by transgenic techniques well known in the art. The nucleic acid is introduced into the cell, directly or indirectly by introduction into a precursor of the cell, by way of deliberate genetic manipulation, such as by microinjection or by infection with a recombinant virus. The term genetic manipulation does not include classical cross-breeding, or in vitro fertilization, but rather is directed to the introduction of a recombinant DNA molecule. The transgenic organisms contemplated in accordance with the present invention include bacteria, cyanobacteria, fungi, plants and animals. The isolated DNA of the present invention can be introduced into the host by methods known in the art, for example infection, transferring to transferring the DNA of the present invention into such organisms are widely known and provided in references such as Sambrook et al. (1989), supra.

A "variant" of a particular nucleic acid sequence is defined as a nucleic acid sequence having at least 40% sequence identity to the particular nucleic acid sequence over a certain length of 15 one of the nucleic acid sequences using blastn with the "BLAST 2 Sequences" tool Version 2.0.9 (May-07-1999) set at default parameters. Such a pair of nucleic acids may show, for example, at least 50%, at least 60%, at least 70%, at least 80%, at least 85%, at least 90%, at least 91%, at least 92%, at least 93%, at least 94%, at least 95%, at least 96%, at least 97%, at least 98%, or at least 99% or greater sequence identity over a certain defined length. A variant may be described as, for 20 example, an "allelic" (as defined above), "splice," "species," or "polymorphic" variant. A splice variant may have significant identity to a reference molecule, but will generally have a greater or lesser number of polynucleotides due to alternate splicing of exons during mRNA processing. The corresponding polypeptide may possess additional functional domains or lack domains that are present in the reference molecule. Species variants are polynucleotide sequences that vary from one 25 species to another. The resulting polypeptides will generally have significant amino acid identity relative to each other. A polymorphic variant is a variation in the polynucleotide sequence of a particular gene between individuals of a given species. Polymorphic variants also may encompass "single nucleotide polymorphisms" (SNPs) in which the polynucleotide sequence varies by one nucleotide base. The presence of SNPs may be indicative of, for example, a certain population, a 30 disease state, or a propensity for a disease state.

A "variant" of a particular polypeptide sequence is defined as a polypeptide sequence having at least 40% sequence identity to the particular polypeptide sequence over a certain length of one of the polypeptide sequences using blastp with the "BLAST 2 Sequences" tool Version 2.0.9 (May-07-1999) set at default parameters. Such a pair of polypeptides may show, for example, at least 50%, at least 60%, at least 70%, at least 80%, at least 91%, at least 92%, at least

93%, at least 94%, at least 95%, at least 96%, at least 97%, at least 98%, or at least 99% or greater sequence identity over a certain defined length of one of the polypeptides.

THE INVENTION

The invention is based on the discovery of new human kinases and phosphatases (KAP), the polynucleotides encoding KAP, and the use of these compositions for the diagnosis, treatment, or prevention of cardiovascular diseases, immune system disorders, neurological disorders, disorders affecting growth and development, lipid disorders, cell proliferative disorders, and cancers.

Table 1 summarizes the nomenclature for the full length polynucleotide and polypeptide 10 sequences of the invention. Each polynucleotide and its corresponding polypeptide are correlated to a single Incyte project identification number (Incyte Project ID). Each polypeptide sequence is denoted by both a polypeptide sequence identification number (Polypeptide SEQ ID NO:) and an Incyte polypeptide sequence number (Incyte Polypeptide ID) as shown. Each polynucleotide sequence is denoted by both a polynucleotide sequence identification number (Polynucleotide SEQ 15 ID NO:) and an Incyte polynucleotide consensus sequence number (Incyte Polynucleotide ID) as shown.

Table 2 shows sequences with homology to the polypeptides of the invention as identified by BLAST analysis against the GenBank protein (genpept) database. Columns 1 and 2 show the polypeptide sequence identification number (Polypeptide SEQ ID NO:) and the corresponding 20 Incyte polypeptide sequence number (Incyte Polypeptide ID) for polypeptides of the invention. Column 3 shows the GenBank identification number (GenBank ID NO:) of the nearest GenBank homolog. Column 4 shows the probability scores for the matches between each polypeptide and its homolog(s). Column 5 shows the annotation of the GenBank homolog(s) along with relevant citations where applicable, all of which are expressly incorporated by reference herein.

Table 3 shows various structural features of the polypeptides of the invention. Columns 1 and 2 show the polypeptide sequence identification number (SEQ ID NO:) and the corresponding Incyte polypeptide sequence number (Incyte Polypeptide ID) for each polypeptide of the invention. Column 3 shows the number of amino acid residues in each polypeptide. Column 4 shows potential phosphorylation sites, and column 5 shows potential glycosylation sites, as determined by the 30 MOTIFS program of the GCG sequence analysis software package (Genetics Computer Group, Madison WI). Column 6 shows amino acid residues comprising signature sequences, domains, and motifs. Column 7 shows analytical methods for protein structure/function analysis and in some cases, searchable databases to which the analytical methods were applied.

Together, Tables 2 and 3 summarize the properties of polypeptides of the invention, and 35 these properties establish that the claimed polypeptides are kinases and phosphatases. For example,

SEQ ID NO:1 is 79% identical to rat protein tyrosine phosphatase TD14 (GenBank ID g3598974) as determined by the Basic Local Alignment Search Tool (BLAST). (See Table 2.) The BLAST probability score is 0.0, which indicates the probability of obtaining the observed polypeptide sequence alignment by chance. SEQ ID NO:1 also contains protein-tyrosine phosphatase domain as 5 determined by searching for statistically significant matches in the hidden Markov model (HMM)-based PFAM database of conserved protein family domains. (See Table 3.) Data from BLIMPS, PROFILESCAN and MOTIFS analyses provide further corroborative evidence that SEQ ID NO:1 is a protein-tyrosine phosphatase.

In an alternative example, SEQ ID NO:3 is 34% identical to Fagus sylvatica protein

10 phosphatase 2C (PP2C, GenBank ID g7768151) as determined by the Basic Local Alignment

Search Tool (BLAST). (See Table 2.) The BLAST probability score is 6.4e-17, which indicates the

probability of obtaining the observed polypeptide sequence alignment by chance. SEQ ID NO:3

also shares 45% identity with a putative Caenorhabditis elegans PP2C (GenBank ID g2804429),

based on BLAST analysis, with a probability score of 2.4e-71. SEQ ID NO:3 contains protein

15 phosphatase 2C domains as determined by searching for statistically significant matches in the

hidden Markov model (HMM)-based PFAM database of conserved protein family domains. (See

Table 3.) Data from BLIMPS analysis provide further corroborative evidence that SEQ ID NO:3 is

a protein phosphatase 2C.

In an alternative example, SEQ ID NO:5 is 25% identical to human protein kinase PAK5

20 (GenBank ID g7649810) as determined by the Basic Local Alignment Search Tool (BLAST). (See Table 2.) The BLAST probability score is 7.2e-14, which indicates the probability of obtaining the observed polypeptide sequence alignment by chance. SEQ ID NO:5 also contains a eukaryotic protein kinase domain as determined by searching for statistically significant matches in the hidden Markov model (HMM)-based PFAM database of conserved protein family domains. (See Table 3.)

25 Data from TMAP analysis as well as BLIMPS and BLAST analyses of the PRODOM and DOMO databases provide further corroborative evidence that SEQ ID NO:5 is a membrane-bound kinase.

In an alternative example, SEQ ID NO:6 is 1511 amino acid residues in length and is 97% identical over 1494 residues to human MEK kinase I (GenBank ID g2815888) as determined by the Basic Local Alignment Search Tool (BLAST). (See Table 2.) The BLAST probability score is 0.0, 30 which indicates the probability of obtaining the observed polypeptide sequence alignment by chance. SEQ ID NO:6 also contains a eukaryotic protein kinase domain as determined by searching for statistically significant matches in the hidden Markov model (HMM)-based PFAM database of conserved protein family domains. (See Table 3.) Data from BLIMPS, MOTIFS, and PROFILESCAN analyses provide further corroborative evidence that SEQ ID NO:6 is protein 15 kinase.

In an alternative example, SEQ ID NO:9 is 87% identical to murine protein kinase

(GenBank ID g406058) as determined by the Basic Local Alignment Search Tool (BLAST). (See Table 2.) The BLAST probability score is 0.0, which indicates the probability of obtaining the observed polypeptide sequence alignment by chance. SEQ ID NO:9 also contains an eukaryotic protein kinase domain and a PDZ domain as determined by searching for statistically significant 5 matches in the hidden Markov model (HMM)-based PFAM database of conserved protein family domains. (See Table 3.) Data from BLIMPS, MOTIFS, and PROFILESCAN analyses provide further corroborative evidence that SEQ ID NO:9 is a protein kinase.

In an alternative example, SEQ ID NO:16 is 61% identical to human mitogen-activated kinase kinase kinase 5 (GenBank ID g1679668) as determined by the Basic Local Alignment Search 10 Tool (BLAST). (See Table 2.) The BLAST probability score is 0.0, which indicates the probability of obtaining the observed polypeptide sequence alignment by chance. SEQ ID NO:16 also contains a eukaryotic protein kinase domain as determined by searching for statistically significant matches in the hidden Markov model (HMM)-based PFAM database of conserved protein family domains. (See Table 3.) Data from BLIMPS, MOTIFS, and PROFILESCAN analyses provide further 15 corroborative evidence that SEQ ID NO:16 is a mitogen activated protein kinase kinase kinase.

In an alternative example, SEQ ID NO:18 is 83% identical from residues 4 to 372 to mouse protein kinase (GenBank ID g406058) as determined by the Basic Local Alignment Search Tool (BLAST). (See Table 2.) The BLAST probability score is 0.0, which indicates the probability of obtaining the observed polypeptide sequence alignment by chance. SEQ ID NO:18 also contains a 20 eukaryotic protein kinase domain and a PDZ domain as determined by searching for statistically significant matches in the hidden Markov model (HMM)-based PFAM database of conserved protein family domains. (See Table 3.) Data from BLIMPS, MOTIFS, and PROFILESCAN analyses provide further corroborative evidence that SEQ ID NO:18 is a serine/threonine protein kinase.

In an alternative example, SEQ ID NO:19 is 95% identical, from residue M1 to residue V988, to Rattus norvegicus mytonic dystrophy kinase-related Cdc42-binding kinase (GenBank ID g2736151) as determined by the Basic Local Alignment Search Tool (BLAST). (See Table 2.) The BLAST probability score is 0.0, which indicates the probability of obtaining the observed polypeptide sequence alignment by chance. SEQ ID NO:19 also contains a protein kinase C 30 terminal domain and a eukaryotic protein kinase domain as determined by searching for statistically significant matches in the hidden Markov model (HMM)-based PFAM database of conserved protein family domains. (See Table 3.) Data from BLIMPS, MOTIES, and additional BLAST analyses provide further corroborative evidence that SEQ ID NO:19 is a protein kinase.

35 SEQ ID NO:2, SEQ ID NO:4, SEQ ID NO:7, SEQ ID NO:8, SEQ ID NO:10-15, SEQ ID NO:17,

and SEQ ID NO:20 were analyzed and annotated in a similar manner. The algorithms and parameters for the analysis of SEQ ID NO:1-20 are described in Table 7.

As shown in Table 4, the full length polynucleotide sequences of the present invention were assembled using cDNA sequences or coding (exon) sequences derived from genomic DNA, or any combination of these two types of sequences. Column 1 lists the polynucleotide sequence identification number (Polynucleotide SEQ ID NO:), the corresponding Incyte polynucleotide consensus sequence number (Incyte ID) for each polynucleotide of the invention, and the length of each polynucleotide sequence in basepairs. Column 2 shows the nucleotide start (5') and stop (3') positions of the cDNA and/or genomic sequences used to assemble the full length polynucleotide sequences of the invention, and of fragments of the polynucleotide sequences which are useful, for example, in hybridization or amplification technologies that identify SEQ ID NO:21-40 or that distinguish between SEQ ID NO:21-40 and related polynucleotide sequences.

The polynucleotide fragments described in Column 2 of Table 4 may refer specifically, for example, to Incyte cDNAs derived from tissue-specific cDNA libraries or from pooled cDNA 15 libraries. Alternatively, the polynucleotide fragments described in column 2 may refer to GenBank cDNAs or ESTs which contributed to the assembly of the full length polynucleotide sequences. In addition, the polynucleotide fragments described in column 2 may identify sequences derived from the ENSEMBL (The Sanger Centre, Cambridge, UK) database (i.e., those sequences including the designation "ENST"). Alternatively, the polynucleotide fragments described in column 2 may be 20 derived from the NCBI RefSeq Nucleotide Sequence Records Database (i.e., those sequences including the designation "NM" or "NT") or the NCBI RefSeq Protein Sequence Records (i.e., those sequences including the designation "NP"). Alternatively, the polynucleotide fragments described in column 2 may refer to assemblages of both cDNA and Genscan-predicted exons brought together by an "exon stitching" algorithm. For example, a polynucleotide sequence 25 identified as FL_XXXXXX_ N_1 _ N_2 _YYYYY_ N_3 _ N_4 represents a "stitched" sequence in which XXXXXX is the identification number of the cluster of sequences to which the algorithm was applied, and YYYYY is the number of the prediction generated by the algorithm, and $N_{1,2,3...}$, if present, represent specific exons that may have been manually edited during analysis (See Example V). Alternatively, the polynucleotide fragments in column 2 may refer to assemblages of exons 30 brought together by an "exon-stretching" algorithm. For example, a polynucleotide sequence identified as FLXXXXXX_gAAAAA_gBBBBB_1_N is a "stretched" sequence, with XXXXXX being the Incyte project identification number, gAAAAA being the GenBank identification number of the human genomic sequence to which the "exon-stretching" algorithm was applied, gBBBBB being the GenBank identification number or NCBI RefSeq identification number of the nearest GenBank 35 protein homolog, and N referring to specific exons (See Example V). In instances where a RefSeq

sequence was used as a protein homolog for the "exon-stretching" algorithm, a RefSeq identifier (denoted by "NM," "NP," or "NT") may be used in place of the GenBank identifier (i.e., gBBBBB).

Alternatively, a prefix identifies component sequences that were hand-edited, predicted from genomic DNA sequences, or derived from a combination of sequence analysis methods. The 5 following Table lists examples of component sequence prefixes and corresponding sequence analysis methods associated with the prefixes (see Example IV and Example V).

Prefix	Type of analysis and/or examples of programs
GNN, GFG,	Exon prediction from genomic sequences using, for example,
ENST	GENSCAN (Stanford University, CA, USA) or FGENES
	(Computer Genomics Group, The Sanger Centre, Cambridge, UK).
GBI	Hand-edited analysis of genomic sequences.
FL	Stitched or stretched genomic sequences (see Example V).
INCY	Full length transcript and exon prediction from mapping of EST
	sequences to the genome. Genomic location and EST composition
	data are combined to predict the exons and resulting transcript.

In some cases, Incyte cDNA coverage redundant with the sequence coverage shown in 15 Table 4 was obtained to confirm the final consensus polynucleotide sequence, but the relevant Incyte cDNA identification numbers are not shown.

Table 5 shows the representative cDNA libraries for those full length polynucleotide sequences which were assembled using Incyte cDNA sequences. The representative cDNA library is the Incyte cDNA library which is most frequently represented by the Incyte cDNA sequences which were used to assemble and confirm the above polynucleotide sequences. The tissues and vectors which were used to construct the cDNA libraries shown in Table 5 are described in Table 6.

The invention also encompasses KAP variants. A preferred KAP variant is one which has at least about 80%, or alternatively at least about 90%, or even at least about 95% amino acid sequence identity to the KAP amino acid sequence, and which contains at least one functional or 25 structural characteristic of KAP.

The invention also encompasses polynucleotides which encode KAP. In a particular embodiment, the invention encompasses a polynucleotide sequence comprising a sequence selected from the group consisting of SEQ ID NO:21-40, which encodes KAP. The polynucleotide sequences of SEQ ID NO:21-40, as presented in the Sequence Listing, embrace the equivalent RNA sequences, wherein occurrences of the nitrogenous base thymine are replaced with uracil, and the sugar backbone is composed of ribose instead of deoxyribose.

The invention also encompasses a variant of a polynucleotide sequence encoding KAP. In

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particular, such a variant polynucleotide sequence will have at least about 70%, or alternatively at least about 85%, or even at least about 95% polynucleotide sequence identity to the polynucleotide sequence encoding KAP. A particular aspect of the invention encompasses a variant of a polynucleotide sequence comprising a sequence selected from the group consisting of SEQ ID NO:21-40 which has at least about 70%, or alternatively at least about 85%, or even at least about 95% polynucleotide sequence identity to a nucleic acid sequence selected from the group consisting of SEQ ID NO:21-40. Any one of the polynucleotide variants described above can encode an amino acid sequence which contains at least one functional or structural characteristic of KAP.

In addition, or in the alternative, a polynucleotide variant of the invention is a splice variant 10 of a polynucleotide sequence encoding KAP. A splice variant may have portions which have significant sequence identity to the polynucleotide sequence encoding KAP, but will generally have a greater or lesser number of polynucleotides due to additions or deletions of blocks of sequence arising from alternate splicing of exons during mRNA processing. A splice variant may have less than about 70%, or alternatively less than about 60%, or alternatively less than about 50% polynucleotide sequence identity to the polynucleotide sequence encoding KAP over its entire length; however, portions of the splice variant will have at least about 70%, or alternatively at least about 85%, or alternatively at least about 95%, or alternatively 100% polynucleotide sequence identity to portions of the polynucleotide sequence encoding KAP. Any one of the splice variants described above can encode an amino acid sequence which contains at least one functional or 20 structural characteristic of KAP.

It will be appreciated by those skilled in the art that as a result of the degeneracy of the genetic code, a multitude of polynucleotide sequences encoding KAP, some bearing minimal similarity to the polynucleotide sequences of any known and naturally occurring gene, may be produced. Thus, the invention contemplates each and every possible variation of polynucleotide sequence that could be made by selecting combinations based on possible codon choices. These combinations are made in accordance with the standard triplet genetic code as applied to the polynucleotide sequence of naturally occurring KAP, and all such variations are to be considered as being specifically disclosed.

Although nucleotide sequences which encode KAP and its variants are generally capable of 30 hybridizing to the nucleotide sequence of the naturally occurring KAP under appropriately selected conditions of stringency, it may be advantageous to produce nucleotide sequences encoding KAP or its derivatives possessing a substantially different codon usage, e.g., inclusion of non-naturally occurring codons. Codons may be selected to increase the rate at which expression of the peptide occurs in a particular prokaryotic or eukaryotic host in accordance with the frequency with which 35 particular codons are utilized by the host. Other reasons for substantially altering the nucleotide

sequence encoding KAP and its derivatives without altering the encoded amino acid sequences include the production of RNA transcripts having more desirable properties, such as a greater half-life, than transcripts produced from the naturally occurring sequence.

The invention also encompasses production of DNA sequences which encode KAP and 5 KAP derivatives, or fragments thereof, entirely by synthetic chemistry. After production, the synthetic sequence may be inserted into any of the many available expression vectors and cell systems using reagents well known in the art. Moreover, synthetic chemistry may be used to introduce mutations into a sequence encoding KAP or any fragment thereof.

Also encompassed by the invention are polynucleotide sequences that are capable of 10 hybridizing to the claimed polynucleotide sequences, and, in particular, to those shown in SEQ ID NO:21-40 and fragments thereof under various conditions of stringency. (See, e.g., Wahl, G.M. and S.L. Berger (1987) Methods Enzymol. 152:399-407; Kimmel, A.R. (1987) Methods Enzymol. 152:507-511.) Hybridization conditions, including annealing and wash conditions, are described in "Definitions."

- Methods for DNA sequencing are well known in the art and may be used to practice any of the embodiments of the invention. The methods may employ such enzymes as the Klenow fragment of DNA polymerase I, SEQUENASE (US Biochemical, Cleveland OH), Taq polymerase (Applied Biosystems), thermostable T7 polymerase (Amersham Pharmacia Biotech, Piscataway NJ), or combinations of polymerases and proofreading exonucleases such as those found in the
- 20 ELONGASE amplification system (Life Technologies, Gaithersburg MD). Preferably, sequence preparation is automated with machines such as the MICROLAB 2200 liquid transfer system (Hamilton, Reno NV), PTC200 thermal cycler (MJ Research, Watertown MA) and ABI CATALYST 800 thermal cycler (Applied Biosystems). Sequencing is then carried out using either the ABI 373 or 377 DNA sequencing system (Applied Biosystems), the MEGABACE 1000 DNA
- 25 sequencing system (Molecular Dynamics, Sunnyvale CA), or other systems known in the art. The resulting sequences are analyzed using a variety of algorithms which are well known in the art. (See, e.g., Ausubel, F.M. (1997) Short Protocols in Molecular Biology, John Wiley & Sons, New York NY, unit 7.7; Meyers, R.A. (1995) Molecular Biology and Biotechnology, Wiley VCH, New York NY, pp. 856-853.)
- The nucleic acid sequences encoding KAP may be extended utilizing a partial nucleotide sequence and employing various PCR-based methods known in the art to detect upstream sequences, such as promoters and regulatory elements. For example, one method which may be employed, restriction-site PCR, uses universal and nested primers to amplify unknown sequence from genomic DNA within a cloning vector. (See, e.g., Sarkar, G. (1993) PCR Methods Applic. 35 2:318-322.) Another method, inverse PCR, uses primers that extend in divergent directions to

amplify unknown sequence from a circularized template. The template is derived from restriction fragments comprising a known genomic locus and surrounding sequences. (See, e.g., Triglia, T. et al. (1988) Nucleic Acids Res. 16:8186.) A third method, capture PCR, involves PCR amplification of DNA fragments adjacent to known sequences in human and yeast artificial chromosome DNA.

5 (See, e.g., Lagerstrom, M. et al. (1991) PCR Methods Applic. 1:111-119.) In this method, multiple restriction enzyme digestions and ligations may be used to insert an engineered double-stranded sequence into a region of unknown sequence before performing PCR. Other methods which may be used to retrieve unknown sequences are known in the art. (See, e.g., Parker, J.D. et al. (1991) Nucleic Acids Res. 19:3055-3060). Additionally, one may use PCR, nested primers, and 10 PROMOTERFINDER libraries (Clontech, Palo Alto CA) to walk genomic DNA. This procedure avoids the need to screen libraries and is useful in finding intron/exon junctions. For all PCR-based methods, primers may be designed using commercially available software, such as OLIGO 4.06 primer analysis software (National Biosciences, Plymouth MN) or another appropriate program, to be about 22 to 30 nucleotides in length, to have a GC content of about 50% or more, and to anneal

15 to the template at temperatures of about 68°C to 72°C.

When screening for full length cDNAs, it is preferable to use libraries that have been size-selected to include larger cDNAs. In addition, random-primed libraries, which often include sequences containing the 5' regions of genes, are preferable for situations in which an oligo d(T) library does not yield a full-length cDNA. Genomic libraries may be useful for extension of 20 sequence into 5' non-transcribed regulatory regions.

Capillary electrophoresis systems which are commercially available may be used to analyze the size or confirm the nucleotide sequence of sequencing or PCR products. In particular, capillary sequencing may employ flowable polymers for electrophoretic separation, four different nucleotide-specific, laser-stimulated fluorescent dyes, and a charge coupled device camera for detection of the emitted wavelengths. Output/light intensity may be converted to electrical signal using appropriate software (e.g., GENOTYPER and SEQUENCE NAVIGATOR, Applied Biosystems), and the entire process from loading of samples to computer analysis and electronic data display may be computer controlled. Capillary electrophoresis is especially preferable for sequencing small DNA fragments which may be present in limited amounts in a particular sample.

In another embodiment of the invention, polynucleotide sequences or fragments thereof which encode KAP may be cloned in recombinant DNA molecules that direct expression of KAP, or fragments or functional equivalents thereof, in appropriate host cells. Due to the inherent degeneracy of the genetic code, other DNA sequences which encode substantially the same or a functionally equivalent amino acid sequence may be produced and used to express KAP.

The nucleotide sequences of the present invention can be engineered using methods

generally known in the art in order to alter KAP-encoding sequences for a variety of purposes including, but not limited to, modification of the cloning, processing, and/or expression of the gene product. DNA shuffling by random fragmentation and PCR reassembly of gene fragments and synthetic oligonucleotides may be used to engineer the nucleotide sequences. For example, oligonucleotide-mediated site-directed mutagenesis may be used to introduce mutations that create new restriction sites, alter glycosylation patterns, change codon preference, produce splice variants, and so forth.

The nucleotides of the present invention may be subjected to DNA shuffling techniques such as MOLECULARBREEDING (Maxygen Inc., Santa Clara CA; described in U.S. Patent No. 10 5,837,458; Chang, C.-C. et al. (1999) Nat. Biotechnol. 17:793-797; Christians, F.C. et al. (1999) Nat. Biotechnol. 17:259-264; and Crameri, A. et al. (1996) Nat. Biotechnol. 14:315-319) to alter or improve the biological properties of KAP, such as its biological or enzymatic activity or its ability to bind to other molecules or compounds. DNA shuffling is a process by which a library of gene variants is produced using PCR-mediated recombination of gene fragments. The library is then 15 subjected to selection or screening procedures that identify those gene variants with the desired properties. These preferred variants may then be pooled and further subjected to recursive rounds of DNA shuffling and selection/screening. Thus, genetic diversity is created through "artificial" breeding and rapid molecular evolution. For example, fragments of a single gene containing random point mutations may be recombined, screened, and then reshuffled until the desired 20 properties are optimized. Alternatively, fragments of a given gene may be recombined with fragments of homologous genes in the same gene family, either from the same or different species, thereby maximizing the genetic diversity of multiple naturally occurring genes in a directed and controllable manner.

In another embodiment, sequences encoding KAP may be synthesized, in whole or in part, 25 using chemical methods well known in the art. (See, e.g., Caruthers, M.H. et al. (1980) Nucleic Acids Symp. Ser. 7:215-223; and Horn, T. et al. (1980) Nucleic Acids Symp. Ser. 7:225-232.)

Alternatively, KAP itself or a fragment thereof may be synthesized using chemical methods. For example, peptide synthesis can be performed using various solution-phase or solid-phase techniques. (See, e.g., Creighton, T. (1984) Proteins, Structures and Molecular Properties, WH 30 Freeman, New York NY, pp. 55-60; and Roberge, J.Y. et al. (1995) Science 269:202-204.)

Automated synthesis may be achieved using the ABI 431A peptide synthesizer (Applied Biosystems). Additionally, the amino acid sequence of KAP, or any part thereof, may be altered during direct synthesis and/or combined with sequences from other proteins, or any part thereof, to produce a variant polypeptide or a polypeptide having a sequence of a naturally occurring 35 polypeptide.

The peptide may be substantially purified by preparative high performance liquid chromatography. (See, e.g., Chiez, R.M. and F.Z. Regnier (1990) Methods Enzymol. 182:392-421.) The composition of the synthetic peptides may be confirmed by amino acid analysis or by sequencing. (See, e.g., Creighton, supra, pp. 28-53.)

In order to express a biologically active KAP, the nucleotide sequences encoding KAP or derivatives thereof may be inserted into an appropriate expression vector, i.e., a vector which contains the necessary elements for transcriptional and translational control of the inserted coding sequence in a suitable host. These elements include regulatory sequences, such as enhancers, constitutive and inducible promoters, and 5' and 3' untranslated regions in the vector and in polynucleotide sequences encoding KAP. Such elements may vary in their strength and specificity. Specific initiation signals may also be used to achieve more efficient translation of sequences encoding KAP. Such signals include the ATG initiation codon and adjacent sequences, e.g. the Kozak sequence. In cases where sequences encoding KAP and its initiation codon and upstream

15 transcriptional or translational control signals may be needed. However, in cases where only coding sequence, or a fragment thereof, is inserted, exogenous translational control signals including an inframe ATG initiation codon should be provided by the vector. Exogenous translational elements and initiation codons may be of various origins, both natural and synthetic. The efficiency of expression may be enhanced by the inclusion of enhancers appropriate for the particular host cell 20 system used. (See, e.g., Scharf, D. et al. (1994) Results Probl. Cell Differ. 20:125-162.)

regulatory sequences are inserted into the appropriate expression vector, no additional

Methods which are well known to those skilled in the art may be used to construct expression vectors containing sequences encoding KAP and appropriate transcriptional and translational control elements. These methods include <u>in vitro</u> recombinant DNA techniques, synthetic techniques, and <u>in vivo</u> genetic recombination. (See, e.g., Sambrook, J. et al. (1989) 25 Molecular Cloning, A Laboratory Manual, Cold Spring Harbor Press, Plainview NY, ch. 4, 8, and

16-17; Ausubel, F.M. et al. (1995) <u>Current Protocols in Molecular Biology</u>, John Wiley & Sons, New York NY, ch. 9, 13, and 16.)

A variety of expression vector/host systems may be utilized to contain and express sequences encoding KAP. These include, but are not limited to, microorganisms such as bacteria 30 transformed with recombinant bacteriophage, plasmid, or cosmid DNA expression vectors; yeast transformed with yeast expression vectors; insect cell systems infected with viral expression vectors (e.g., baculovirus); plant cell systems transformed with viral expression vectors (e.g., cauliflower mosaic virus, CaMV, or tobacco mosaic virus, TMV) or with bacterial expression vectors (e.g., Ti or pBR322 plasmids); or animal cell systems. (See, e.g., Sambrook, supra; Ausubel, supra; Van 35 Heeke, G. and S.M. Schuster (1989) J. Biol. Chem. 264:5503-5509; Engelhard, E.K. et al. (1994)

Proc. Natl. Acad. Sci. USA 91:3224-3227; Sandig, V. et al. (1996) Hum. Gene Ther. 7:1937-1945; Takamatsu, N. (1987) EMBO J. 6:307-311; The McGraw Hill Yearbook of Science and Technology (1992) McGraw Hill, New York NY, pp. 191-196; Logan, J. and T. Shenk (1984) Proc. Natl. Acad. Sci. USA 81:3655-3659; and Harrington, J.J. et al. (1997) Nat. Genet. 15:345-355.) Expression 5 vectors derived from retroviruses, adenoviruses, or herpes or vaccinia viruses, or from various bacterial plasmids, may be used for delivery of nucleotide sequences to the targeted organ, tissue, or cell population. (See, e.g., Di Nicola, M. et al. (1998) Cancer Gen. Ther. 5(6):350-356; Yu, M. et al. (1993) Proc. Natl. Acad. Sci. USA 90(13):6340-6344; Buller, R.M. et al. (1985) Nature 317(6040):813-815; McGregor, D.P. et al. (1994) Mol. Immunol. 31(3):219-226; and Verma, I.M. 10 and N. Somia (1997) Nature 389:239-242.) The invention is not limited by the host cell employed.

In bacterial systems, a number of cloning and expression vectors may be selected depending upon the use intended for polynucleotide sequences encoding KAP. For example, routine cloning, subcloning, and propagation of polynucleotide sequences encoding KAP can be achieved using a multifunctional E. coli vector such as PBLUESCRIPT (Stratagene, La Jolla CA) or PSPORT1

15 plasmid (Life Technologies). Ligation of sequences encoding KAP into the vector's multiple cloning site disrupts the *lacZ* gene, allowing a colorimetric screening procedure for identification of transformed bacteria containing recombinant molecules. In addition, these vectors may be useful for in vitro transcription, dideoxy sequencing, single strand rescue with helper phage, and creation of nested deletions in the cloned sequence. (See, e.g., Van Heeke, G. and S.M. Schuster (1989) J.

20 Biol. Chem. 264:5503-5509.) When large quantities of KAP are needed, e.g. for the production of antibodies, vectors which direct high level expression of KAP may be used. For example, vectors containing the strong, inducible SP6 or T7 bacteriophage promoter may be used.

Yeast expression systems may be used for production of KAP. A number of vectors containing constitutive or inducible promoters, such as alpha factor, alcohol oxidase, and PGH 25 promoters, may be used in the yeast <u>Saccharomyces cerevisiae</u> or <u>Pichia pastoris</u>. In addition, such vectors direct either the secretion or intracellular retention of expressed proteins and enable integration of foreign sequences into the host genome for stable propagation. (See, e.g., Ausubel, 1995, <u>supra</u>; Bitter, G.A. et al. (1987) Methods Enzymol. 153:516-544; and Scorer, C.A. et al. (1994) Bio/Technology 12:181-184.)

Plant systems may also be used for expression of KAP. Transcription of sequences encoding KAP may be driven by viral promoters, e.g., the 35S and 19S promoters of CaMV used alone or in combination with the omega leader sequence from TMV (Takamatsu, N. (1987) EMBO J. 6:307-311). Alternatively, plant promoters such as the small subunit of RUBISCO or heat shock promoters may be used. (See, e.g., Coruzzi, G. et al. (1984) EMBO J. 3:1671-1680; Broglie, R. et 35 al. (1984) Science 224:838-843; and Winter, J. et al. (1991) Results Probl. Cell Differ. 17:85-105.)

These constructs can be introduced into plant cells by direct DNA transformation or pathogen-mediated transfection. (See, e.g., <u>The McGraw Hill Yearbook of Science and Technology</u> (1992) McGraw Hill, New York NY, pp. 191-196.)

In mammalian cells, a number of viral-based expression systems may be utilized. In cases 5 where an adenovirus is used as an expression vector, sequences encoding KAP may be ligated into an adenovirus transcription/translation complex consisting of the late promoter and tripartite leader sequence. Insertion in a non-essential E1 or E3 region of the viral genome may be used to obtain infective virus which expresses KAP in host cells. (See, e.g., Logan, J. and T. Shenk (1984) Proc. Natl. Acad. Sci. USA 81:3655-3659.) In addition, transcription enhancers, such as the Rous 10 sarcoma virus (RSV) enhancer, may be used to increase expression in mammalian host cells. SV40 or EBV-based vectors may also be used for high-level protein expression.

Human artificial chromosomes (HACs) may also be employed to deliver larger fragments of DNA than can be contained in and expressed from a plasmid. HACs of about 6 kb to 10 Mb are constructed and delivered via conventional delivery methods (liposomes, polycationic amino polymers, or vesicles) for therapeutic purposes. (See, e.g., Harrington, J.J. et al. (1997) Nat. Genet. 15:345-355.)

For long term production of recombinant proteins in mammalian systems, stable expression of KAP in cell lines is preferred. For example, sequences encoding KAP can be transformed into cell lines using expression vectors which may contain viral origins of replication and/or endogenous 20 expression elements and a selectable marker gene on the same or on a separate vector. Following the introduction of the vector, cells may be allowed to grow for about 1 to 2 days in enriched media before being switched to selective media. The purpose of the selectable marker is to confer resistance to a selective agent, and its presence allows growth and recovery of cells which successfully express the introduced sequences. Resistant clones of stably transformed cells may be 25 propagated using tissue culture techniques appropriate to the cell type.

Any number of selection systems may be used to recover transformed cell lines. These include, but are not limited to, the herpes simplex virus thymidine kinase and adenine phosphoribosyltransferase genes, for use in tk and apr cells, respectively. (See, e.g., Wigler, M. et al. (1977) Cell 11:223-232; Lowy, I. et al. (1980) Cell 22:817-823.) Also, antimetabolite, antibiotic, or herbicide resistance can be used as the basis for selection. For example, dhfr confers resistance to methotrexate; neo confers resistance to the aminoglycosides neomycin and G-418; and als and pat confer resistance to chlorsulfuron and phosphinotricin acetyltransferase, respectively. (See, e.g., Wigler, M. et al. (1980) Proc. Natl. Acad. Sci. USA 77:3567-3570; Colbere-Garapin, F. et al. (1981) J. Mol. Biol. 150:1-14.) Additional selectable genes have been described, e.g., trpB and hisD, which alter cellular requirements for metabolites. (See, e.g., Hartman, S.C. and R.C. Mulligan

(1988) Proc. Natl. Acad. Sci. USA 85:8047-8051.) Visible markers, e.g., anthocyanins, green fluorescent proteins (GFP; Clontech), ß glucuronidase and its substrate ß-glucuronide, or luciferase and its substrate luciferin may be used. These markers can be used not only to identify transformants, but also to quantify the amount of transient or stable protein expression attributable 5 to a specific vector system. (See, e.g., Rhodes, C.A. (1995) Methods Mol. Biol. 55:121-131.)

Although the presence/absence of marker gene expression suggests that the gene of interest is also present, the presence and expression of the gene may need to be confirmed. For example, if the sequence encoding KAP is inserted within a marker gene sequence, transformed cells containing sequences encoding KAP can be identified by the absence of marker gene function. Alternatively, a 10 marker gene can be placed in tandem with a sequence encoding KAP under the control of a single promoter. Expression of the marker gene in response to induction or selection usually indicates expression of the tandem gene as well.

In general, host cells that contain the nucleic acid sequence encoding KAP and that express KAP may be identified by a variety of procedures known to those of skill in the art. These 15 procedures include, but are not limited to, DNA-DNA or DNA-RNA hybridizations, PCR amplification, and protein bioassay or immunoassay techniques which include membrane, solution, or chip based technologies for the detection and/or quantification of nucleic acid or protein sequences.

Immunological methods for detecting and measuring the expression of KAP using either 20 specific polyclonal or monoclonal antibodies are known in the art. Examples of such techniques include enzyme-linked immunosorbent assays (ELISAs), radioimmunoassays (RIAs), and fluorescence activated cell sorting (FACS). A two-site, monoclonal-based immunoassay utilizing monoclonal antibodies reactive to two non-interfering epitopes on KAP is preferred, but a competitive binding assay may be employed. These and other assays are well known in the art.

25 (See, e.g., Hampton, R. et al. (1990) Serological Methods, a Laboratory Manual, APS Press, St. Paul MN, Sect. IV; Coligan, J.E. et al. (1997) Current Protocols in Immunology, Greene Pub. Associates and Wiley-Interscience, New York NY; and Pound, J.D. (1998) Immunochemical Protocols, Humana Press, Totowa NJ.)

A wide variety of labels and conjugation techniques are known by those skilled in the art 30 and may be used in various nucleic acid and amino acid assays. Means for producing labeled hybridization or PCR probes for detecting sequences related to polynucleotides encoding KAP include oligelabeling, nick translation, end labeling, or PCR amplification using a labeled nucleotide. Alternatively, the sequences encoding KAP, or any fragments thereof, may be cloned into a vector for the production of an mRNA probe. Such vectors are known in the art, are 35 commercially available, and may be used to synthesize RNA probes in vitro by addition of an

appropriate RNA polymerase such as T7, T3, or SP6 and labeled nucleotides. These procedures may be conducted using a variety of commercially available kits, such as those provided by Amersham Pharmacia Biotech, Promega (Madison WI), and US Biochemical. Suitable reporter molecules or labels which may be used for ease of detection include radionuclides, enzymes, 5 fluorescent, chemiluminescent, or chromogenic agents, as well as substrates, cofactors, inhibitors, magnetic particles, and the like.

Host cells transformed with nucleotide sequences encoding KAP may be cultured under conditions suitable for the expression and recovery of the protein from cell culture. The protein produced by a transformed cell may be secreted or retained intracellularly depending on the sequence and/or the vector used. As will be understood by those of skill in the art, expression vectors containing polynucleotides which encode KAP may be designed to contain signal sequences which direct secretion of KAP through a prokaryotic or eukaryotic cell membrane.

In addition, a host cell strain may be chosen for its ability to modulate expression of the inserted sequences or to process the expressed protein in the desired fashion. Such modifications of 15 the polypeptide include, but are not limited to, acetylation, carboxylation, glycosylation, phosphorylation, lipidation, and acylation. Post-translational processing which cleaves a "prepro" or "pro" form of the protein may also be used to specify protein targeting, folding, and/or activity. Different host cells which have specific cellular machinery and characteristic mechanisms for post-translational activities (e.g., CHO, HeLa, MDCK, HEK293, and WI38) are available from the 20 American Type Culture Collection (ATCC, Manassas VA) and may be chosen to ensure the correct modification and processing of the foreign protein.

In another embodiment of the invention, natural, modified, or recombinant nucleic acid sequences encoding KAP may be ligated to a heterologous sequence resulting in translation of a fusion protein in any of the aforementioned host systems. For example, a chimeric KAP protein containing a heterologous moiety that can be recognized by a commercially available antibody may facilitate the screening of peptide libraries for inhibitors of KAP activity. Heterologous protein and peptide moieties may also facilitate purification of fusion proteins using commercially available affinity matrices. Such moieties include, but are not limited to, glutathione S-transferase (GST), maltose binding protein (MBP), thioredoxin (Trx), calmodulin binding peptide (CBP), 6-His, 30 FLAG, *c-myc*, and hemagglutinin (HA). GST, MBP, Trx, CBP, and 6-His enable purification of their cognate fusion proteins on immobilized glutathione, maltose, phenylarsine oxide, calmodulin, and metal-chelate resins, respectively. FLAG, *c-myc*, and hemagglutinin (HA) enable immunoaffinity purification of fusion proteins using commercially available monoclonal and polyclonal antibodies that specifically recognize these epitope tags. A fusion protein may also be 35 engineered to contain a proteolytic cleavage site located between the KAP encoding sequence and

the heterologous protein sequence, so that KAP may be cleaved away from the heterologous moiety following purification. Methods for fusion protein expression and purification are discussed in Ausubel (1995, <u>supra</u>, ch. 10). A variety of commercially available kits may also be used to facilitate expression and purification of fusion proteins.

- In a further embodiment of the invention, synthesis of radiolabeled KAP may be achieved in vitro using the TNT rabbit reticulocyte lysate or wheat germ extract system (Promega). These systems couple transcription and translation of protein-coding sequences operably associated with the T7, T3, or SP6 promoters. Translation takes place in the presence of a radiolabeled amino acid precursor, for example, ³⁵S-methionine.
- 10 KAP of the present invention or fragments thereof may be used to screen for compounds that specifically bind to KAP. At least one and up to a plurality of test compounds may be screened for specific binding to KAP. Examples of test compounds include antibodies, oligonucleotides, proteins (e.g., receptors), or small molecules.
- In one embodiment, the compound thus identified is closely related to the natural ligand of 15 KAP, e.g., a ligand or fragment thereof, a natural substrate, a structural or functional mimetic, or a natural binding partner. (See, e.g., Coligan, J.E. et al. (1991) <u>Current Protocols in Immunology</u> 1(2): Chapter 5.) Similarly, the compound can be closely related to the natural receptor to which KAP binds, or to at least a fragment of the receptor, e.g., the ligand binding site. In either case, the compound can be rationally designed using known techniques. In one embodiment, screening for 20 these compounds involves producing appropriate cells which express KAP, either as a secreted protein or on the cell membrane. Preferred cells include cells from mammals, yeast, <u>Drosophila</u>, or <u>E. coli</u>. Cells expressing KAP or cell membrane fractions which contain KAP are then contacted with a test compound and binding, stimulation, or inhibition of activity of either KAP or the compound is analyzed.
- An assay may simply test binding of a test compound to the polypeptide, wherein binding is detected by a fluorophore, radioisotope, enzyme conjugate, or other detectable label. For example, the assay may comprise the steps of combining at least one test compound with KAP, either in solution or affixed to a solid support, and detecting the binding of KAP to the compound. Alternatively, the assay may detect or measure binding of a test compound in the presence of a labeled competitor. Additionally, the assay may be carried out using cell-free preparations, chemical libraries, or natural product mixtures, and the test compound(s) may be free in solution or affixed to a solid support.

KAP of the present invention or fragments thereof may be used to screen for compounds that modulate the activity of KAP. Such compounds may include agonists, antagonists, or partial or 35 inverse agonists. In one embodiment, an assay is performed under conditions permissive for KAP

activity, wherein KAP is combined with at least one test compound, and the activity of KAP in the presence of a test compound is compared with the activity of KAP in the absence of the test compound. A change in the activity of KAP in the presence of the test compound is indicative of a compound that modulates the activity of KAP. Alternatively, a test compound is combined with an 5 in vitro or cell-free system comprising KAP under conditions suitable for KAP activity, and the assay is performed. In either of these assays, a test compound which modulates the activity of KAP may do so indirectly and need not come in direct contact with the test compound. At least one and up to a plurality of test compounds may be screened.

In another embodiment, polynucleotides encoding KAP or their mammalian homologs may 10 be "knocked out" in an animal model system using homologous recombination in embryonic stem (ES) cells. Such techniques are well known in the art and are useful for the generation of animal models of human disease. (See, e.g., U.S. Patent No. 5,175,383 and U.S. Patent No. 5,767,337.) For example, mouse ES cells, such as the mouse 129/SvJ cell line, are derived from the early mouse embryo and grown in culture. The ES cells are transformed with a vector containing the gene of 15 interest disrupted by a marker gene, e.g., the neomycin phosphotransferase gene (neo; Capecchi, M.R. (1989) Science 244:1288-1292). The vector integrates into the corresponding region of the host genome by homologous recombination. Alternatively, homologous recombination takes place using the Cre-loxP system to knockout a gene of interest in a tissue- or developmental stage-specific manner (Marth, J.D. (1996) Clin. Invest. 97:1999-2002; Wagner, K.U. et al. (1997) Nucleic Acids 20 Res. 25:4323-4330). Transformed ES cells are identified and microinjected into mouse cell blastocysts such as those from the C57BL/6 mouse strain. The blastocysts are surgically transferred to pseudopregnant dams, and the resulting chimeric progeny are genotyped and bred to produce heterozygous or homozygous strains. Transgenic animals thus generated may be tested with potential therapeutic or toxic agents.

Polynucleotides encoding KAP may also be manipulated <u>in vitro</u> in ES cells derived from human blastocysts. Human ES cells have the potential to differentiate into at least eight separate cell lineages including endoderm, mesoderm, and ectodermal cell types. These cell lineages differentiate into, for example, neural cells, hematopoietic lineages, and cardiomyocytes (Thomson, J.A. et al. (1998) Science 282:1145-1147).

Polynucleotides encoding KAP can also be used to create "knockin" humanized animals (pigs) or transgenic animals (mice or rats) to model human disease. With knockin technology, a region of a polynucleotide encoding KAP is injected into animal ES cells, and the injected sequence integrates into the animal cell genome. Transformed cells are injected into blastulae, and the blastulae are implanted as described above. Transgenic progeny or inbred lines are studied and treated with potential pharmaceutical agents to obtain information on treatment of a human disease.

Alternatively, a mammal inbred to overexpress KAP, e.g., by secreting KAP in its milk, may also serve as a convenient source of that protein (Janne, J. et al. (1998) Biotechnol. Annu. Rev. 4:55-74). **THERAPEUTICS**

Chemical and structural similarity, e.g., in the context of sequences and motifs, exists

5 between regions of KAP and kinases and phosphatases. In addition, examples of tissues expressing
KAP can be found in Table 6. Therefore, KAP appears to play a role in cardiovascular diseases,
immune system disorders, neurological disorders, disorders affecting growth and development, lipid
disorders, cell proliferative disorders, and cancers. In the treatment of disorders associated with
increased KAP expression or activity, it is desirable to decrease the expression or activity of KAP.

10 In the treatment of disorders associated with decreased KAP expression or activity, it is desirable to
increase the expression or activity of KAP.

Therefore, in one embodiment, KAP or a fragment or derivative thereof may be administered to a subject to treat or prevent a disorder associated with decreased expression or activity of KAP. Examples of such disorders include, but are not limited to, a cardiovascular 15 disorder such as arteriovenous fistula, atherosclerosis, hypertension, vasculitis, Raynaud's disease, aneurysms, arterial dissections, varicose veins, thrombophlebitis and phlebothrombosis, vascular tumors, and complications of thrombolysis, balloon angioplasty, vascular replacement, and coronary artery bypass graft surgery, congestive heart failure, ischemic heart disease, angina pectoris, myocardial infarction, hypertensive heart disease, degenerative valvular heart disease, calcific aortic 20 valve stenosis, congenitally bicuspid aortic valve, mitral annular calcification, mitral valve prolapse, rheumatic fever and rheumatic heart disease, infective endocarditis, nonbacterial thrombotic endocarditis, endocarditis of systemic lupus erythematosus, carcinoid heart disease, cardiomyopathy, myocarditis, pericarditis, neoplastic heart disease, congenital heart disease, and complications of cardiac transplantation, congenital lung anomalies, atelectasis, pulmonary 25 congestion and edema, pulmonary embolism, pulmonary hemorrhage, pulmonary infarction, pulmonary hypertension, vascular sclerosis, obstructive pulmonary disease, restrictive pulmonary disease, chronic obstructive pulmonary disease, emphysema, chronic bronchitis, bronchial asthma, bronchiectasis, bacterial pneumonia, viral and mycoplasmal pneumonia, lung abscess, pulmonary tuberculosis, diffuse interstitial diseases, pneumoconioses, sarcoidosis, idiopathic pulmonary 30 fibrosis, desquamative interstitial pneumonitis, hypersensitivity pneumonitis, pulmonary eosinophilia bronchiolitis obliterans-organizing pneumonia, diffuse pulmonary hemorrhage syndromes, Goodpasture's syndromes, idiopathic pulmonary hemosiderosis, pulmonary involvement in collagen-vascular disorders, pulmonary alveolar proteinosis, lung tumors, inflammatory and

noninflammatory pleural effusions, pneumothorax, pleural tumors, drug-induced lung disease, 35 radiation-induced lung disease, and complications of lung transplantation; an immune disorder such

as acquired immunodeficiency syndrome (AIDS), Addison's disease, adult respiratory distress syndrome, allergies, ankylosing spondylitis, amyloidosis, anemia, asthma, atherosclerosis, autoimmune hemolytic anemia, autoimmune thyroiditis, autoimmune polyendocrinopathycandidiasis-ectodermal dystrophy (APECED), bronchitis, cholecystitis, contact dermatitis, Crohn's 5 disease, atopic dermatitis, dermatomyositis, diabetes mellitus, emphysema, episodic lymphopenia with lymphocytotoxins, erythroblastosis fetalis, erythema nodosum, atrophic gastritis, glomerulonephritis, Goodpasture's syndrome, gout, Graves' disease, Hashimoto's thyroiditis, hypereosinophilia, irritable bowel syndrome, multiple sclerosis, myasthenia gravis, myocardial or pericardial inflammation, osteoarthritis, osteoporosis, pancreatitis, polymyositis, psoriasis, Reiter's 10 syndrome, rheumatoid arthritis, scleroderma, Sjögren's syndrome, systemic anaphylaxis, systemic lupus erythematosus, systemic sclerosis, thrombocytopenic purpura, ulcerative colitis, uveitis, Werner syndrome, complications of cancer, hemodialysis, and extracorporeal circulation, viral, bacterial, fungal, parasitic, protozoal, and helminthic infections, and trauma; a neurological disorder such as epilepsy, ischemic cerebrovascular disease, stroke, cerebral neoplasms, Alzheimer's 15 disease, Pick's disease, Huntington's disease, dementia, Parkinson's disease and other extrapyramidal disorders, amyotrophic lateral sclerosis and other motor neuron disorders, progressive neural muscular atrophy, retinitis pigmentosa, hereditary ataxias, multiple sclerosis and other demyelinating diseases, bacterial and viral meningitis, brain abscess, subdural empyema, epidural abscess, suppurative intracranial thrombophlebitis, myelitis and radiculitis, viral central 20 nervous system disease, prion diseases including kuru, Creutzfeldt-Jakob disease, and Gerstmann-Straussler-Scheinker syndrome, fatal familial insomnia, nutritional and metabolic diseases of the nervous system, neurofibromatosis, tuberous sclerosis, cerebelloretinal hemangioblastomatosis, encephalotrigeminal syndrome, mental retardation and other developmental disorders of the central nervous system including Down syndrome, cerebral palsy, neuroskeletal disorders, autonomic 25 nervous system disorders, cranial nerve disorders, spinal cord diseases, muscular dystrophy and other neuromuscular disorders, peripheral nervous system disorders, dermatomyositis and polymyositis, inherited, metabolic, endocrine, and toxic myopathies, myasthenia gravis, periodic paralysis, mental disorders including mood, anxiety, and schizophrenic disorders, seasonal affective disorder (SAD), akathesia, amnesia, catatonia, diabetic neuropathy, tardive dyskinesia, dystonias, 30 paranoid psychoses, postherpetic neuralgia, Tourette's disorder, progressive supranuclear palsy, corticobasal degeneration, and familial frontotemporal dementia; a growth and developmental disorder such as actinic keratosis, arteriosclerosis, atherosclerosis, bursitis, cirrhosis, hepatitis, mixed connective tissue disease (MCTD), myelofibrosis, paroxysmal nocturnal hemoglobinuria, polycythemia vera, psoriasis, primary thrombocythemia, renal tubular acidosis, anemia, Cushing's 35 syndrome, achondroplastic dwarfism, Duchenne and Becker muscular dystrophy, epilepsy, gonadal

dysgenesis, WAGR syndrome (Wilms' tumor, aniridia, genitourinary abnormalities, and mental retardation), Smith-Magenis syndrome, myelodysplastic syndrome, hereditary mucoepithelial dysplasia, hereditary keratodermas, hereditary neuropathies such as Charcot-Marie-Tooth disease and neurofibromatosis, hypothyroidism, hydrocephalus, seizure disorders such as Syndenham's 5 chorea and cerebral palsy, spina bifida, anencephaly, craniorachischisis, congenital glaucoma, cataract, and sensorineural hearing loss; a lipid disorder such as fatty liver, cholestasis, primary biliary cirrhosis, carnitine deficiency, carnitine palmitoyltransferase deficiency, myoadenylate deaminase deficiency, hypertriglyceridemia, lipid storage disorders such Fabry's disease, Gaucher's disease, Niemann-Pick's disease, metachromatic leukodystrophy, adrenoleukodystrophy, GM, 10 gangliosidosis, and ceroid lipofuscinosis, abetalipoproteinemia, Tangier disease, hyperlipoproteinemia, diabetes mellitus, lipodystrophy, lipomatoses, acute panniculitis, disseminated fat necrosis, adiposis dolorosa, lipoid adrenal hyperplasia, minimal change disease, lipomas, atherosclerosis, hypercholesterolemia, hypercholesterolemia with hypertriglyceridemia, primary hypoalphalipoproteinemia, hypothyroidism, renal disease, liver disease, lecithin:cholesterol 15 acyltransferase deficiency, cerebrotendinous xanthomatosis, sitosterolemia, hypocholesterolemia, Tay-Sachs disease, Sandhoff's disease, hyperlipidemia, hyperlipemia, lipid myopathies, and obesity; and a cell proliferative disorder such as actinic keratosis, arteriosclerosis, atherosclerosis, bursitis, cirrhosis, hepatitis, mixed connective tissue disease (MCTD), myelofibrosis, paroxysmal nocturnal hemoglobinuria, polycythemia vera, psoriasis, primary thrombocythemia, and cancers 20 including adenocarcinoma, leukemia, lymphoma, melanoma, myeloma, sarcoma, teratocarcinoma, and, in particular, cancers of the adrenal gland, bladder, bone, bone marrow, brain, breast, cervix, gall bladder, ganglia, gastrointestinal tract, heart, kidney, liver, lung, muscle, ovary, pancreas, parathyroid, penis, prostate, salivary glands, skin, spleen, testis, thymus, thyroid, uterus, leukemias

In another embodiment, a vector capable of expressing KAP or a fragment or derivative thereof may be administered to a subject to treat or prevent a disorder associated with decreased expression or activity of KAP including, but not limited to, those described above.

such as multiple myeloma, and lymphomas such as Hodgkin's disease..

In a further embodiment, a composition comprising a substantially purified KAP in conjunction with a suitable pharmaceutical carrier may be administered to a subject to treat or 30 prevent a disorder associated with decreased expression or activity of KAP including, but not limited to, those provided above.

In still another embodiment, an agonist which modulates the activity of KAP may be administered to a subject to treat or prevent a disorder associated with decreased expression or activity of KAP including, but not limited to, those listed above.

In a further embodiment, an antagonist of KAP may be administered to a subject to treat or

prevent a disorder associated with increased expression or activity of KAP. Examples of such disorders include, but are not limited to, those cardiovascular diseases, immune system disorders, neurological disorders, disorders affecting growth and development, lipid disorders, cell proliferative disorders, and cancers described above. In one aspect, an antibody which specifically binds KAP may be used directly as an antagonist or indirectly as a targeting or delivery mechanism for bringing a pharmaceutical agent to cells or tissues which express KAP.

In an additional embodiment, a vector expressing the complement of the polynucleotide encoding KAP may be administered to a subject to treat or prevent a disorder associated with increased expression or activity of KAP including, but not limited to, those described above.

In other embodiments, any of the proteins, antagonists, antibodies, agonists, complementary sequences, or vectors of the invention may be administered in combination with other appropriate therapeutic agents. Selection of the appropriate agents for use in combination therapy may be made by one of ordinary skill in the art, according to conventional pharmaceutical principles. The combination of therapeutic agents may act synergistically to effect the treatment or prevention of the various disorders described above. Using this approach, one may be able to achieve therapeutic efficacy with lower dosages of each agent, thus reducing the potential for adverse side effects.

An antagonist of KAP may be produced using methods which are generally known in the art. In particular, purified KAP may be used to produce antibodies or to screen libraries of pharmaceutical agents to identify those which specifically bind KAP. Antibodies to KAP may also 20 be generated using methods that are well known in the art. Such antibodies may include, but are not limited to, polyclonal, monoclonal, chimeric, and single chain antibodies, Fab fragments, and fragments produced by a Fab expression library. Neutralizing antibodies (i.e., those which inhibit dimer formation) are generally preferred for therapeutic use.

For the production of antibodies, various hosts including goats, rabbits, rats, mice, humans, 25 and others may be immunized by injection with KAP or with any fragment or oligopeptide thereof which has immunogenic properties. Depending on the host species, various adjuvants may be used to increase immunological response. Such adjuvants include, but are not limited to, Freund's, mineral gels such as aluminum hydroxide, and surface active substances such as lysolecithin, pluronic polyols, polyanions, peptides, oil emulsions, KLH, and dinitrophenol. Among adjuvants 30 used in humans, BCG (bacilli Calmette-Guerin) and Corynebacterium parvum are especially preferable.

It is preferred that the oligopeptides, peptides, or fragments used to induce antibodies to KAP have an amino acid sequence consisting of at least about 5 amino acids, and generally will consist of at least about 10 amino acids. It is also preferable that these oligopeptides, peptides, or fragments are identical to a portion of the amino acid sequence of the natural protein. Short

stretches of KAP amino acids may be fused with those of another protein, such as KLH, and antibodies to the chimeric molecule may be produced.

Monoclonal antibodies to KAP may be prepared using any technique which provides for the production of antibody molecules by continuous cell lines in culture. These include, but are not 5 limited to, the hybridoma technique, the human B-cell hybridoma technique, and the EBV-hybridoma technique. (See, e.g., Kohler, G. et al. (1975) Nature 256:495-497; Kozbor, D. et al. (1985) J. Immunol. Methods 81:31-42; Cote, R.J. et al. (1983) Proc. Natl. Acad. Sci. USA 80:2026-2030; and Cole, S.P. et al. (1984) Mol. Cell Biol. 62:109-120.)

In addition, techniques developed for the production of "chimeric antibodies," such as the 10 splicing of mouse antibody genes to human antibody genes to obtain a molecule with appropriate antigen specificity and biological activity, can be used. (See, e.g., Morrison, S.L. et al. (1984) Proc. Natl. Acad. Sci. USA 81:6851-6855; Neuberger, M.S. et al. (1984) Nature 312:604-608; and Takeda, S. et al. (1985) Nature 314:452-454.) Alternatively, techniques described for the production of single chain antibodies may be adapted, using methods known in the art, to produce 15 KAP-specific single chain antibodies. Antibodies with related specificity, but of distinct idiotypic composition, may be generated by chain shuffling from random combinatorial immunoglobulin libraries. (See, e.g., Burton, D.R. (1991) Proc. Natl. Acad. Sci. USA 88:10134-10137.)

Antibodies may also be produced by inducing in vivo production in the lymphocyte population or by screening immunoglobulin libraries or panels of highly specific binding reagents as 20 disclosed in the literature. (See, e.g., Orlandi, R. et al. (1989) Proc. Natl. Acad. Sci. USA 86:3833-3837; Winter, G. et al. (1991) Nature 349:293-299.)

Antibody fragments which contain specific binding sites for KAP may also be generated. For example, such fragments include, but are not limited to, F(ab')₂ fragments produced by pepsin digestion of the antibody molecule and Fab fragments generated by reducing the disulfide bridges of 25 the F(ab')₂ fragments. Alternatively, Fab expression libraries may be constructed to allow rapid and easy identification of monoclonal Fab fragments with the desired specificity. (See, e.g., Huse, W.D. et al. (1989) Science 246:1275-1281.)

Various immunoassays may be used for screening to identify antibodies having the desired specificity. Numerous protocols for competitive binding or immunoradiometric assays using either 30 polyclonal or monoclonal antibodies with established specificities are well known in the art. Such immunoassays typically involve the measurement of complex formation between KAP and its specific antibody. A two site, monoclonal-based immunoassay utilizing monoclonal antibodies reactive to two non-interfering KAP epitopes is generally used, but a competitive binding assay may also be employed (Pound, supra).

Various methods such as Scatchard analysis in conjunction with radioimmunoassay

techniques may be used to assess the affinity of antibodies for KAP. Affinity is expressed as an association constant, K_a , which is defined as the molar concentration of KAP-antibody complex divided by the molar concentrations of free antigen and free antibody under equilibrium conditions. The K_a determined for a preparation of polyclonal antibodies, which are heterogeneous in their 5 affinities for multiple KAP epitopes, represents the average affinity, or avidity, of the antibodies for KAP. The K_a determined for a preparation of monoclonal antibodies, which are monospecific for a particular KAP epitope, represents a true measure of affinity. High-affinity antibody preparations with K_a ranging from about 10° to 10¹2 L/mole are preferred for use in immunoassays in which the KAP-antibody complex must withstand rigorous manipulations. Low-affinity antibody preparations 10 with K_a ranging from about 10° to 10¹ L/mole are preferred for use in immunopurification and similar procedures which ultimately require dissociation of KAP, preferably in active form, from the antibody (Catty, D. (1988) Antibodies, Volume I: A Practical Approach, IRL Press, Washington DC; Liddell, J.E. and A. Cryer (1991) A Practical Guide to Monoclonal Antibodies, John Wiley & Sons, New York NY).

The titer and avidity of polyclonal antibody preparations may be further evaluated to determine the quality and suitability of such preparations for certain downstream applications. For example, a polyclonal antibody preparation containing at least 1-2 mg specific antibody/ml, preferably 5-10 mg specific antibody/ml, is generally employed in procedures requiring precipitation of KAP-antibody complexes. Procedures for evaluating antibody specificity, titer, and avidity, and guidelines for antibody quality and usage in various applications, are generally available. (See, e.g., Catty, supra, and Coligan et al. supra.)

In another embodiment of the invention, the polynucleotides encoding KAP, or any fragment or complement thereof, may be used for therapeutic purposes. In one aspect, modifications of gene expression can be achieved by designing complementary sequences or antisense molecules (DNA, RNA, PNA, or modified oligonucleotides) to the coding or regulatory regions of the gene encoding KAP. Such technology is well known in the art, and antisense oligonucleotides or larger fragments can be designed from various locations along the coding or control regions of sequences encoding KAP. (See, e.g., Agrawal, S., ed. (1996) Antisense Therapeutics, Humana Press Inc., Totawa NJ.)

In therapeutic use, any gene delivery system suitable for introduction of the antisense sequences into appropriate target cells can be used. Antisense sequences can be delivered intracellularly in the form of an expression plasmid which, upon transcription, produces a sequence complementary to at least a portion of the cellular sequence encoding the target protein. (See, e.g., Slater, J.E. et al. (1998) J. Allergy Clin. Immunol. 102(3):469-475; and Scanlon, K.J. et al. (1995) 9(13):1288-1296.) Antisense sequences can also be introduced intracellularly through the use of

viral vectors, such as retrovirus and adeno-associated virus vectors. (See, e.g., Miller, A.D. (1990) Blood 76:271; Ausubel, supra; Uckert, W. and W. Walther (1994) Pharmacol. Ther. 63(3):323-347.) Other gene delivery mechanisms include liposome-derived systems, artificial viral envelopes, and other systems known in the art. (See, e.g., Rossi, J.J. (1995) Br. Med. Bull. 51(1):217-225; 5 Boado, R.J. et al. (1998) J. Pharm. Sci. 87(11):1308-1315; and Morris, M.C. et al. (1997) Nucleic Acids Res. 25(14):2730-2736.)

In another embodiment of the invention, polynucleotides encoding KAP may be used for somatic or germline gene therapy. Gene therapy may be performed to (i) correct a genetic deficiency (e.g., in the cases of severe combined immunodeficiency (SCID)-X1 disease 10 characterized by X-linked inheritance (Cavazzana-Calvo, M. et al. (2000) Science 288:669-672), severe combined immunodeficiency syndrome associated with an inherited adenosine deaminase (ADA) deficiency (Blaese, R.M. et al. (1995) Science 270:475-480; Bordignon, C. et al. (1995) Science 270:470-475), cystic fibrosis (Zabner, J. et al. (1993) Cell 75:207-216; Crystal, R.G. et al. (1995) Hum. Gene Therapy 6:643-666; Crystal, R.G. et al. (1995) Hum. Gene Therapy 6:667-703), 15 thalassamias, familial hypercholesterolemia, and hemophilia resulting from Factor VIII or Factor IX deficiencies (Crystal, R.G. (1995) Science 270:404-410; Verma, I.M. and N. Somia (1997) Nature 389:239-242)), (ii) express a conditionally lethal gene product (e.g., in the case of cancers which result from unregulated cell proliferation), or (iii) express a protein which affords protection against intracellular parasites (e.g., against human retroviruses, such as human immunodeficiency virus 20 (HIV) (Baltimore, D. (1988) Nature 335:395-396; Poeschla, E. et al. (1996) Proc. Natl. Acad. Sci. USA 93:11395-11399), hepatitis B or C virus (HBV, HCV); fungal parasites, such as Candida albicans and Paracoccidioides brasiliensis; and protozoan parasites such as Plasmodium falciparum and Trypanosoma cruzi). In the case where a genetic deficiency in KAP expression or regulation causes disease, the expression of KAP from an appropriate population of transduced cells may 25 alleviate the clinical manifestations caused by the genetic deficiency.

In a further embodiment of the invention, diseases or disorders caused by deficiencies in KAP are treated by constructing mammalian expression vectors encoding KAP and introducing these vectors by mechanical means into KAP-deficient cells. Mechanical transfer technologies for use with cells in vivo or ex vitro include (i) direct DNA microinjection into individual cells, (ii) 30 ballistic gold particle delivery, (iii) liposome-mediated transfection, (iv) receptor-mediated gene transfer, and (v) the use of DNA transposons (Morgan, R.A. and W.F. Anderson (1993) Annu. Rev. Biochem. 62:191-217; Ivies; Z. (1997) Cell-91:501-510; Boulay; J-L. and H. Récipon (1998) Curr. Opin. Biotechnol. 9:445-450).

Expression vectors that may be effective for the expression of KAP include, but are not 35 limited to, the PCDNA 3.1, EPITAG, PRCCMV2, PREP, PVAX, PCR2-TOPOTA vectors

(Invitrogen, Carlsbad CA), PCMV-SCRIPT, PCMV-TAG, PEGSH/PERV (Stratagene, La Jolla CA), and PTET-OFF, PTET-ON, PTRE2, PTRE2-LUC, PTK-HYG (Clontech, Palo Alto CA). KAP may be expressed using (i) a constitutively active promoter, (e.g., from cytomegalovirus (CMV), Rous sarcoma virus (RSV), SV40 virus, thymidine kinase (TK), or β-actin genes), (ii) an 5 inducible promoter (e.g., the tetracycline-regulated promoter (Gossen, M. and H. Bujard (1992) Proc. Natl. Acad. Sci. USA 89:5547-5551; Gossen, M. et al. (1995) Science 268:1766-1769; Rossi, F.M.V. and H.M. Blau (1998) Curr. Opin. Biotechnol. 9:451-456), commercially available in the T-REX plasmid (Invitrogen)); the ecdysone-inducible promoter (available in the plasmids PVGRXR and PIND; Invitrogen); the FK506/rapamycin inducible promoter; or the RU486/mifepristone 10 inducible promoter (Rossi, F.M.V. and H.M. Blau, supra)), or (iii) a tissue-specific promoter or the native promoter of the endogenous gene encoding KAP from a normal individual.

Commercially available liposome transformation kits (e.g., the PERFECT LIPID TRANSFECTION KIT, available from Invitrogen) allow one with ordinary skill in the art to deliver polynucleotides to target cells in culture and require minimal effort to optimize experimental parameters. In the alternative, transformation is performed using the calcium phosphate method (Graham, F.L. and A.J. Eb (1973) Virology 52:456-467), or by electroporation (Neumann, E. et al. (1982) EMBO J. 1:841-845). The introduction of DNA to primary cells requires modification of these standardized mammalian transfection protocols.

In another embodiment of the invention, diseases or disorders caused by genetic defects 20 with respect to KAP expression are treated by constructing a retrovirus vector consisting of (i) the polynucleotide encoding KAP under the control of an independent promoter or the retrovirus long terminal repeat (LTR) promoter, (ii) appropriate RNA packaging signals, and (iii) a Rev-responsive element (RRE) along with additional retrovirus cis-acting RNA sequences and coding sequences required for efficient vector propagation. Retrovirus vectors (e.g., PFB and PFBNEO) are 25 commercially available (Stratagene) and are based on published data (Riviere, I. et al. (1995) Proc. Natl. Acad. Sci. USA 92:6733-6737), incorporated by reference herein. The vector is propagated in an appropriate vector producing cell line (VPCL) that expresses an envelope gene with a tropism for receptors on the target cells or a promiscuous envelope protein such as VSVg (Armentano, D. et al. (1987) J. Virol. 61:1647-1650; Bender, M.A. et al. (1987) J. Virol. 61:1639-1646; Adam, M.A. and 30 A.D. Miller (1988) J. Virol. 62:3802-3806; Dull, T. et al. (1998) J. Virol. 72:8463-8471; Zufferey, R. et al. (1998) J. Virol. 72:9873-9880). U.S. Patent No. 5,910,434 to Rigg ("Method for obtaining retrovirus packaging cell lines producing high transducing efficiency retroviral supernatant") discloses a method for obtaining retrovirus packaging cell lines and is hereby incorporated by reference. Propagation of retrovirus vectors, transduction of a population of cells (e.g., CD4+T-35 cells), and the return of transduced cells to a patient are procedures well known to persons skilled in

the art of gene therapy and have been well documented (Ranga, U. et al. (1997) J. Virol. 71:7020-7029; Bauer, G. et al. (1997) Blood 89:2259-2267; Bonyhadi, M.L. (1997) J. Virol. 71:4707-4716; Ranga, U. et al. (1998) Proc. Natl. Acad. Sci. USA 95:1201-1206; Su, L. (1997) Blood 89:2283-2290).

- In the alternative, an adenovirus-based gene therapy delivery system is used to deliver polynucleotides encoding KAP to cells which have one or more genetic abnormalities with respect to the expression of KAP. The construction and packaging of adenovirus-based vectors are well known to those with ordinary skill in the art. Replication defective adenovirus vectors have proven to be versatile for importing genes encoding immunoregulatory proteins into intact islets in the 10 pancreas (Csete, M.E. et al. (1995) Transplantation 27:263-268). Potentially useful adenoviral vectors are described in U.S. Patent No. 5,707,618 to Armentano ("Adenovirus vectors for gene therapy"), hereby incorporated by reference. For adenoviral vectors, see also Antinozzi, P.A. et al. (1999) Annu. Rev. Nutr. 19:511-544 and Verma, I.M. and N. Somia (1997) Nature 18:389:239-242, both incorporated by reference herein.
- 15 In another alternative, a herpes-based, gene therapy delivery system is used to deliver polynucleotides encoding KAP to target cells which have one or more genetic abnormalities with respect to the expression of KAP. The use of herpes simplex virus (HSV)-based vectors may be especially valuable for introducing KAP to cells of the central nervous system, for which HSV has a tropism. The construction and packaging of herpes-based vectors are well known to those with 20 ordinary skill in the art. A replication-competent herpes simplex virus (HSV) type 1-based vector has been used to deliver a reporter gene to the eyes of primates (Liu, X. et al. (1999) Exp. Eye Res. 169:385-395). The construction of a HSV-1 virus vector has also been disclosed in detail in U.S. Patent No. 5,804,413 to DeLuca ("Herpes simplex virus strains for gene transfer"), which is hereby incorporated by reference. U.S. Patent No. 5,804,413 teaches the use of recombinant HSV d92 25 which consists of a genome containing at least one exogenous gene to be transferred to a cell under the control of the appropriate promoter for purposes including human gene therapy. Also taught by this patent are the construction and use of recombinant HSV strains deleted for ICP4, ICP27 and ICP22. For HSV vectors, see also Goins, W.F. et al. (1999) J. Virol. 73:519-532 and Xu, H. et al. (1994) Dev. Biol. 163:152-161, hereby incorporated by reference. The manipulation of cloned 30 herpesvirus sequences, the generation of recombinant virus following the transfection of multiple plasmids containing different segments of the large herpesvirus genomes, the growth and propagation of herpesvirus, and the infection of cells with herpesvirus are techniques well known to those of ordinary skill in the art.

In another alternative, an alphavirus (positive, single-stranded RNA virus) vector is used to 35 deliver polynucleotides encoding KAP to target cells. The biology of the prototypic alphavirus,

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Semliki Forest Virus (SFV), has been studied extensively and gene transfer vectors have been based on the SFV genome (Garoff, H. and K.-J. Li (1998) Curr. Opin. Biotechnol. 9:464-469). During alphavirus RNA replication, a subgenomic RNA is generated that normally encodes the viral capsid proteins. This subgenomic RNA replicates to higher levels than the full length genomic RNA, 5 resulting in the overproduction of capsid proteins relative to the viral proteins with enzymatic activity (e.g., protease and polymerase). Similarly, inserting the coding sequence for KAP into the alphavirus genome in place of the capsid-coding region results in the production of a large number of KAP-coding RNAs and the synthesis of high levels of KAP in vector transduced cells. While alphavirus infection is typically associated with cell lysis within a few days, the ability to establish a 10 persistent infection in hamster normal kidney cells (BHK-21) with a variant of Sindbis virus (SIN) indicates that the lytic replication of alphaviruses can be altered to suit the needs of the gene therapy application (Dryga, S.A. et al. (1997) Virology 228:74-83). The wide host range of alphaviruses will allow the introduction of KAP into a variety of cell types. The specific transduction of a subset of cells in a population may require the sorting of cells prior to transduction. The methods of 15 manipulating infectious cDNA clones of alphaviruses, performing alphavirus cDNA and RNA transfections, and performing alphavirus infections, are well known to those with ordinary skill in the art.

Oligonucleotides derived from the transcription initiation site, e.g., between about positions
-10 and +10 from the start site, may also be employed to inhibit gene expression. Similarly,
20 inhibition can be achieved using triple helix base-pairing methodology. Triple helix pairing is
useful because it causes inhibition of the ability of the double helix to open sufficiently for the
binding of polymerases, transcription factors, or regulatory molecules. Recent therapeutic advances
using triplex DNA have been described in the literature. (See, e.g., Gee, J.E. et al. (1994) in Huber,
B.E. and B.I. Carr, Molecular and Immunologic Approaches, Futura Publishing, Mt. Kisco NY, pp.
25 163-177.) A complementary sequence or antisense molecule may also be designed to block
translation of mRNA by preventing the transcript from binding to ribosomes.

Ribozymes, enzymatic RNA molecules, may also be used to catalyze the specific cleavage of RNA. The mechanism of ribozyme action involves sequence-specific hybridization of the ribozyme molecule to complementary target RNA, followed by endonucleolytic cleavage. For 30 example, engineered hammerhead motif ribozyme molecules may specifically and efficiently catalyze endonucleolytic cleavage of sequences encoding KAP.

Specific ribozyme cleavage sites within any potential RNA target are initially identified by scanning the target molecule for ribozyme cleavage sites, including the following sequences: GUA, GUU, and GUC. Once identified, short RNA sequences of between 15 and 20 ribonucleotides, 35 corresponding to the region of the target gene containing the cleavage site, may be evaluated for

secondary structural features which may render the oligonucleotide inoperable. The suitability of candidate targets may also be evaluated by testing accessibility to hybridization with complementary oligonucleotides using ribonuclease protection assays.

Complementary ribonucleic acid molecules and ribozymes of the invention may be prepared 5 by any method known in the art for the synthesis of nucleic acid molecules. These include techniques for chemically synthesizing oligonucleotides such as solid phase phosphoramidite chemical synthesis. Alternatively, RNA molecules may be generated by in vitro and in vivo transcription of DNA sequences encoding KAP. Such DNA sequences may be incorporated into a wide variety of vectors with suitable RNA polymerase promoters such as T7 or SP6. Alternatively, these cDNA constructs that synthesize complementary RNA, constitutively or inducibly, can be introduced into cell lines, cells, or tissues.

RNA molecules may be modified to increase intracellular stability and half-life. Possible modifications include, but are not limited to, the addition of flanking sequences at the 5' and/or 3' ends of the molecule, or the use of phosphorothioate or 2'O-methyl rather than phosphodiesterase 15 linkages within the backbone of the molecule. This concept is inherent in the production of PNAs and can be extended in all of these molecules by the inclusion of nontraditional bases such as inosine, queosine, and wybutosine, as well as acetyl-, methyl-, thio-, and similarly modified forms of adenine, cytidine, guanine, thymine, and uridine which are not as easily recognized by endogenous endonucleases.

An additional embodiment of the invention encompasses a method for screening for a compound which is effective in altering expression of a polynucleotide encoding KAP. Compounds which may be effective in altering expression of a specific polynucleotide may include, but are not limited to, oligonucleotides, antisense oligonucleotides, triple helix-forming oligonucleotides, transcription factors and other polypeptide transcriptional regulators, and non-macromolecular compounds may alter polynucleotide expression by acting as either inhibitors or promoters of polynucleotide expression. Thus, in the treatment of disorders associated with increased KAP expression or activity, a compound which specifically inhibits expression of the polynucleotide encoding KAP may be therapeutically useful, and in the treatment of disorders associated with 30 decreased KAP expression or activity, a compound which specifically promotes expression of the polynucleotide encoding KAP may be therapeutically useful.

At least one, and up to a plurality, of test compounds may be screened for effectiveness in altering expression of a specific polynucleotide. A test compound may be obtained by any method commonly known in the art, including chemical modification of a compound known to be effective 35 in altering polynucleotide expression; selection from an existing, commercially-available or

proprietary library of naturally-occurring or non-natural chemical compounds; rational design of a compound based on chemical and/or structural properties of the target polynucleotide; and selection from a library of chemical compounds created combinatorially or randomly. A sample comprising a polynucleotide encoding KAP is exposed to at least one test compound thus obtained. The sample 5 may comprise, for example, an intact or permeabilized cell, or an in vitro cell-free or reconstituted biochemical system. Alterations in the expression of a polynucleotide encoding KAP are assayed by any method commonly known in the art. Typically, the expression of a specific nucleotide is detected by hybridization with a probe having a nucleotide sequence complementary to the sequence of the polynucleotide encoding KAP. The amount of hybridization may be quantified, 10 thus forming the basis for a comparison of the expression of the polynucleotide both with and without exposure to one or more test compounds. Detection of a change in the expression of a polynucleotide exposed to a test compound indicates that the test compound is effective in altering the expression of the polynucleotide. A screen for a compound effective in altering expression of a specific polynucleotide can be carried out, for example, using a Schizosaccharomyces pombe gene 15 expression system (Atkins, D. et al. (1999) U.S. Patent No. 5,932,435; Arndt, G.M. et al. (2000) Nucleic Acids Res. 28:E15) or a human cell line such as HeLa cell (Clarke, M.L. et al. (2000) Biochem. Biophys. Res. Commun. 268:8-13). A particular embodiment of the present invention involves screening a combinatorial library of oligonucleotides (such as deoxyribonucleotides, ribonucleotides, peptide nucleic acids, and modified oligonucleotides) for antisense activity against 20 a specific polynucleotide sequence (Bruice, T.W. et al. (1997) U.S. Patent No. 5,686,242; Bruice, T.W. et al. (2000) U.S. Patent No. 6,022,691).

Many methods for introducing vectors into cells or tissues are available and equally suitable for use in vivo, in vitro, and ex vivo. For ex vivo therapy, vectors may be introduced into stem cells taken from the patient and clonally propagated for autologous transplant back into that same patient.

25 Delivery by transfection, by liposome injections, or by polycationic amino polymers may be achieved using methods which are well known in the art. (See, e.g., Goldman, C.K. et al. (1997)

Nat. Biotechnol. 15:462-466.)

Any of the therapeutic methods described above may be applied to any subject in need of such therapy, including, for example, mammals such as humans, dogs, cats, cows, horses, rabbits, 30 and monkeys.

An additional embodiment of the invention relates to the administration of a composition which generally comprises an active ingredient formulated with a pharmaceutically acceptable excipient. Excipients may include, for example, sugars, starches, celluloses, gums, and proteins. Various formulations are commonly known and are thoroughly discussed in the latest edition of Remington's Pharmaceutical Sciences (Maack Publishing, Easton PA). Such compositions may

consist of KAP, antibodies to KAP, and mimetics, agonists, antagonists, or inhibitors of KAP.

The compositions utilized in this invention may be administered by any number of routes including, but not limited to, oral, intravenous, intramuscular, intra-arterial, intramedullary, intrathecal, intraventricular, pulmonary, transdermal, subcutaneous, intraperitoneal, intranasal, 5 enteral, topical, sublingual, or rectal means.

Compositions for pulmonary administration may be prepared in liquid or dry powder form. These compositions are generally aerosolized immediately prior to inhalation by the patient. In the case of small molecules (e.g. traditional low molecular weight organic drugs), aerosol delivery of fast-acting formulations is well-known in the art. In the case of macromolecules (e.g. larger 10 peptides and proteins), recent developments in the field of pulmonary delivery via the alveolar region of the lung have enabled the practical delivery of drugs such as insulin to blood circulation (see, e.g., Patton, J.S. et al., U.S. Patent No. 5,997,848). Pulmonary delivery has the advantage of administration without needle injection, and obviates the need for potentially toxic penetration enhancers.

15 Compositions suitable for use in the invention include compositions wherein the active ingredients are contained in an effective amount to achieve the intended purpose. The determination of an effective dose is well within the capability of those skilled in the art.

Specialized forms of compositions may be prepared for direct intracellular delivery of macromolecules comprising KAP or fragments thereof. For example, liposome preparations 20 containing a cell-impermeable macromolecule may promote cell fusion and intracellular delivery of the macromolecule. Alternatively, KAP or a fragment thereof may be joined to a short cationic N-terminal portion from the HIV Tat-1 protein. Fusion proteins thus generated have been found to transduce into the cells of all tissues, including the brain, in a mouse model system (Schwarze, S.R. et al. (1999) Science 285:1569-1572).

For any compound, the therapeutically effective dose can be estimated initially either in cell culture assays, e.g., of neoplastic cells, or in animal models such as mice, rats, rabbits, dogs, monkeys, or pigs. An animal model may also be used to determine the appropriate concentration range and route of administration. Such information can then be used to determine useful doses and routes for administration in humans.

A therapeutically effective dose refers to that amount of active ingredient, for example KAP or fragments thereof, antibodies of KAP, and agonists, antagonists or inhibitors of KAP, which ameliorates the symptoms or condition. Therapeutic efficacy and toxicity may be determined by standard pharmaceutical procedures in cell cultures or with experimental animals, such as by calculating the ED₅₀ (the dose therapeutically effective in 50% of the population) or LD₅₀ (the dose 35 lethal to 50% of the population) statistics. The dose ratio of toxic to therapeutic effects is the

therapeutic index, which can be expressed as the LD₅₀/ED₅₀ ratio. Compositions which exhibit large therapeutic indices are preferred. The data obtained from cell culture assays and animal studies are used to formulate a range of dosage for human use. The dosage contained in such compositions is preferably within a range of circulating concentrations that includes the ED₅₀ with little or no toxicity. The dosage varies within this range depending upon the dosage form employed, the sensitivity of the patient, and the route of administration.

The exact dosage will be determined by the practitioner, in light of factors related to the subject requiring treatment. Dosage and administration are adjusted to provide sufficient levels of the active moiety or to maintain the desired effect. Factors which may be taken into account include 10 the severity of the disease state, the general health of the subject, the age, weight, and gender of the subject, time and frequency of administration, drug combination(s), reaction sensitivities, and response to therapy. Long-acting compositions may be administered every 3 to 4 days, every week, or biweekly depending on the half-life and clearance rate of the particular formulation.

Normal dosage amounts may vary from about $0.1 \mu g$ to $100,000 \mu g$, up to a total dose of 15 about 1 gram, depending upon the route of administration. Guidance as to particular dosages and methods of delivery is provided in the literature and generally available to practitioners in the art. Those skilled in the art will employ different formulations for nucleotides than for proteins or their inhibitors. Similarly, delivery of polynucleotides or polypeptides will be specific to particular cells, conditions, locations, etc.

20 DIAGNOSTICS

In another embodiment, antibodies which specifically bind KAP may be used for the diagnosis of disorders characterized by expression of KAP, or in assays to monitor patients being treated with KAP or agonists, antagonists, or inhibitors of KAP. Antibodies useful for diagnostic purposes may be prepared in the same manner as described above for therapeutics. Diagnostic 25 assays for KAP include methods which utilize the antibody and a label to detect KAP in human body fluids or in extracts of cells or tissues. The antibodies may be used with or without modification, and may be labeled by covalent or non-covalent attachment of a reporter molecule. A wide variety of reporter molecules, several of which are described above, are known in the art and may be used.

A variety of protocols for measuring KAP, including ELISAs, RIAs, and FACS, are known in the art and provide a basis for diagnosing altered or abnormal levels of KAP expression. Normal or standard values for KAP expression are established by combining body fluids or cell extracts taken from normal mammalian subjects, for example, human subjects, with antibodies to KAP under conditions suitable for complex formation. The amount of standard complex formation may 35 be quantitated by various methods, such as photometric means. Quantities of KAP expressed in

subject, control, and disease samples from biopsied tissues are compared with the standard values. Deviation between standard and subject values establishes the parameters for diagnosing disease.

In another embodiment of the invention, the polynucleotides encoding KAP may be used for diagnostic purposes. The polynucleotides which may be used include oligonucleotide sequences, 5 complementary RNA and DNA molecules, and PNAs. The polynucleotides may be used to detect and quantify gene expression in biopsied tissues in which expression of KAP may be correlated with disease. The diagnostic assay may be used to determine absence, presence, and excess expression of KAP, and to monitor regulation of KAP levels during therapeutic intervention.

In one aspect, hybridization with PCR probes which are capable of detecting polynucleotide sequences, including genomic sequences, encoding KAP or closely related molecules may be used to identify nucleic acid sequences which encode KAP. The specificity of the probe, whether it is made from a highly specific region, e.g., the 5' regulatory region, or from a less specific region, e.g., a conserved motif, and the stringency of the hybridization or amplification will determine whether the probe identifies only naturally occurring sequences encoding KAP, allelic variants, or related 15 sequences.

Probes may also be used for the detection of related sequences, and may have at least 50% sequence identity to any of the KAP encoding sequences. The hybridization probes of the subject invention may be DNA or RNA and may be derived from the sequence of SEQ ID NO:21-40 or from genomic sequences including promoters, enhancers, and introns of the KAP gene.

Means for producing specific hybridization probes for DNAs encoding KAP include the cloning of polynucleotide sequences encoding KAP or KAP derivatives into vectors for the production of mRNA probes. Such vectors are known in the art, are commercially available, and may be used to synthesize RNA probes in vitro by means of the addition of the appropriate RNA polymerases and the appropriate labeled nucleotides. Hybridization probes may be labeled by a variety of reporter groups, for example, by radionuclides such as ³²P or ³⁵S, or by enzymatic labels, such as alkaline phosphatase coupled to the probe via avidin/biotin coupling systems, and the like.

Polynucleotide sequences encoding KAP may be used for the diagnosis of disorders associated with expression of KAP. Examples of such disorders include, but are not limited to, a cardiovascular disorder such as arteriovenous fistula, atherosclerosis, hypertension, vasculitis, 30 Raynaud's disease, aneurysms, arterial dissections, varicose veins, thrombophlebitis and phlebothrombosis, vascular tumors, and complications of thrombolysis, balloon angioplasty, vascular replacement, and coronary artery bypass graft surgery, congestive heart failure, ischemic heart disease, angina pectoris, myocardial infarction, hypertensive heart disease, degenerative valvular heart disease, calcific aortic valve stenosis, congenitally bicuspid aortic valve, mitral 35 annular calcification, mitral valve prolapse, rheumatic fever and rheumatic heart disease, infective

endocarditis, nonbacterial thrombotic endocarditis, endocarditis of systemic lupus erythematosus, carcinoid heart disease, cardiomyopathy, myocarditis, pericarditis, neoplastic heart disease, congenital heart disease, and complications of cardiac transplantation, congenital lung anomalies, atelectasis, pulmonary congestion and edema, pulmonary embolism, pulmonary hemorrhage, 5 pulmonary infarction, pulmonary hypertension, vascular sclerosis, obstructive pulmonary disease, restrictive pulmonary disease, chronic obstructive pulmonary disease, emphysema, chronic bronchitis, bronchial asthma, bronchiectasis, bacterial pneumonia, viral and mycoplasmal pneumonia, lung abscess, pulmonary tuberculosis, diffuse interstitial diseases, pneumoconioses, sarcoidosis, idiopathic pulmonary fibrosis, desquamative interstitial pneumonitis, hypersensitivity 10 pneumonitis, pulmonary eosinophilia bronchiolitis obliterans-organizing pneumonia, diffuse pulmonary hemorrhage syndromes, Goodpasture's syndromes, idiopathic pulmonary hemosiderosis, pulmonary involvement in collagen-vascular disorders, pulmonary alveolar proteinosis, lung tumors, inflammatory and noninflammatory pleural effusions, pneumothorax, pleural tumors, drug-induced lung disease, radiation-induced lung disease, and complications of lung transplantation; an immune 15 disorder such as acquired immunodeficiency syndrome (AIDS), Addison's disease, adult respiratory distress syndrome, allergies, ankylosing spondylitis, amyloidosis, anemia, asthma, atherosclerosis, autoimmune hemolytic anemia, autoimmune thyroiditis, autoimmune polyendocrinopathycandidiasis-ectodermal dystrophy (APECED), bronchitis, cholecystitis, contact dermatitis, Crohn's disease, atopic dermatitis, dermatomyositis, diabetes mellitus, emphysema, episodic lymphopenia 20 with lymphocytotoxins, erythroblastosis fetalis, erythema nodosum, atrophic gastritis, glomerulonephritis, Goodpasture's syndrome, gout, Graves' disease, Hashimoto's thyroiditis, hypereosinophilia, irritable bowel syndrome, multiple sclerosis, myasthenia gravis, myocardial or pericardial inflammation, osteoarthritis, osteoporosis, pancreatitis, polymyositis, psoriasis, Reiter's syndrome, rheumatoid arthritis, scleroderma, Sjögren's syndrome, systemic anaphylaxis, systemic 25 lupus erythematosus, systemic sclerosis, thrombocytopenic purpura, ulcerative colitis, uveitis, Werner syndrome, complications of cancer, hemodialysis, and extracorporeal circulation, viral, bacterial, fungal, parasitic, protozoal, and helminthic infections, and trauma; a neurological disorder such as epilepsy, ischemic cerebrovascular disease, stroke, cerebral neoplasms, Alzheimer's disease, Pick's disease, Huntington's disease, dementia, Parkinson's disease and other 30 extrapyramidal disorders, amyotrophic lateral sclerosis and other motor neuron disorders, progressive neural muscular atrophy, retinitis pigmentosa, hereditary ataxias, multiple sclerosis and other demyelinating diseases, bacterial and viral meningitis, brain abscess, subdural empyema, epidural abscess, suppurative intracranial thrombophlebitis, myelitis and radiculitis, viral central nervous system disease, prion diseases including kuru, Creutzfeldt-Jakob disease, and Gerstmann-35 Straussler-Scheinker syndrome, fatal familial insomnia, nutritional and metabolic diseases of the

nervous system, neurofibromatosis, tuberous sclerosis, cerebelloretinal hemangioblastomatosis, encephalotrigeminal syndrome, mental retardation and other developmental disorders of the central nervous system including Down syndrome, cerebral palsy, neuroskeletal disorders, autonomic nervous system disorders, cranial nerve disorders, spinal cord diseases, muscular dystrophy and

- 5 other neuromuscular disorders, peripheral nervous system disorders, dermatomyositis and polymyositis, inherited, metabolic, endocrine, and toxic myopathies, myasthenia gravis, periodic paralysis, mental disorders including mood, anxiety, and schizophrenic disorders, seasonal affective disorder (SAD), akathesia, amnesia, catatonia, diabetic neuropathy, tardive dyskinesia, dystonias, paranoid psychoses, postherpetic neuralgia, Tourette's disorder, progressive supranuclear palsy,
- 10 corticobasal degeneration, and familial frontotemporal dementia; a growth and developmental disorder such as actinic keratosis, arteriosclerosis, atherosclerosis, bursitis, cirrhosis, hepatitis, mixed connective tissue disease (MCTD), myelofibrosis, paroxysmal nocturnal hemoglobinuria, polycythemia vera, psoriasis, primary thrombocythemia, renal tubular acidosis, anemia, Cushing's syndrome, achondroplastic dwarfism, Duchenne and Becker muscular dystrophy, epilepsy, gonadal
- 15 dysgenesis, WAGR syndrome (Wilms' tumor, aniridia, genitourinary abnormalities, and mental retardation), Smith-Magenis syndrome, myelodysplastic syndrome, hereditary mucoepithelial dysplasia, hereditary keratodermas, hereditary neuropathies such as Charcot-Marie-Tooth disease and neurofibromatosis, hypothyroidism, hydrocephalus, seizure disorders such as Syndenham's chorea and cerebral palsy, spina bifida, anencephaly, craniorachischisis, congenital glaucoma,
- 20 cataract, and sensorineural hearing loss; a lipid disorder such as fatty liver, cholestasis, primary biliary cirrhosis, carnitine deficiency, carnitine palmitoyltransferase deficiency, myoadenylate deaminase deficiency, hypertriglyceridemia, lipid storage disorders such Fabry's disease, Gaucher's disease, Niemann-Pick's disease, metachromatic leukodystrophy, adrenoleukodystrophy, GM₂ gangliosidosis, and ceroid lipofuscinosis, abetalipoproteinemia, Tangier disease,
- 25 hyperlipoproteinemia, diabetes mellitus, lipodystrophy, lipomatoses, acute panniculitis, disseminated fat necrosis, adiposis dolorosa, lipoid adrenal hyperplasia, minimal change disease, lipomas, atherosclerosis, hypercholesterolemia, hypercholesterolemia with hypertriglyceridemia, primary hypoalphalipoproteinemia, hypothyroidism, renal disease, liver disease, lecithin:cholesterol acyltransferase deficiency, cerebrotendinous xanthomatosis, sitosterolemia, hypocholesterolemia,
- 30 Tay-Sachs disease, Sandhoff's disease, hyperlipidemia, hyperlipemia, lipid myopathies, and obesity; and a cell proliferative disorder such as actinic keratosis, arteriosclerosis, atherosclerosis, bursitis, cirrhosis, hepatitis, mixed connective tissue disease (MCTD), myelofibrosis, paroxysmal nocturnal hemoglobinuria, polycythemia vera, psoriasis, primary thrombocythemia, and cancers including adenocarcinoma, leukemia, lymphoma, melanoma, myeloma, sarcoma, teratocarcinoma, 35 and, in particular, cancers of the adrenal gland, bladder, bone, bone marrow, brain, breast, cervix,

gall bladder, ganglia, gastrointestinal tract, heart, kidney, liver, lung, muscle, ovary, pancreas, parathyroid, penis, prostate, salivary glands, skin, spleen, testis, thymus, thyroid, uterus, leukemias such as multiple myeloma, and lymphomas such as Hodgkin's disease. The polynucleotide sequences encoding KAP may be used in Southern or northern analysis, dot blot, or other membrane-based technologies; in PCR technologies; in dipstick, pin, and multiformat ELISA-like assays; and in microarrays utilizing fluids or tissues from patients to detect altered KAP expression. Such qualitative or quantitative methods are well known in the art.

In a particular aspect, the nucleotide sequences encoding KAP may be useful in assays that detect the presence of associated disorders, particularly those mentioned above. The nucleotide sequences encoding KAP may be labeled by standard methods and added to a fluid or tissue sample from a patient under conditions suitable for the formation of hybridization complexes. After a suitable incubation period, the sample is washed and the signal is quantified and compared with a standard value. If the amount of signal in the patient sample is significantly altered in comparison to a control sample then the presence of altered levels of nucleotide sequences encoding KAP in the sample indicates the presence of the associated disorder. Such assays may also be used to evaluate the efficacy of a particular therapeutic treatment regimen in animal studies, in clinical trials, or to monitor the treatment of an individual patient.

In order to provide a basis for the diagnosis of a disorder associated with expression of KAP, a normal or standard profile for expression is established. This may be accomplished by 20 combining body fluids or cell extracts taken from normal subjects, either animal or human, with a sequence, or a fragment thereof, encoding KAP, under conditions suitable for hybridization or amplification. Standard hybridization may be quantified by comparing the values obtained from normal subjects with values from an experiment in which a known amount of a substantially purified polynucleotide is used. Standard values obtained in this manner may be compared with 25 values obtained from samples from patients who are symptomatic for a disorder. Deviation from standard values is used to establish the presence of a disorder.

Once the presence of a disorder is established and a treatment protocol is initiated, hybridization assays may be repeated on a regular basis to determine if the level of expression in the patient begins to approximate that which is observed in the normal subject. The results obtained 30 from successive assays may be used to show the efficacy of treatment over a period ranging from several days to months.

With respect to cancer, the presence of an abnormal amount of transcript (either under- or overexpressed) in biopsied tissue from an individual may indicate a predisposition for the development of the disease, or may provide a means for detecting the disease prior to the 35 appearance of actual clinical symptoms. A more definitive diagnosis of this type may allow health

professionals to employ preventative measures or aggressive treatment earlier thereby preventing the development or further progression of the cancer.

Additional diagnostic uses for oligonucleotides designed from the sequences encoding KAP may involve the use of PCR. These oligomers may be chemically synthesized, generated 5 enzymatically, or produced in vitro. Oligomers will preferably contain a fragment of a polynucleotide encoding KAP, or a fragment of a polynucleotide complementary to the polynucleotide encoding KAP, and will be employed under optimized conditions for identification of a specific gene or condition. Oligomers may also be employed under less stringent conditions for detection or quantification of closely related DNA or RNA sequences.

In a particular aspect, oligonucleotide primers derived from the polynucleotide sequences encoding KAP may be used to detect single nucleotide polymorphisms (SNPs). SNPs are substitutions, insertions and deletions that are a frequent cause of inherited or acquired genetic disease in humans. Methods of SNP detection include, but are not limited to, single-stranded conformation polymorphism (SSCP) and fluorescent SSCP (fSSCP) methods. In SSCP,

- 15 oligonucleotide primers derived from the polynucleotide sequences encoding KAP are used to amplify DNA using the polymerase chain reaction (PCR). The DNA may be derived, for example, from diseased or normal tissue, biopsy samples, bodily fluids, and the like. SNPs in the DNA cause differences in the secondary and tertiary structures of PCR products in single-stranded form, and these differences are detectable using gel electrophoresis in non-denaturing gels. In fSCCP, the
- 20 oligonucleotide primers are fluorescently labeled, which allows detection of the amplimers in high-throughput equipment such as DNA sequencing machines. Additionally, sequence database analysis methods, termed in silico SNP (isSNP), are capable of identifying polymorphisms by comparing the sequence of individual overlapping DNA fragments which assemble into a common consensus sequence. These computer-based methods filter out sequence variations due to
- 25 laboratory preparation of DNA and sequencing errors using statistical models and automated analyses of DNA sequence chromatograms. In the alternative, SNPs may be detected and characterized by mass spectrometry using, for example, the high throughput MASSARRAY system (Sequenom, Inc., San Diego CA).

Methods which may also be used to quantify the expression of KAP include radiolabeling 30 or biotinylating nucleotides, coamplification of a control nucleic acid, and interpolating results from standard curves. (See, e.g., Melby, P.C. et al. (1993) J. Immunol. Methods 159:235-244; Duplaa, C. et al. (1993) Anal. Biochem. 212:229 236.) The speed of quantitation of multiple samples may be accelerated by running the assay in a high-throughput format where the oligomer or polynucleotide of interest is presented in various dilutions and a spectrophotometric or colorimetric response gives 35 rapid quantitation.

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In further embodiments, oligonucleotides or longer fragments derived from any of the polynucleotide sequences described herein may be used as elements on a microarray. The microarray can be used in transcript imaging techniques which monitor the relative expression levels of large numbers of genes simultaneously as described below. The microarray may also be 5 used to identify genetic variants, mutations, and polymorphisms. This information may be used to determine gene function, to understand the genetic basis of a disorder, to diagnose a disorder, to monitor progression/regression of disease as a function of gene expression, and to develop and monitor the activities of therapeutic agents in the treatment of disease. In particular, this information may be used to develop a pharmacogenomic profile of a patient in order to select the 10 most appropriate and effective treatment regimen for that patient. For example, therapeutic agents which are highly effective and display the fewest side effects may be selected for a patient based on his/her pharmacogenomic profile.

In another embodiment, KAP, fragments of KAP, or antibodies specific for KAP may be used as elements on a microarray. The microarray may be used to monitor or measure protein15 protein interactions, drug-target interactions, and gene expression profiles, as described above.

A particular embodiment relates to the use of the polynucleotides of the present invention to generate a transcript image of a tissue or cell type. A transcript image represents the global pattern of gene expression by a particular tissue or cell type. Global gene expression patterns are analyzed by quantifying the number of expressed genes and their relative abundance under given conditions 20 and at a given time. (See Seilhamer et al., "Comparative Gene Transcript Analysis," U.S. Patent No. 5,840,484, expressly incorporated by reference herein.) Thus a transcript image may be generated by hybridizing the polynucleotides of the present invention or their complements to the totality of transcripts or reverse transcripts of a particular tissue or cell type. In one embodiment, the hybridization takes place in high-throughput format, wherein the polynucleotides of the present 25 invention or their complements comprise a subset of a plurality of elements on a microarray. The resultant transcript image would provide a profile of gene activity.

Transcript images may be generated using transcripts isolated from tissues, cell lines, biopsies, or other biological samples. The transcript image may thus reflect gene expression in vivo, as in the case of a tissue or biopsy sample, or in vitro, as in the case of a cell line.

30 Transcript images which profile the expression of the polynucleotides of the present invention may also be used in conjunction with <u>in vitro</u> model systems and preclinical evaluation of pharmaceuticals, as well as toxicological testing of industrial and naturally-occurring environmental compounds. All compounds induce characteristic gene expression patterns, frequently termed molecular fingerprints or toxicant signatures, which are indicative of mechanisms of action and 35 toxicity (Nuwaysir, E.F. et al. (1999) Mol. Carcinog. 24:153-159; Steiner, S. and N.L. Anderson

(2000) Toxicol. Lett. 112-113:467-471, expressly incorporated by reference herein). If a test compound has a signature similar to that of a compound with known toxicity, it is likely to share those toxic properties. These fingerprints or signatures are most useful and refined when they contain expression information from a large number of genes and gene families. Ideally, a genome-5 wide measurement of expression provides the highest quality signature. Even genes whose expression is not altered by any tested compounds are important as well, as the levels of expression of these genes are used to normalize the rest of the expression data. The normalization procedure is useful for comparison of expression data after treatment with different compounds. While the assignment of gene function to elements of a toxicant signature aids in interpretation of toxicity mechanisms, knowledge of gene function is not necessary for the statistical matching of signatures which leads to prediction of toxicity. (See, for example, Press Release 00-02 from the National Institute of Environmental Health Sciences, released February 29, 2000, available at http://www.niehs.nih.gov/oc/news/toxchip.htm.) Therefore, it is important and desirable in toxicological screening using toxicant signatures to include all expressed gene sequences.

In one embodiment, the toxicity of a test compound is assessed by treating a biological sample containing nucleic acids with the test compound. Nucleic acids that are expressed in the treated biological sample are hybridized with one or more probes specific to the polynucleotides of the present invention, so that transcript levels corresponding to the polynucleotides of the present invention may be quantified. The transcript levels in the treated biological sample are compared 20 with levels in an untreated biological sample. Differences in the transcript levels between the two samples are indicative of a toxic response caused by the test compound in the treated sample.

Another particular embodiment relates to the use of the polypeptide sequences of the present invention to analyze the proteome of a tissue or cell type. The term proteome refers to the global pattern of protein expression in a particular tissue or cell type. Each protein component of a 25 proteome can be subjected individually to further analysis. Proteome expression patterns, or profiles, are analyzed by quantifying the number of expressed proteins and their relative abundance under given conditions and at a given time. A profile of a cell's proteome may thus be generated by separating and analyzing the polypeptides of a particular tissue or cell type. In one embodiment, the separation is achieved using two-dimensional gel electrophoresis, in which proteins from a sample 30 are separated by isoelectric focusing in the first dimension, and then according to molecular weight by sodium dodecyl sulfate slab gel electrophoresis in the second dimension (Steiner and Anderson,

supra). The proteins are visualized in the gel as discrete and uniquely positioned spots, typically by staining the gel with an agent such as Coomassie Blue or silver or fluorescent stains. The optical density of each protein spot is generally proportional to the level of the protein in the sample. The

biological samples either treated or untreated with a test compound or therapeutic agent, are compared to identify any changes in protein spot density related to the treatment. The proteins in the spots are partially sequenced using, for example, standard methods employing chemical or enzymatic cleavage followed by mass spectrometry. The identity of the protein in a spot may be determined by comparing its partial sequence, preferably of at least 5 contiguous amino acid residues, to the polypeptide sequences of the present invention. In some cases, further sequence data may be obtained for definitive protein identification.

A proteomic profile may also be generated using antibodies specific for KAP to quantify the levels of KAP expression. In one embodiment, the antibodies are used as elements on a microarray, 10 and protein expression levels are quantified by exposing the microarray to the sample and detecting the levels of protein bound to each array element (Lueking, A. et al. (1999) Anal. Biochem. 270:103-111; Mendoze, L.G. et al. (1999) Biotechniques 27:778-788). Detection may be performed by a variety of methods known in the art, for example, by reacting the proteins in the sample with a thiol- or amino-reactive fluorescent compound and detecting the amount of fluorescence bound at 15 each array element.

Toxicant signatures at the proteome level are also useful for toxicological screening, and should be analyzed in parallel with toxicant signatures at the transcript level. There is a poor correlation between transcript and protein abundances for some proteins in some tissues (Anderson, N.L. and J. Seilhamer (1997) Electrophoresis 18:533-537), so proteome toxicant signatures may be useful in the analysis of compounds which do not significantly affect the transcript image, but which alter the proteomic profile. In addition, the analysis of transcripts in body fluids is difficult, due to rapid degradation of mRNA, so proteomic profiling may be more reliable and informative in such cases.

In another embodiment, the toxicity of a test compound is assessed by treating a biological sample containing proteins with the test compound. Proteins that are expressed in the treated biological sample are separated so that the amount of each protein can be quantified. The amount of each protein is compared to the amount of the corresponding protein in an untreated biological sample. A difference in the amount of protein between the two samples is indicative of a toxic response to the test compound in the treated sample. Individual proteins are identified by sequencing the amino acid residues of the individual proteins and comparing these partial sequences to the polypeptides of the present invention.

In another embodiment, the toxicity of a test compound is assessed by treating a biological sample containing proteins with the test compound. Proteins from the biological sample are incubated with antibodies specific to the polypeptides of the present invention. The amount of protein recognized by the antibodies is quantified. The amount of protein in the treated biological

sample is compared with the amount in an untreated biological sample. A difference in the amount of protein between the two samples is indicative of a toxic response to the test compound in the treated sample.

Microarrays may be prepared, used, and analyzed using methods known in the art. (See, 5 e.g., Brennan, T.M. et al. (1995) U.S. Patent No. 5,474,796; Schena, M. et al. (1996) Proc. Natl. Acad. Sci. USA 93:10614-10619; Baldeschweiler et al. (1995) PCT application WO95/251116; Shalon, D. et al. (1995) PCT application WO95/35505; Heller, R.A. et al. (1997) Proc. Natl. Acad. Sci. USA 94:2150-2155; and Heller, M.J. et al. (1997) U.S. Patent No. 5,605,662.) Various types of microarrays are well known and thoroughly described in DNA Microarrays: A Practical Approach, 10 M. Schena, ed. (1999) Oxford University Press, London, hereby expressly incorporated by reference.

In another embodiment of the invention, nucleic acid sequences encoding KAP may be used to generate hybridization probes useful in mapping the naturally occurring genomic sequence. Either coding or noncoding sequences may be used, and in some instances, noncoding sequences 15 may be preferable over coding sequences. For example, conservation of a coding sequence among members of a multi-gene family may potentially cause undesired cross hybridization during chromosomal mapping. The sequences may be mapped to a particular chromosome, to a specific region of a chromosome, or to artificial chromosome constructions, e.g., human artificial chromosomes (HACs), yeast artificial chromosomes (YACs), bacterial artificial chromosomes 20 (BACs), bacterial P1 constructions, or single chromosome cDNA libraries. (See, e.g., Harrington, J.J. et al. (1997) Nat. Genet. 15:345-355; Price, C.M. (1993) Blood Rev. 7:127-134; and Trask, B.J. (1991) Trends Genet. 7:149-154.) Once mapped, the nucleic acid sequences of the invention may be used to develop genetic linkage maps, for example, which correlate the inheritance of a disease state with the inheritance of a particular chromosome region or restriction fragment length 25 polymorphism (RFLP). (See, for example, Lander, E.S. and D. Botstein (1986) Proc. Natl. Acad. Sci. USA 83:7353-7357.)

Fluorescent in situ hybridization (FISH) may be correlated with other physical and genetic map data. (See, e.g., Heinz-Ulrich, et al. (1995) in Meyers, supra, pp. 965-968.) Examples of genetic map data can be found in various scientific journals or at the Online Mendelian Inheritance 30 in Man (OMIM) World Wide Web site. Correlation between the location of the gene encoding KAP on a physical map and a specific disorder, or a predisposition to a specific disorder, may help define the region of DNA associated with that disorder and thus may further positional cioning efforts.

<u>In situ</u> hybridization of chromosomal preparations and physical mapping techniques, such as 35 linkage analysis using established chromosomal markers, may be used for extending genetic maps.

Often the placement of a gene on the chromosome of another mammalian species, such as mouse, may reveal associated markers even if the exact chromosomal locus is not known. This information is valuable to investigators searching for disease genes using positional cloning or other gene discovery techniques. Once the gene or genes responsible for a disease or syndrome have been 5 crudely localized by genetic linkage to a particular genomic region, e.g., ataxia-telangiectasia to 11q22-23, any sequences mapping to that area may represent associated or regulatory genes for further investigation. (See, e.g., Gatti, R.A. et al. (1988) Nature 336:577-580.) The nucleotide sequence of the instant invention may also be used to detect differences in the chromosomal location due to translocation, inversion, etc., among normal, carrier, or affected individuals.

In another embodiment of the invention, KAP, its catalytic or immunogenic fragments, or oligopeptides thereof can be used for screening libraries of compounds in any of a variety of drug screening techniques. The fragment employed in such screening may be free in solution, affixed to a solid support, borne on a cell surface, or located intracellularly. The formation of binding complexes between KAP and the agent being tested may be measured.

Another technique for drug screening provides for high throughput screening of compounds having suitable binding affinity to the protein of interest. (See, e.g., Geysen, et al. (1984) PCT application WO84/03564.) In this method, large numbers of different small test compounds are synthesized on a solid substrate. The test compounds are reacted with KAP, or fragments thereof, and washed. Bound KAP is then detected by methods well known in the art. Purified KAP can also be coated directly onto plates for use in the aforementioned drug screening techniques.

Alternatively, non-neutralizing antibodies can be used to capture the peptide and immobilize it on a

In another embodiment, one may use competitive drug screening assays in which neutralizing antibodies capable of binding KAP specifically compete with a test compound for 25 binding KAP. In this manner, antibodies can be used to detect the presence of any peptide which shares one or more antigenic determinants with KAP.

In additional embodiments, the nucleotide sequences which encode KAP may be used in any molecular biology techniques that have yet to be developed, provided the new techniques rely on properties of nucleotide sequences that are currently known, including, but not limited to, such 30 properties as the triplet genetic code and specific base pair interactions.

Without further elaboration, it is believed that one skilled in the art can, using the preceding description, utilize the present invention to its fullest extent. The following embodiments are, therefore, to be construed as merely illustrative, and not limitative of the remainder of the disclosure in any way whatsoever.

The disclosures of all patents, applications and publications, mentioned above and below,

including U.S. Ser. No. 60/254,034, U.S. Ser. No. 60/255,756, U.S. Ser. No. 60/251,814, U.S. Ser. No. 60/256,172, U.S. Ser. No. 60/257,416, U.S. Ser. No. 60/260,912, U.S. Ser. No. 60/264,344, and U.S. Ser. No.60/266,017, are expressly incorporated by reference herein.

5 EXAMPLES

I. Construction of cDNA Libraries

Incyte cDNAs were derived from cDNA libraries described in the LIFESEQ GOLD database (Incyte Genomics, Palo Alto CA). Some tissues were homogenized and lysed in guanidinium isothiocyanate, while others were homogenized and lysed in phenol or in a suitable 10 mixture of denaturants, such as TRIZOL (Life Technologies), a monophasic solution of phenol and guanidine isothiocyanate. The resulting lysates were centrifuged over CsCl cushions or extracted with chloroform. RNA was precipitated from the lysates with either isopropanol or sodium acetate and ethanol, or by other routine methods.

Phenol extraction and precipitation of RNA were repeated as necessary to increase RNA 15 purity. In some cases, RNA was treated with DNase. For most libraries, poly(A)+ RNA was isolated using oligo d(T)-coupled paramagnetic particles (Promega), OLIGOTEX latex particles (QIAGEN, Chatsworth CA), or an OLIGOTEX mRNA purification kit (QIAGEN). Alternatively, RNA was isolated directly from tissue lysates using other RNA isolation kits, e.g., the POLY(A)PURE mRNA purification kit (Ambion, Austin TX).

20 In some cases, Stratagene was provided with RNA and constructed the corresponding cDNA libraries. Otherwise, cDNA was synthesized and cDNA libraries were constructed with the UNIZAP vector system (Stratagene) or SUPERSCRIPT plasmid system (Life Technologies), using the recommended procedures or similar methods known in the art. (See, e.g., Ausubel, 1997, supra, units 5.1-6.6.) Reverse transcription was initiated using oligo d(T) or random primers. Synthetic 25 oligonucleotide adapters were ligated to double stranded cDNA, and the cDNA was digested with the appropriate restriction enzyme or enzymes. For most libraries, the cDNA was size-selected (300-1000 bp) using SEPHACRYL S1000, SEPHAROSE CL2B, or SEPHAROSE CL4B column chromatography (Amersham Pharmacia Biotech) or preparative agarose gel electrophoresis. cDNAs were ligated into compatible restriction enzyme sites of the polylinker of a suitable plasmid, 30 e.g., PBLUESCRIPT plasmid (Stratagene), PSPORT1 plasmid (Life Technologies), PCDNA2.1 plasmid (Invitrogen, Carlsbad CA), PBK-CMV plasmid (Stratagene), PCR2-TOPOTA plasmid (Invitrogen), PCMV-ICIS plasmid (Stratagene), pIGEN (Incyte Genomics, Palo Alto CA), pRARE (Incyte Genomics), or pINCY (Incyte Genomics), or derivatives thereof. Recombinant plasmids were transformed into competent E. coli cells including XL1-Blue, XL1-BlueMRF, or SOLR from 35 Stratagene or DH5a, DH10B, or ElectroMAX DH10B from Life Technologies.

II. Isolation of cDNA Clones

Plasmids obtained as described in Example I were recovered from host cells by in vivo excision using the UNIZAP vector system (Stratagene) or by cell lysis. Plasmids were purified using at least one of the following: a Magic or WIZARD Minipreps DNA purification system 5 (Promega); an AGTC Miniprep purification kit (Edge Biosystems, Gaithersburg MD); and QIAWELL 8 Plasmid, QIAWELL 8 Plasmid, QIAWELL 8 Ultra Plasmid purification systems or the R.E.A.L. PREP 96 plasmid purification kit from QIAGEN. Following precipitation, plasmids were resuspended in 0.1 ml of distilled water and stored, with or without lyophilization, at 4°C.

Alternatively, plasmid DNA was amplified from host cell lysates using direct link PCR in a 10 high-throughput format (Rao, V.B. (1994) Anal. Biochem. 216:1-14). Host cell lysis and thermal cycling steps were carried out in a single reaction mixture. Samples were processed and stored in 384-well plates, and the concentration of amplified plasmid DNA was quantified fluorometrically using PICOGREEN dye (Molecular Probes, Eugene OR) and a FLUOROSKAN II fluorescence scanner (Labsystems Oy, Helsinki, Finland).

15 III. Sequencing and Analysis

Incyte cDNA recovered in plasmids as described in Example II were sequenced as follows. Sequencing reactions were processed using standard methods or high-throughput instrumentation such as the ABI CATALYST 800 (Applied Biosystems) thermal cycler or the PTC-200 thermal cycler (MJ Research) in conjunction with the HYDRA microdispenser (Robbins Scientific) or the 20 MICROLAB 2200 (Hamilton) liquid transfer system. cDNA sequencing reactions were prepared using reagents provided by Amersham Pharmacia Biotech or supplied in ABI sequencing kits such as the ABI PRISM BIGDYE Terminator cycle sequencing ready reaction kit (Applied Biosystems). Electrophoretic separation of cDNA sequencing reactions and detection of labeled polynucleotides were carried out using the MEGABACE 1000 DNA sequencing system (Molecular Dynamics); the 25 ABI PRISM 373 or 377 sequencing system (Applied Biosystems) in conjunction with standard ABI protocols and base calling software; or other sequence analysis systems known in the art. Reading frames within the cDNA sequences were identified using standard methods (reviewed in Ausubel, 1997, supra, unit 7.7). Some of the cDNA sequences were selected for extension using the techniques disclosed in Example VIII.

The polynucleotide sequences derived from Incyte cDNAs were validated by removing vector, linker, and poly(A) sequences and by masking ambiguous bases, using algorithms and programs based on BLAST, dynamic programming, and dinucleotide nearest neighbor analysis. The Incyte cDNA sequences or translations thereof were then queried against a selection of public databases such as the GenBank primate, rodent, mammalian, vertebrate, and eukaryote databases, and BLOCKS, PRINTS, DOMO, PRODOM; PROTEOME databases with sequences from Homo

sapiens, Rattus norvegicus, Mus musculus, Caenorhabditis elegans, Saccharomyces cerevisiae, Schizosaccharomyces pombe, and Candida albicans (Incyte Genomics, Palo Alto CA); and hidden Markov model (HMM)-based protein family databases such as PFAM. (HMM is a probabilistic approach which analyzes consensus primary structures of gene families. See, for example, Eddy,

- 5 S.R. (1996) Curr. Opin. Struct. Biol. 6:361-365.) The queries were performed using programs based on BLAST, FASTA, BLIMPS, and HMMER. The Incyte cDNA sequences were assembled to produce full length polynucleotide sequences. Alternatively, GenBank cDNAs, GenBank ESTs, stitched sequences, stretched sequences, or Genscan-predicted coding sequences (see Examples IV and V) were used to extend Incyte cDNA assemblages to full length. Assembly was performed
- 10 using programs based on Phred, Phrap, and Consed, and cDNA assemblages were screened for open reading frames using programs based on GeneMark, BLAST, and FASTA. The full length polynucleotide sequences were translated to derive the corresponding full length polypeptide sequences. Alternatively, a polypeptide of the invention may begin at any of the methionine residues of the full length translated polypeptide. Full length polypeptide sequences were
- 15 subsequently analyzed by querying against databases such as the GenBank protein databases (genpept), SwissProt, the PROTEOME databases, BLOCKS, PRINTS, DOMO, PRODOM, Prosite, and hidden Markov model (HMM)-based protein family databases such as PFAM. Full length polynucleotide sequences are also analyzed using MACDNASIS PRO software (Hitachi Software Engineering, South San Francisco CA) and LASERGENE software (DNASTAR). Polynucleotide
- 20 and polypeptide sequence alignments are generated using default parameters specified by the CLUSTAL algorithm as incorporated into the MEGALIGN multisequence alignment program (DNASTAR), which also calculates the percent identity between aligned sequences.

Table 7 summarizes the tools, programs, and algorithms used for the analysis and assembly of Incyte cDNA and full length sequences and provides applicable descriptions, references, and 25 threshold parameters. The first column of Table 7 shows the tools, programs, and algorithms used, the second column provides brief descriptions thereof, the third column presents appropriate references, all of which are incorporated by reference herein in their entirety, and the fourth column presents, where applicable, the scores, probability values, and other parameters used to evaluate the strength of a match between two sequences (the higher the score or the lower the probability value, 30 the greater the identity between two sequences).

The programs described above for the assembly and analysis of full length polynucleotide and polypeptide sequences were also used to identify polynucleotide sequence fragments from SEQ ID NO:21-40. Fragments from about 20 to about 4000 nucleotides which are useful in hybridization and amplification technologies are described in Table 4, column 2.

35 IV. Identification and Editing of Coding Sequences from Genomic DNA

Putative kinases and phosphatases were initially identified by running the Genscan gene identification program against public genomic sequence databases (e.g., gbpri and gbhtg). Genscan is a general-purpose gene identification program which analyzes genomic DNA sequences from a variety of organisms (See Burge, C. and S. Karlin (1997) J. Mol. Biol. 268:78-94, and Burge, C. and 5 S. Karlin (1998) Curr. Opin. Struct. Biol. 8:346-354). The program concatenates predicted exons to form an assembled cDNA sequence extending from a methionine to a stop codon. The output of Genscan is a FASTA database of polynucleotide and polypeptide sequences. The maximum range of sequence for Genscan to analyze at once was set to 30 kb. To determine which of these Genscan predicted cDNA sequences encode kinases and phosphatases, the encoded polypeptides were 10 analyzed by querying against PFAM models for kinases and phosphatases. Potential kinases and phosphatases were also identified by homology to Incyte cDNA sequences that had been annotated as kinases and phosphatases. These selected Genscan-predicted sequences were then compared by BLAST analysis to the genpept and gbpri public databases. Where necessary, the Genscanpredicted sequences were then edited by comparison to the top BLAST hit from genpept to correct 15 errors in the sequence predicted by Genscan, such as extra or omitted exons. BLAST analysis was also used to find any Incyte cDNA or public cDNA coverage of the Genscan-predicted sequences, thus providing evidence for transcription. When Incyte cDNA coverage was available, this information was used to correct or confirm the Genscan predicted sequence. Full length polynucleotide sequences were obtained by assembling Genscan-predicted coding sequences with 20 Incyte cDNA sequences and/or public cDNA sequences using the assembly process described in Example III. Alternatively, full length polynucleotide sequences were derived entirely from edited or unedited Genscan-predicted coding sequences.

V. Assembly of Genomic Sequence Data with cDNA Sequence Data "Stitched" Sequences

Partial cDNA sequences were extended with exons predicted by the Genscan gene identification program described in Example IV. Partial cDNAs assembled as described in Example III were mapped to genomic DNA and parsed into clusters containing related cDNAs and Genscan exon predictions from one or more genomic sequences. Each cluster was analyzed using an algorithm based on graph theory and dynamic programming to integrate cDNA and genomic information, generating possible splice variants that were subsequently confirmed, edited, or extended to create a full length sequence. Sequence intervals in which the entire length of the interval was present on more than one sequence in the cluster were identified, and intervals thus identified were considered to be equivalent by transitivity. For example, if an interval was present on a cDNA and two genomic sequences, then all three intervals were considered to be equivalent.

35 This process allows unrelated but consecutive genomic sequences to be brought together, bridged by

cDNA sequence. Intervals thus identified were then "stitched" together by the stitching algorithm in the order that they appear along their parent sequences to generate the longest possible sequence, as well as sequence variants. Linkages between intervals which proceed along one type of parent sequence (cDNA to cDNA or genomic sequence to genomic sequence) were given preference over 5 linkages which change parent type (cDNA to genomic sequence). The resultant stitched sequences were translated and compared by BLAST analysis to the genpept and gbpri public databases.

Incorrect exons predicted by Genscan were corrected by comparison to the top BLAST hit from genpept. Sequences were further extended with additional cDNA sequences, or by inspection of genomic DNA, when necessary.

10 "Stretched" Sequences

Partial DNA sequences were extended to full length with an algorithm based on BLAST analysis. First, partial cDNAs assembled as described in Example III were queried against public databases such as the GenBank primate, rodent, mammalian, vertebrate, and eukaryote databases using the BLAST program. The nearest GenBank protein homolog was then compared by BLAST analysis to either Incyte cDNA sequences or GenScan exon predicted sequences described in Example IV. A chimeric protein was generated by using the resultant high-scoring segment pairs (HSPs) to map the translated sequences onto the GenBank protein homolog. Insertions or deletions may occur in the chimeric protein with respect to the original GenBank protein homolog. The GenBank protein homolog, the chimeric protein, or both were used as probes to search for 20 homologous genomic sequences from the public human genome databases. Partial DNA sequences were therefore "stretched" or extended by the addition of homologous genomic sequences. The resultant stretched sequences were examined to determine whether it contained a complete gene.

VI. Chromosomal Mapping of KAP Encoding Polynucleotides

The sequences which were used to assemble SEQ ID NO:21-40 were compared with sequences from the Incyte LIFESEQ database and public domain databases using BLAST and other implementations of the Smith-Waterman algorithm. Sequences from these databases that matched SEQ ID NO:21-40 were assembled into clusters of contiguous and overlapping sequences using assembly algorithms such as Phrap (Table 7). Radiation hybrid and genetic mapping data available from public resources such as the Stanford Human Genome Center (SHGC), Whitehead Institute for 30 Genome Research (WIGR), and Généthon were used to determine if any of the clustered sequences had been previously mapped. Inclusion of a mapped sequence in a cluster resulted in the assignment of all sequences of that cluster, including its particular SEQ ID NO:, to that map location.

Map locations are represented by ranges, or intervals, of human chromosomes. The map 35 position of an interval, in centiMorgans, is measured relative to the terminus of the chromosome's

p-arm. (The centiMorgan (cM) is a unit of measurement based on recombination frequencies between chromosomal markers. On average, 1 cM is roughly equivalent to 1 megabase (Mb) of DNA in humans, although this can vary widely due to hot and cold spots of recombination.) The cM distances are based on genetic markers mapped by Généthon which provide boundaries for radiation hybrid markers whose sequences were included in each of the clusters. Human genome maps and other resources available to the public, such as the NCBI "GeneMap'99" World Wide Web site (http://www.ncbi.nlm.nih.gov/genemap/), can be employed to determine if previously identified disease genes map within or in proximity to the intervals indicated above.

In this manner, SEQ ID NO:33 was mapped to chromosome 12 within the interval from 10 97.10 to 113.30 centiMorgans. SEQ ID NO:35 was mapped to chromosome 3 within the interval from 16.50 to 30.40 centiMorgans. SEQ ID NO:29 was mapped to chromosome 13 within the interval from 11.60 to 22.80 centiMorgans, to chromosome 15 within the interval from 72.30 to 77.30 centiMorgans, and to chromosome 20 within the interval from 57.70 to 64.10 centiMorgans. More than one map location is reported for SEQ ID NO:29, indicating that sequences having 15 different map locations were assembled into a single cluster. This situation occurs, for example, when sequences having strong similarity, but not complete identity, are assembled into a single cluster.

VII. Analysis of Polynucleotide Expression

Northern analysis is a laboratory technique used to detect the presence of a transcript of a 20 gene and involves the hybridization of a labeled nucleotide sequence to a membrane on which RNAs from a particular cell type or tissue have been bound. (See, e.g., Sambrook, supra, ch. 7; Ausubel (1995) supra, ch. 4 and 16.)

. Analogous computer techniques applying BLAST were used to search for identical or related molecules in cDNA databases such as GenBank or LIFESEQ (Incyte Genomics). This 25 analysis is much faster than multiple membrane-based hybridizations. In addition, the sensitivity of the computer search can be modified to determine whether any particular match is categorized as exact or similar. The basis of the search is the product score, which is defined as:

BLAST Score x Percent Identity

5 x minimum {length(Seq. 1), length(Seq. 2)}

The product score takes into account both the degree of similarity between two sequences and the length of the sequence match. The product score is a normalized value between 0 and 100, and is calculated as follows: the BLAST score is multiplied by the percent nucleotide identity and the product is divided by (5 times the length of the shorter of the two sequences). The BLAST score is

calculated by assigning a score of +5 for every base that matches in a high-scoring segment pair (HSP), and -4 for every mismatch. Two sequences may share more than one HSP (separated by gaps). If there is more than one HSP, then the pair with the highest BLAST score is used to calculate the product score. The product score represents a balance between fractional overlap and 5 quality in a BLAST alignment. For example, a product score of 100 is produced only for 100% identity over the entire length of the shorter of the two sequences being compared. A product score of 70 is produced either by 100% identity and 70% overlap at one end, or by 88% identity and 100% overlap at the other. A product score of 50 is produced either by 100% identity and 50% overlap at one end, or 79% identity and 100% overlap.

10 Alternatively, polynucleotide sequences encoding KAP are analyzed with respect to the tissue sources from which they were derived. For example, some full length sequences are assembled, at least in part, with overlapping Incyte cDNA sequences (see Example III). Each cDNA sequence is derived from a cDNA library constructed from a human tissue. Each human tissue is classified into one of the following organ/tissue categories: cardiovascular system; connective 15 tissue; digestive system; embryonic structures; endocrine system; exocrine glands; genitalia, female; genitalia, male; germ cells; hemic and immune system; liver; musculoskeletal system; nervous system; pancreas; respiratory system; sense organs; skin; stomatognathic system; unclassified/mixed; or urinary tract. The number of libraries in each category is counted and divided by the total number of libraries across all categories. Similarly, each human tissue is 20 classified into one of the following disease/condition categories: cancer, cell line, developmental, inflammation, neurological, trauma, cardiovascular, pooled, and other, and the number of libraries in each category is counted and divided by the total number of libraries across all categories. The resulting percentages reflect the tissue- and disease-specific expression of cDNA encoding KAP. cDNA sequences and cDNA library/tissue information are found in the LIFESEQ GOLD database 25 (Incyte Genomics, Palo Alto CA).

VIII. Extension of KAP Encoding Polynucleotides

Full length polynucleotide sequences were also produced by extension of an appropriate fragment of the full length molecule using oligonucleotide primers designed from this fragment. One primer was synthesized to initiate 5' extension of the known fragment, and the other primer was 30 synthesized to initiate 3' extension of the known fragment. The initial primers were designed using OLIGO 4.06 software (National Biosciences), or another appropriate program, to be about 22 to 30 nucleotides in length, to have a GC content of about 50% or more, and to anneal to the target sequence at temperatures of about 68°C to about 72°C. Any stretch of nucleotides which would result in hairpin structures and primer-primer dimerizations was avoided.

35 Selected human cDNA libraries were used to extend the sequence. If more than one

extension was necessary or desired, additional or nested sets of primers were designed.

High fidelity amplification was obtained by PCR using methods well known in the art. PCR was performed in 96-well plates using the PTC-200 thermal cycler (MJ Research, Inc.). The reaction mix contained DNA template, 200 nmol of each primer, reaction buffer containing Mg²⁺, 5 (NH₄)₂SO₄, and 2-mercaptoethanol, Taq DNA polymerase (Amersham Pharmacia Biotech), ELONGASE enzyme (Life Technologies), and Pfu DNA polymerase (Stratagene), with the following parameters for primer pair PCI A and PCI B: Step 1: 94°C, 3 min; Step 2: 94°C, 15 sec; Step 3: 60°C, 1 min; Step 4: 68°C, 2 min; Step 5: Steps 2, 3, and 4 repeated 20 times; Step 6: 68°C, 5 min; Step 7: storage at 4°C. In the alternative, the parameters for primer pair T7 and SK+ were as 10 follows: Step 1: 94°C, 3 min; Step 2: 94°C, 15 sec; Step 3: 57°C, 1 min; Step 4: 68°C, 2 min; Step 5: Steps 2, 3, and 4 repeated 20 times; Step 6: 68°C, 5 min; Step 7: storage at 4°C.

The concentration of DNA in each well was determined by dispensing 100 μ l PICOGREEN quantitation reagent (0.25% (v/v) PICOGREEN; Molecular Probes, Eugene OR) dissolved in 1X TE and 0.5 μ l of undiluted PCR product into each well of an opaque fluorimeter plate (Corning Costar, 15 Acton MA), allowing the DNA to bind to the reagent. The plate was scanned in a Fluoroskan II (Labsystems Oy, Helsinki, Finland) to measure the fluorescence of the sample and to quantify the concentration of DNA. A 5 μ l to 10 μ l aliquot of the reaction mixture was analyzed by electrophoresis on a 1% agarose gel to determine which reactions were successful in extending the sequence.

The extended nucleotides were desalted and concentrated, transferred to 384-well plates, digested with CviJI cholera virus endonuclease (Molecular Biology Research, Madison WI), and sonicated or sheared prior to religation into pUC 18 vector (Amersham Pharmacia Biotech). For shotgun sequencing, the digested nucleotides were separated on low concentration (0.6 to 0.8%) agarose gels, fragments were excised, and agar digested with Agar ACE (Promega). Extended clones were religated using T4 ligase (New England Biolabs, Beverly MA) into pUC 18 vector (Amersham Pharmacia Biotech), treated with Pfu DNA polymerase (Stratagene) to fill-in restriction site overhangs, and transfected into competent <u>E. coli</u> cells. Transformed cells were selected on antibiotic-containing media, and individual colonies were picked and cultured overnight at 37°C in 384-well plates in LB/2x carb liquid media.

The cells were lysed, and DNA was amplified by PCR using Taq DNA polymerase (Amersham Pharmacia Biotech) and Pfu DNA polymerase (Stratagene) with the following parameters: Step 1: 94°C, 3 min; Step 2: 94°C, 15 sec; Step 3: 60°C, 1 min; Step 4: 72°C, 2 min; Step 5: steps 2, 3, and 4 repeated 29 times; Step 6: 72°C, 5 min; Step 7: storage at 4°C. DNA was quantified by PICOGREEN reagent (Molecular Probes) as described above. Samples with low 35 DNA recoveries were reamplified using the same conditions as described above. Samples were

diluted with 20% dimethysulfoxide (1:2, v/v), and sequenced using DYENAMIC energy transfer sequencing primers and the DYENAMIC DIRECT kit (Amersham Pharmacia Biotech) or the ABI PRISM BIGDYE Terminator cycle sequencing ready reaction kit (Applied Biosystems).

In like manner, full length polynucleotide sequences are verified using the above procedure 5 or are used to obtain 5' regulatory sequences using the above procedure along with oligonucleotides designed for such extension, and an appropriate genomic library.

IX. Labeling and Use of Individual Hybridization Probes

Hybridization probes derived from SEQ ID NO:21-40 are employed to screen cDNAs, genomic DNAs, or mRNAs. Although the labeling of oligonucleotides, consisting of about 20 base 10 pairs, is specifically described, essentially the same procedure is used with larger nucleotide fragments. Oligonucleotides are designed using state-of-the-art software such as OLIGO 4.06 software (National Biosciences) and labeled by combining 50 pmol of each oligomer, 250 μCi of [γ-32P] adenosine triphosphate (Amersham Pharmacia Biotech), and T4 polynucleotide kinase (DuPont NEN, Boston MA). The labeled oligonucleotides are substantially purified using a 15 SEPHADEX G-25 superfine size exclusion dextran bead column (Amersham Pharmacia Biotech). An aliquot containing 10⁷ counts per minute of the labeled probe is used in a typical membrane-based hybridization analysis of human genomic DNA digested with one of the following endonucleases: Ase I, Bgl II, Eco RI, Pst I, Xba I, or Pvu II (DuPont NEN).

The DNA from each digest is fractionated on a 0.7% agarose gel and transferred to nylon 20 membranes (Nytran Plus, Schleicher & Schuell, Durham NH). Hybridization is carried out for 16 hours at 40°C. To remove nonspecific signals, blots are sequentially washed at room temperature under conditions of up to, for example, 0.1 x saline sodium citrate and 0.5% sodium dodecyl sulfate. Hybridization patterns are visualized using autoradiography or an alternative imaging means and compared.

25 X. Microarrays

The linkage or synthesis of array elements upon a microarray can be achieved utilizing photolithography, piezoelectric printing (ink-jet printing, See, e.g., Baldeschweiler, supra.), mechanical microspotting technologies, and derivatives thereof. The substrate in each of the aforementioned technologies should be uniform and solid with a non-porous surface (Schena 30 (1999), supra). Suggested substrates include silicon, silica, glass slides, glass chips, and silicon wafers. Alternatively, a procedure analogous to a dot or slot blot may also be used to arrange and link elements to the surface of a substrate using thermal, UV, chemical, or mechanical bonding procedures. A typical array may be produced using available methods and machines well known to those of ordinary skill in the art and may contain any appropriate number of elements. (See, e.g., 35 Schena, M. et al. (1995) Science 270:467-470; Shalon, D. et al. (1996) Genome Res. 6:639-645;

Marshall, A. and J. Hodgson (1998) Nat. Biotechnol. 16:27-31.)

Full length cDNAs, Expressed Sequence Tags (ESTs), or fragments or oligomers thereof may comprise the elements of the microarray. Fragments or oligomers suitable for hybridization can be selected using software well known in the art such as LASERGENE software (DNASTAR).

5 The array elements are hybridized with polynucleotides in a biological sample. The polynucleotides in the biological sample are conjugated to a fluorescent label or other molecular tag for ease of detection. After hybridization, nonhybridized nucleotides from the biological sample are removed, and a fluorescence scanner is used to detect hybridization at each array element. Alternatively, laser desorbtion and mass spectrometry may be used for detection of hybridization. The degree of 10 complementarity and the relative abundance of each polynucleotide which hybridizes to an element on the microarray may be assessed. In one embodiment, microarray preparation and usage is described in detail below.

Tissue or Cell Sample Preparation

Total RNA is isolated from tissue samples using the guanidinium thiocyanate method and 15 poly(A)⁺ RNA is purified using the oligo-(dT) cellulose method. Each poly(A)⁺ RNA sample is reverse transcribed using MMLV reverse-transcriptase, 0.05 pg/ μ l oligo-(dT) primer (21mer), 1X first strand buffer, 0.03 units/ μ l RNase inhibitor, 500 μ M dATP, 500 μ M dGTP, 500 μ M dTTP, 40 μ M dCTP-Cy3 (BDS) or dCTP-Cy5 (Amersham Pharmacia Biotech). The reverse transcription reaction is performed in a 25 ml volume containing 200 ng poly(A)⁺ RNA with

- 20 GEMBRIGHT kits (Incyte). Specific control poly(A)* RNAs are synthesized by in vitro transcription from non-coding yeast genomic DNA. After incubation at 37°C for 2 hr, each reaction sample (one with Cy3 and another with Cy5 labeling) is treated with 2.5 ml of 0.5M sodium hydroxide and incubated for 20 minutes at 85°C to the stop the reaction and degrade the RNA. Samples are purified using two successive CHROMA SPIN 30 gel filtration spin columns
- 25 (CLONTECH Laboratories, Inc. (CLONTECH), Palo Alto CA) and after combining, both reaction samples are ethanol precipitated using 1 ml of glycogen (1 mg/ml), 60 ml sodium acetate, and 300 ml of 100% ethanol. The sample is then dried to completion using a SpeedVAC (Savant Instruments Inc., Holbrook NY) and resuspended in 14 μl 5X SSC/0.2% SDS.

Microarray Preparation

Sequences of the present invention are used to generate array elements. Each array element is amplified from bacterial cells containing vectors with cloned cDNA inserts. PCR amplification uses primers complementary to the vector sequences flanking the cDNA insert. Array elements are amplified in thirty cycles of PCR from an initial quantity of 1-2 ng to a final quantity greater than 5 μg. Amplified array elements are then purified using SEPHACRYL-400 (Amersham Pharmacia 35 Biotech).

Purified array elements are immobilized on polymer-coated glass slides. Glass microscope slides (Corning) are cleaned by ultrasound in 0.1% SDS and acetone, with extensive distilled water washes between and after treatments. Glass slides are etched in 4% hydrofluoric acid (VWR Scientific Products Corporation (VWR), West Chester PA), washed extensively in distilled water, 5 and coated with 0.05% aminopropyl silane (Sigma) in 95% ethanol. Coated slides are cured in a 110°C oven.

Array elements are applied to the coated glass substrate using a procedure described in U.S. Patent No. 5,807,522, incorporated herein by reference. 1 μ l of the array element DNA, at an average concentration of 100 ng/ μ l, is loaded into the open capillary printing element by a high-10 speed robotic apparatus. The apparatus then deposits about 5 nl of array element sample per slide.

Microarrays are UV-crosslinked using a STRATALINKER UV-crosslinker (Stratagene). Microarrays are washed at room temperature once in 0.2% SDS and three times in distilled water. Non-specific binding sites are blocked by incubation of microarrays in 0.2% casein in phosphate buffered saline (PBS) (Tropix, Inc., Bedford MA) for 30 minutes at 60°C followed by washes in 15 0.2% SDS and distilled water as before.

Hybridization

Hybridization reactions contain 9 μl of sample mixture consisting of 0.2 μg each of Cy3 and Cy5 labeled cDNA synthesis products in 5X SSC, 0.2% SDS hybridization buffer. The sample mixture is heated to 65°C for 5 minutes and is aliquoted onto the microarray surface and covered 20 with an 1.8 cm² coverslip. The arrays are transferred to a waterproof chamber having a cavity just slightly larger than a microscope slide. The chamber is kept at 100% humidity internally by the addition of 140 μl of 5X SSC in a corner of the chamber. The chamber containing the arrays is incubated for about 6.5 hours at 60°C. The arrays are washed for 10 min at 45°C in a first wash buffer (1X SSC, 0.1% SDS), three times for 10 minutes each at 45°C in a second wash buffer (0.1X 25 SSC), and dried.

Detection

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Reporter-labeled hybridization complexes are detected with a microscope equipped with an Innova 70 mixed gas 10 W laser (Coherent, Inc., Santa Clara CA) capable of generating spectral lines at 488 nm for excitation of Cy3 and at 632 nm for excitation of Cy5. The excitation laser light 30 is focused on the array using a 20X microscope objective (Nikon, Inc., Melville NY). The slide containing the array is placed on a computer-controlled X-Y stage on the microscope and raster-scanned past the objective. The 1.8 cm x 1.8 cm array used in the present example is scanned with a resolution of 20 micrometers.

In two separate scans, a mixed gas multiline laser excites the two fluorophores sequentially.

35 Emitted light is split, based on wavelength, into two photomultiplier tube detectors (PMT R1477,

Hamamatsu Photonics Systems, Bridgewater NJ) corresponding to the two fluorophores.

Appropriate filters positioned between the array and the photomultiplier tubes are used to filter the signals. The emission maxima of the fluorophores used are 565 nm for Cy3 and 650 nm for Cy5.

Each array is typically scanned twice, one scan per fluorophore using the appropriate filters at the 5 laser source, although the apparatus is capable of recording the spectra from both fluorophores simultaneously.

The sensitivity of the scans is typically calibrated using the signal intensity generated by a cDNA control species added to the sample mixture at a known concentration. A specific location on the array contains a complementary DNA sequence, allowing the intensity of the signal at that 10 location to be correlated with a weight ratio of hybridizing species of 1:100,000. When two samples from different sources (e.g., representing test and control cells), each labeled with a different fluorophore, are hybridized to a single array for the purpose of identifying genes that are differentially expressed, the calibration is done by labeling samples of the calibrating cDNA with the two fluorophores and adding identical amounts of each to the hybridization mixture.

The output of the photomultiplier tube is digitized using a 12-bit RTI-835H analog-to-digital (A/D) conversion board (Analog Devices, Inc., Norwood MA) installed in an IBM-compatible PC computer. The digitized data are displayed as an image where the signal intensity is mapped using a linear 20-color transformation to a pseudocolor scale ranging from blue (low signal) to red (high signal). The data is also analyzed quantitatively. Where two different 20 fluorophores are excited and measured simultaneously, the data are first corrected for optical crosstalk (due to overlapping emission spectra) between the fluorophores using each fluorophore's emission spectrum.

A grid is superimposed over the fluorescence signal image such that the signal from each spot is centered in each element of the grid. The fluorescence signal within each element is then 25 integrated to obtain a numerical value corresponding to the average intensity of the signal. The software used for signal analysis is the GEMTOOLS gene expression analysis program (Incyte).

XI. Complementary Polynucleotides

Sequences complementary to the KAP-encoding sequences, or any parts thereof, are used to detect, decrease, or inhibit expression of naturally occurring KAP. Although use of 30 oligonucleotides comprising from about 15 to 30 base pairs is described, essentially the same procedure is used with smaller or with larger sequence fragments. Appropriate oligonucleotides are designed using OLIGO 4.06 software (National Biosciences) and the coding sequence of KAP. To inhibit transcription, a complementary oligonucleotide is designed from the most unique 5' sequence and used to prevent promoter binding to the coding sequence. To inhibit translation, a 35 complementary oligonucleotide is designed to prevent ribosomal binding to the KAP-encoding

transcript.

XII. Expression of KAP

systems. For expression of KAP in bacteria, cDNA is subcloned into an appropriate vector 5 containing an antibiotic resistance gene and an inducible promoter that directs high levels of cDNA transcription. Examples of such promoters include, but are not limited to, the *trp-lac* (*tac*) hybrid promoter and the T5 or T7 bacteriophage promoter in conjunction with the *lac* operator regulatory element. Recombinant vectors are transformed into suitable bacterial hosts, e.g., BL21(DE3). Antibiotic resistant bacteria express KAP upon induction with isopropyl beta-D-

Expression and purification of KAP is achieved using bacterial or virus-based expression

- 10 thiogalactopyranoside (IPTG). Expression of KAP in eukaryotic cells is achieved by infecting insect or mammalian cell lines with recombinant <u>Autographica californica</u> nuclear polyhedrosis virus (AcMNPV), commonly known as baculovirus. The nonessential polyhedrin gene of baculovirus is replaced with cDNA encoding KAP by either homologous recombination or bacterial-mediated transposition involving transfer plasmid intermediates. Viral infectivity is
- 15 maintained and the strong polyhedrin promoter drives high levels of cDNA transcription.

 Recombinant baculovirus is used to infect Spodoptera frugiperda (Sf9) insect cells in most cases, or human hepatocytes, in some cases. Infection of the latter requires additional genetic modifications to baculovirus. (See Engelhard, E.K. et al. (1994) Proc. Natl. Acad. Sci. USA 91:3224-3227; Sandig, V. et al. (1996) Hum. Gene Ther. 7:1937-1945.)
- In most expression systems, KAP is synthesized as a fusion protein with, e.g., glutathione Stransferase (GST) or a peptide epitope tag, such as FLAG or 6-His, permitting rapid, single-step, affinity-based purification of recombinant fusion protein from crude cell lysates. GST, a 26-kilodalton enzyme from Schistosoma japonicum, enables the purification of fusion proteins on immobilized glutathione under conditions that maintain protein activity and antigenicity (Amersham
- 25 Pharmacia Biotech). Following purification, the GST moiety can be proteolytically cleaved from KAP at specifically engineered sites. FLAG, an 8-amino acid peptide, enables immunoaffinity purification using commercially available monoclonal and polyclonal anti-FLAG antibodies (Eastman Kodak). 6-His, a stretch of six consecutive histidine residues, enables purification on metal-chelate resins (QIAGEN). Methods for protein expression and purification are discussed in
- 30 Ausubel (1995, <u>supra</u>, ch. 10 and 16). Purified KAP obtained by these methods can be used directly in the assays shown in Examples XVI, XVII, XVIII, XIX, XX, and XXI where applicable.

XIII. Functional Assays

KAP function is assessed by expressing the sequences encoding KAP at physiologically elevated levels in mammalian cell culture systems. cDNA is subcloned into a mammalian 35 expression vector containing a strong promoter that drives high levels of cDNA expression. Vectors

of choice include PCMV SPORT (Life Technologies) and PCR3.1 (Invitrogen, Carlsbad CA), both of which contain the cytomegalovirus promoter. 5-10 μ g of recombinant vector are transiently transfected into a human cell line, for example, an endothelial or hematopoietic cell line, using either liposome formulations or electroporation. 1-2 μ g of an additional plasmid containing 5 sequences encoding a marker protein are co-transfected. Expression of a marker protein provides a means to distinguish transfected cells from nontransfected cells and is a reliable predictor of cDNA expression from the recombinant vector. Marker proteins of choice include, e.g., Green Fluorescent Protein (GFP; Clontech), CD64, or a CD64-GFP fusion protein. Flow cytometry (FCM), an automated, laser optics-based technique, is used to identify transfected cells expressing GFP or 10 CD64-GFP and to evaluate the apoptotic state of the cells and other cellular properties. FCM detects and quantifies the uptake of fluorescent molecules that diagnose events preceding or coincident with cell death. These events include changes in nuclear DNA content as measured by staining of DNA with propidium iodide; changes in cell size and granularity as measured by forward light scatter and 90 degree side light scatter; down-regulation of DNA synthesis as measured by 15 decrease in bromodeoxyuridine uptake; alterations in expression of cell surface and intracellular proteins as measured by reactivity with specific antibodies; and alterations in plasma membrane composition as measured by the binding of fluorescein-conjugated Annexin V protein to the cell surface. Methods in flow cytometry are discussed in Ormerod, M.G. (1994) Flow Cytometry, Oxford, New York NY.

The influence of KAP on gene expression can be assessed using highly purified populations of cells transfected with sequences encoding KAP and either CD64 or CD64-GFP. CD64 and CD64-GFP are expressed on the surface of transfected cells and bind to conserved regions of human immunoglobulin G (IgG). Transfected cells are efficiently separated from nontransfected cells using magnetic beads coated with either human IgG or antibody against CD64 (DYNAL, Lake Success NY). mRNA can be purified from the cells using methods well known by those of skill in the art. Expression of mRNA encoding KAP and other genes of interest can be analyzed by northern analysis or microarray techniques.

XIV. Production of KAP Specific Antibodies

KAP substantially purified using polyacrylamide gel electrophoresis (PAGE; see, e.g., 30 Harrington, M.G. (1990) Methods Enzymol. 182:488-495), or other purification techniques, is used to immunize rabbits and to produce antibodies using standard protocols.

Alternatively, the KAP amino acid sequence is analyzed using LASERGENE software (DNASTAR) to determine regions of high immunogenicity, and a corresponding oligopeptide is synthesized and used to raise antibodies by means known to those of skill in the art. Methods for selection of appropriate epitopes, such as those near the C-terminus or in hydrophilic regions are

well described in the art. (See, e.g., Ausubel, 1995, supra, ch. 11.)

Typically, oligopeptides of about 15 residues in length are synthesized using an ABI 431A peptide synthesizer (Applied Biosystems) using FMOC chemistry and coupled to KLH (Sigma-Aldrich, St. Louis MO) by reaction with N-maleimidobenzoyl-N-hydroxysuccinimide ester (MBS) to increase immunogenicity. (See, e.g., Ausubel, 1995, supra.) Rabbits are immunized with the oligopeptide-KLH complex in complete Freund's adjuvant. Resulting antisera are tested for antipeptide and anti-KAP activity by, for example, binding the peptide or KAP to a substrate, blocking with 1% BSA, reacting with rabbit antisera, washing, and reacting with radio-iodinated goat anti-rabbit IgG.

10 XV. Purification of Naturally Occurring KAP Using Specific Antibodies

Naturally occurring or recombinant KAP is substantially purified by immunoaffinity chromatography using antibodies specific for KAP. An immunoaffinity column is constructed by covalently coupling anti-KAP antibody to an activated chromatographic resin, such as CNBr-activated SEPHAROSE (Amersham Pharmacia Biotech). After the coupling, the resin is blocked and washed according to the manufacturer's instructions.

Media containing KAP are passed over the immunoaffinity column, and the column is washed under conditions that allow the preferential absorbance of KAP (e.g., high ionic strength buffers in the presence of detergent). The column is eluted under conditions that disrupt antibody/KAP binding (e.g., a buffer of pH 2 to pH 3, or a high concentration of a chaotrope, such 20 as urea or thiocyanate ion), and KAP is collected.

XVI. Identification of Molecules Which Interact with KAP

KAP, or biologically active fragments thereof, are labeled with ¹²⁵I Bolton-Hunter reagent. (See, e.g., Bolton, A.E. and W.M. Hunter (1973) Biochem. J. 133:529-539.) Candidate molecules previously arrayed in the wells of a multi-well plate are incubated with the labeled KAP, washed, 25 and any wells with labeled KAP complex are assayed. Data obtained using different concentrations of KAP are used to calculate values for the number, affinity, and association of KAP with the candidate molecules.

Alternatively, molecules interacting with KAP are analyzed using the yeast two-hybrid system as described in Fields, S. and O. Song (1989) Nature 340:245-246, or using commercially 30 available kits based on the two-hybrid system, such as the MATCHMAKER system (Clontech).

KAP may also be used in the PATHCALLING process (CuraGen Corp., New Haven CT) which employs the yeast two-hybrid system in a high throughput manner to determine all interactions between the proteins encoded by two large libraries of genes (Nandabalan, K. et al. (2000) U.S. Patent No. 6,057,101).

35 XVII. Demonstration of KAP Activity

Generally, protein kinase activity is measured by quantifying the phosphorylation of a protein substrate by KAP in the presence of [γ-³²P]ATP. KAP is incubated with the protein substrate, ³²P-ATP, and an appropriate kinase buffer. The ³²P incorporated into the substrate is separated from free ³²P-ATP by electrophoresis and the incorporated ³²P is counted using a 5 radioisotope counter. The amount of incorporated ³²P is proportional to the activity of KAP. A determination of the specific amino acid residue phosphorylated is made by phosphoamino acid analysis of the hydrolyzed protein.

In one alternative, protein kinase activity is measured by quantifying the transfer of gamma phosphate from adenosine triphosphate (ATP) to a serine, threonine or tyrosine residue in a protein substrate. The reaction occurs between a protein kinase sample with a biotinylated peptide substrate and gamma ³²P-ATP. Following the reaction, free avidin in solution is added for binding to the biotinylated ³²P-peptide product. The binding sample then undergoes a centrifugal ultrafiltration process with a membrane which will retain the product-avidin complex and allow passage of free gamma ³²P-ATP. The reservoir of the centrifuged unit containing the ³²P-peptide product as retentate is then counted in a scintillation counter. This procedure allows the assay of any type of protein kinase sample, depending on the peptide substrate and kinase reaction buffer selected. This assay is provided in kit form (ASUA, Affinity Ultrafiltration Separation Assay, Transbio Corporation, Baltimore MD, U.S. Patent No. 5,869,275). Suggested substrates and their respective enzymes include but are not limited to: Histone H1 (Sigma) and p34^{ede2}kinase, Annexin I, 20 Angiotensin (Sigma) and EGF receptor kinase, Annexin II and *src* kinase, ERK1 & ERK2 substrates and MEK, and myelin basic protein and ERK (Pearson, J.D. et al. (1991) Methods Enzymol. 200:62-81).

In another alternative, protein kinase activity of KAP is demonstrated in an assay containing KAP, 50 μl of kinase buffer, 1 μg substrate, such as myelin basic protein (MBP) or synthetic 25 peptide substrates, 1 mM DTT, 10 μg ATP, and 0.5 μCi [γ-³²P]ATP. The reaction is incubated at 30°C for 30 minutes and stopped by pipetting onto P81 paper. The unincorporated [γ-³²P]ATP is removed by washing and the incorporated radioactivity is measured using a scintillation counter. Alternatively, the reaction is stopped by heating to 100°C in the presence of SDS loading buffer and resolved on a 12% SDS polyacrylamide gel followed by autoradiography. The amount of 30 incorporated ³²P is proportional to the activity of KAP.

In yet another alternative, adenylate kinase or guanylate kinase activity of KAP may be measured by the incorporation of ³²P from [γ-³²P]ATP into ADP or GDP using a gamma radioisotope counter. KAP, in a kinase buffer, is incubated together with the appropriate nucleotide mono-phosphate substrate (AMP or GMP) and ³²P-labeled ATP as the phosphate donor. The 35 reaction is incubated at 37°C and terminated by addition of trichloroacetic acid. The acid extract is

neutralized and subjected to gel electrophoresis to separate the mono-, di-, and triphosphonucleotide fractions. The diphosphonucleotide fraction is excised and counted. The radioactivity recovered is proportional to the activity of KAP.

In yet another alternative, other assays for KAP include scintillation proximity assays 5 (SPA), scintillation plate technology and filter binding assays. Useful substrates include recombinant proteins tagged with glutathione transferase, or synthetic peptide substrates tagged with biotin. Inhibitors of KAP activity, such as small organic molecules, proteins or peptides, may be identified by such assays.

In another alternative, phosphatase activity of KAP is measured by the hydrolysis of para10 nitrophenyl phosphate (PNPP). KAP is incubated together with PNPP in HEPES buffer pH 7.5, in
the presence of 0.1% β-mercaptoethanol at 37°C for 60 min. The reaction is stopped by the addition
of 6 ml of 10 N NaOH (Diamond, R.H. et al. (1994) Mol. Cell. Biol. 14:3752-62). Alternatively,
acid phosphatase activity of KAP is demonstrated by incubating KAP-containing extract with 100
μl of 10 mM PNPP in 0.1 M sodium citrate, pH 4.5, and 50 μl of 40 mM NaCl at 37°C for 20 min.
15 The reaction is stopped by the addition of 0.5 ml of 0.4 M glycine/NaOH, pH 10.4 (Saftig, P. et al.

(1997) J. Biol. Chem. 272:18628-18635). The increase in light absorbance at 410 nm resulting from the hydrolysis of PNPP is measured using a spectrophotometer. The increase in light absorbance is proportional to the activity of KAP in the assay.

In the alternative, KAP activity is determined by measuring the amount of phosphate 20 removed from a phosphorylated protein substrate. Reactions are performed with 2 or 4 nM KAP in a final volume of 30 μl containing 60 mM Tris, pH 7.6, 1 mM EDTA, 1 mM EGTA, 0.1% β-mercaptoethanol and 10 μM substrate, ³²P-labeled on serine/threonine or tyrosine, as appropriate. Reactions are initiated with substrate and incubated at 30° C for 10-15 min. Reactions are quenched with 450 μl of 4% (w/v) activated charcoal in 0.6 M HCl, 90 mM Na₄P₂O₇, and 2 mM NaH₂PO₄,

25 then centrifuged at $12,000 \times g$ for 5 min. Acid-soluble ³²Pi is quantified by liquid scintillation counting (Sinclair, C. et al. (1999) J. Biol. Chem. 274:23666-23672).

XVIII. Kinase Binding Assay

Binding of KAP to a FLAG-CD44 cyt fusion protein can be determined by incubating KAP with anti-KAP-conjugated immunoaffinity beads followed by incubating portions of the beads 30 (having 10-20 ng of protein) with 0.5 ml of a binding buffer (20 mM Tris-HCL (pH 7.4), 150 mM NaCl, 0.1% bovine serum albumin, and 0.05% Triton X-100) in the presence of ¹²⁵I-labeled FLAG-CD44cyt fusion protein (5,000 cpm/ng protein) at 4 °C for 5 hours. Following binding, beads were washed thoroughly in the binding buffer and the bead-bound radioactivity measured in a scintillation counter (Bourguignon, L.Y.W. et al. (2001) J. Biol. Chem. 276:7327-7336). The 35 amount of incorporated ³²P is proportional to the amount of bound KAP.

XIX. Identification of KAP Inhibitors

Compounds to be tested are arrayed in the wells of a 384-well plate in varying concentrations along with an appropriate buffer and substrate, as described in the assays in Example XVII. KAP activity is measured for each well and the ability of each compound to inhibit KAP activity can be determined, as well as the dose-response kinetics. This assay could also be used to identify molecules which enhance KAP activity.

XX. Identification of KAP Substrates

A KAP "substrate-trapping" assay takes advantage of the increased substrate affinity that may be conferred by certain mutations in the PTP signature sequence of protein tyrosine

10 phosphatases. KAP bearing these mutations form a stable complex with their substrate; this complex may be isolated biochemically. Site-directed mutagenesis of invariant residues in the PTP signature sequence in a clone encoding the catalytic domain of KAP is performed using a method standard in the art or a commercial kit, such as the MUTA-GENE kit from BIO-RAD. For expression of KAP mutants in Escherichia coli, DNA fragments containing the mutation are

15 exchanged with the corresponding wild-type sequence in an expression vector bearing the sequence encoding KAP or a glutathione S-transferase (GST)-KAP fusion protein. KAP mutants are expressed in E. coli and purified by chromatography.

The expression vector is transfected into COS1 or 293 cells via calcium phosphate-mediated transfection with 20 μg of CsCl-purified DNA per 10-cm dish of cells or 8 μg per 6-cm dish. Forty-20 eight hours after transfection, cells are stimulated with 100 ng/ml epidermal growth factor to increase tyrosine phosphorylation in cells, as the tyrosine kinase EGFR is abundant in COS cells. Cells are lysed in 50 mM Tris·HCl, pH 7.5/5 mM EDTA/150 mM NaCl/1% Triton X-100/5 mM iodoacetic acid/10 mM sodium phosphate/10 mM NaF/5 μg/ml leupeptin/5 μg/ml aprotinin/1 mM benzamidine (1 ml per 10-cm dish, 0.5 ml per 6-cm dish). KAP is immunoprecipitated from lysates with an appropriate antibody. GST-KAP fusion proteins are precipitated with glutathione-Sepharose, 4 μg of mAb or 10 μl of beads respectively per mg of cell lysate. Complexes can be visualized by PAGE or further purified to identify substrate molecules (Flint, A.J. et al. (1997) Proc. Natl. Acad. Sci. USA 94:1680-1685).

XXI. Enhancement/Inhibition of Protein Kinase Activity

Agonists or antagonists of KAP activation or inhibition may be tested using assays described in section XVII. Agonists cause an increase in KAP activity and antagonists cause a decrease in KAP activity.

Various modifications and variations of the described methods and systems of the invention 35 will be apparent to those skilled in the art without departing from the scope and spirit of the

invention. Although the invention has been described in connection with certain embodiments, it should be understood that the invention as claimed should not be unduly limited to such specific embodiments. Indeed, various modifications of the described modes for carrying out the invention which are obvious to those skilled in molecular biology or related fields are intended to be within 5 the scope of the following claims.

Table 1

Incote Project ID	Polypeptide	Incyte	Polynucleotide	Incyte
	SEQ ID NO:	Polypeptide ID	SEQ ID NO:	Polynucleotide
				3
4615110		4615110CD1	21	4615110CB1
4622230	2	4622229CD1	22	4622229CB1
77358203	3 (2)	72358203CD1	23	72358203CB1
4885040	4	4885040CD1	24	4885040CB1
7484507	v	7484507CD1	25	7484507CB1
7108031	9	7198931CD1	26	7198931CB1
7482905	7	7482905CD1	27	7482905CB1
7483019	. ∞	7483019CD1	28	7483019CB1
5455490	6	5455490CD1	29	5455490CB1
5547067	10	5547067CD1	30	5547067CB1
71675660	11	71675660CD1	31	71675660CB1
71678683	12	71678683CD1	32	71678683CB1
7474567	13	7474567CD1	33	7474567CB1
3838946	14	3838946CD1	34	3838946CB1
72001176	15	72001176CD1	35	72001176CB1
55064363	16	55064363CD1	36	55064363CB1
7482044	17	7482044CD1	37	7482044CB1
7476595	18	7476595CD1	38	7476595CB1
71824382	19	71824382CD1	39	71824382CB1
3566882	20	3566882CD1	40	3566882CB1
20000				

Poly SEQ	Polyneptide SEQ ID NO:	Incyte Polypeptide ID	Incyte Polypeptide GenBank ID NO: Probability ID or PROTEOME Score ID NO:	Probability Score	Annotation
-		4615110CD1	g3598974	0	[Rattus norvegicus] protein tyrosine phosphatase TD14. Cao, L. et al. (1998) J. Biol Chem 273:21077-21083
2		4622229CD1	g4079673	0	myotubularin related 1 [Homo sapiens]. Kioschis, P. et al. (1998) Genomics
					54:256-266
3		72358203CD1	g7768151	6.40E-17	Protein phpsphatase 2C (PP2C) [Fagus sylvatica].
4		4885040CD1	g6468206	1.20E-119	[Mus musculus] thiamin pyrophosphokinase. Nosaka, K. et al. (1999) J. Biol. Chem. 274:34129-34133
2		7484507CD1	g7649810	7.20E-14	[Homo sapiens] protein kinase PAK5
9		7198931CD1	g2815888	0	[Homo sapiens] MEK kinase 1. Xia, Y. et al. (1998) Genes Dev. 12:3369-3381
7		7482905CD1	g256855	2.10E-161	[Mus sp.] serine/threonine- and tyrosine-specific protein kinase, Nek1=NIMA cell cycle regulator homolog. Letwin, K., et al. (1992) EMBO J. 11:3521-3531
∞		7483019CD1	g6552404	8.40E-197	[Rattus norvegicus] DLG6 alpha. Inagaki, H. et al. (1999) Biochem. Biophys.
					INCS: COMMITTEE SOCIALS TOO
6		5455490CD1	g406058	0	protein kinase [Mus musculus]. (Walden, P.D. and Cowan, N.J. (1993) Mol. Cell. Biol. 13: 7625-7635)
10		5547067CD1	g1033033	5.90E-41	ribosomal S6 kinase [Homo sapiens]. (Zhao, Y. et al. (1995) Mol. Cell. Biol. 15: 4353-4363)
111		71675660CD1	g2738898	9.40E-175	protein kinase [Mus musculus]. (Kueng,P. et al. (1997) J. Cell Biol. 139: 1851- 1859)
12		71678683CD1	g2738898	4.00E-174	protein kinase [Mus musculus]. (Kueng,P. et al. (1997) J. Cell Biol. 139: 1851-1859)
13		7474567CD1	g6723964	2.50E-72	putative serine/threonine protein kinase [Schizosaccharomyces pombe]
14		3838946CD1	g4982155	2.80E-53	glycerate kinase, putative [Thermotoga maritima]. (Nelson, K.E. et al. (1999) Nature 399: 323-329)
15		72001176CD1	g11177010	5.70E-232	casein kinase 1 gamma 1L [Homo sapiens]

Table 2

	Wano X S et al.	11 au 6, 13.5. 5. a.	1	NT (1000) A Mound	N.J. (1993) A INOVELASSOCIATED With 13, 7625-7635	
Annotation) [Supplementation of III and saniens]	Mitogen-activated kinase kinase of filolilo sapicus) (17 alis) 2 cm. 171.31607-31611)	J) J. DIOI. CHOIN 4/ 1:3100/ 3101/	Mitogen-activated protein kinase kinase kinase i Homo sapicins	[Mus musculus] protein kinase. Walden, P.D. and Cowan, N.J. (1993) A NOVEL 205-kDa Testis-specific Serine/Threonine Protein Kinase Associated with Microtubules of the Spermatid Manchette. Mol. Cell. Biol. 13, 7625-7635	
		Mito	(122	Mito	[Mus 205- Micr	-
Probal Score		<u> </u>		0	0	_
GenBank ID NO: Probat or PROTEOME Score ID NO:		g1679668		g11527775	g406058	
ncyte Polypeptide D		55064363CD1		7482044CD1		
Polypeptide SEQ ID NO:		16		17	18	

CEO	Though	-	Amino Acid	Dotential	Dotential	Signature Sequences Domains and Motife	Analytical Methods and
	Polypeptide ID	E E	Residues	돗	ation Sites Glycosylation Sites		Databases
1	4615110QD1		1636	S86 S101 S136 S193 S275 S311 S429 S455 S487 S546 S645 S869 S1056 S1122 S1218 S1231 S1238 S1247 S1290 S1322 S1342 S1475 S1506 S1533 S1575 S1593 S1625 T95 T293 T352 T434 T450 T486 T511 T882 T1068 T1144 T1269 T1305 T1328 T1354 Y272 Y320	N652 N1245 N1634	Protein-tyrosine phosphatase: Y1217-R1451	HMMER_PFAM
						Tyrosine specific protein phosphatases proteins BL00383: K1220-V1234, D1241-V1249, D1272-V1282, H1349-P1361, V1390-G1400, R1429-F1444	BLIMPS_BLOCKS
				,		Tyrosine specific protein phosphatases signature and PROFILESCAN profiles: L1367-M1428	PROFILESCAN
						stase signature PR00700: 1279, R1345-D1362, P1387- 11435-C1445	BLIMPS_PRINTS
						PROTEIN TYROSINE PHOSPHATASE TD14 EC BLAST_PRODOM 3.1.3.48 HYDROLASE PD180360: F967-L1219	BLAST_PRODOM

ds and	M	M						
Analytical Methods and Databases	BLAST_PRODO	BLAST_PRODO	BLAST_DOMO	BLAST_DOMO	BLAST_DOMO	BLAST_DOMO	MOTIFS	TMAP
Signature Sequences, Domains and Motifs	PROTEIN TYROSINE PHOSPHATASE TD14 EC BLAST_PRODOM 3.1.3.48 HYDROLASE PD184907: K713-G952	PROTEIN TYROSINE PHOSPHATASE TD14 EC BLAST_PRODOM 3.1.3.48 HYDROLASE PD169419: A1567-T1636	PROTEIN-TYROSINE-PHOSPHATASE DM00089[P17706[4-277: K1220-V1450	PROTEIN-TYROSINE-PHOSPHATASE	PROTEIN-TYROSINE-PHOSPHATASE DM00089 P29074 641-914: K1220-Q1455	PROTEIN-TYROSINE-PHOSPHATASE DM00089 P43378 285-577: K1220-Q1455	Tyrosine specific protein phosphatases active site: V1390-F1402	Transmembrane domains: W517-S543; N-terminus is cytosolic
Potential Glycosylation Sites								113 S163 S172 N78 N251 N359 \$253 S261 \$342 S354 \$402 S410 \$525 S575 \$654 S656 T334 T358 T476 T536 Y400 Y563
Potential Potential Phosphorylation Sites								S53 S113 S163 S172 S225 S253 S261 S278 S342 S354 S391 S402 S410 S437 S525 S575 S600 S654 S656 T136 T334 T358 T470 T476 T536 Y331 Y400 Y563
cid								673
Incyte Amino A Polypeptide ID Residues								4622229CD1
SEQ T	NO.							2

SEO	Incyte	Amino Acid		Potential	Signature Sequences. Domains and Motifs	Analytical Methods and
e ë	ID Polypeptide ID NO:		Phosphory	lation Sites Glycosylation Sites		Databases
2 cont		·			Tyrosine specific protein phosphatases proteins BL00383: W570-D578, Q511-R521, V444-A454	BLMPS_BLOCKS
					Tyrosine specific protein phosphatases signature and PROFILESCAN profiles: L424-K480	PROFILESCAN
					HYDROLASE PROTEIN MYOTUBULARIN DISEASE MUTATION F53A2.8 PROTEIN TYROSINE PHOSPHATASE C19A8.03	BLAST_PRODOM
					CPAZNNF1 PD014611; C1/8-Y3/2, D304-H391	
					MYOTUBULARIN DISEASE MUTATION HYDROLASE PD144999: H601-T671	BLAST_PRODOM
					Tyrosine specific protein phosphatases active site: V444-L456	MOTIFS
6	72358203CD	459	S50, T257, T278, S306, T364, S430, S438		Protein phosphatase 2C: Q326-K415, L187-L265	HMMER-PFAM
					Protein phosphatase 2C: BL01032: Y120-G129, L187-G204, G214-S223, N232-E271, R328-D341, D376-D388	BLIMPS-BLOCKS
					PROTEIN PHOSPHATASE 2C MAGNESIUM HYDROLASE MANGANESE MULTIGENE FAMILY PP2C ISOFORM: PD001101: G322- L403, Y120-D289	BLAST-PRODOM

Table 3

					Gomesine and Motifs	Analytical Methods and
18 e	SEQ Incyte Amino Av D Polypeptide ID Residues	cid	Potential Potential Phosphorylation Sites Clycosylation Sites Phosphorylation Sites Phosphorylation Sites Potential Potent	Potential Glycosylation Sites	Signature Sequences, Domains and recess	Databases
ö	•				PROTECTIVE DELOCEDITATA SE 20.	BLAST-DOMO
3					PROTEIN FROM THE TANK TOOL TOO. R328-S456, DM00377[P49596]1-295. A191-I262, R328-S456,	
cont					Y120-E149	
	1.400	cro	74 CO2 TF T56	N203	Ribokinase signature PR00990 V121-F132	BLIMPS_PRINTS
4	4885040CD1	243	T176		CU MOUT PRINT	
					THIAMIN PYROPHOSPHOKINASE PUTALIVE THIK KINASE, PD106295; H170-M239;	BLASI_TRODOM
					PD036502: L21-Q144	
					Endeamotic protein kinase domain: V55-L173, W201 HMMER_PFAM	HIMMER_PFAM
5	7484507CD1	632	S6 S20 S114 S212	N208	Eukar your provin America Comments	
			S283 S300 S318			
			S504 S575 S587			
			S601 S607 T12 T183			
			T258 T269 T287			
			1338 1410		Transmembrane domains: E421-N448 M472-G487, TMAP	TMAP
					N terminus cytosolic	Cuth
					Tyrosine kinase catalytic domain PROO109, Y147- BLIMPS_PRINTS	- BLIMPS_PRINTS
					L165, F197-L207, S215-E237	MOUDO SOUTH
					PHOSPHORYLASE KINASE ALF PD01841: L422/BLLKU S_1XCD CST.	
				-	L438, A404-1303, G307 13003, 225 27-17-18-18-18-18-18-18-18-18-18-18-18-18-18-	
				-		

Analytical Methods and	Databases	BLAST_DOMO	HMMER_PFAM	ТМАР	PROFILESCAN
Signature Sequences, Domains and Motifs	•	PROTEIN KINASE DOMAIN DM00004; P51955 10-261: V30-M233; S43968 28-311: Q33- K289, R271-I288 A55480 28-320: Q33-K289, R271- L297; P49186 28-320: Q33-K289, R271-L297	Eukaryotic protein kinase domain: W1242-F1507	Transmembrane domains: S348-L368, A1392-L1420; N-terminus is cytosolic	Protein kinases signatures and profile: V1344-G1398
Potential	Glycosylation Sites		N346 N540 N744 N806 N1068 N1085 N1099 N1128 N1278 N1347		
Potential	ylation Sites		S35 S118 S232 S258 N346 N540 N744 S275 S281 S300 N806 N1068 N10 S394 S397 S398 N1099 N1128 N1 S429 S434 S507 N1347 S514 S531 S588 N1347 S669 S782 S816 S823 S900 S923 S823 S900 S923 S928 S1025 S1038 S1087 S1088 S1129 S1130 S1281 T20 T169 T261 T304 T379 T457 T657 T705 T911 T946 T996 T1020 T1069 T1113 T1147 T1165 T1279 Y1166		
Amino Acid			1511		
Though	Polypeptide ID		719893 CD1		
נבט		5 cont	9		

Analytical Methods and Databases	Out there you a re-	BLIMPS_PRINTS	BLAST_PRODOM		BLAST_PRODOM			ON COLUMN TO A TEL	BLAS1_DOMO		BLAST_DOMO	חזיטת דיז א זת	DLASI_DOMO	BLAST_DOMO		MOTIFS		MOTIFS		
Signature Sequences, Domains and Motifs		Tyrosine kinase catalytic domain signature PR00109: L1476-S1498, Y1358-I1376, G1410- L1420, C1429-E1451		PROTEIN ATP BINDING PHOSPHORY LATION PD144583: M1-E601	MAPK/ERK KINASE 1 EC 2.7.1. MEK MEKK	PROTEIN ATP BINDING PHOSPHORYLATION	PD146039: Q624-Q1247		PROTEIN KINASE DOMAIN	DIMINUOU4/173349/403-036. 1X1244-01476	PROTEIN KINASE DOMAIN	DIVIDUOU4 A46064 76-546. DAZ44-10175	PROTEIN KINASE DOMAIN DM00004 001389 1176-1430: L1243-P1496	PROTEIN KINASE DOMAIN	DM00004 Q1040/ 826-1084; L1243-L1466	Protein kinases ATP-binding region signature: I1248 MOTIFS	K1271	Serine/Threonine protein kinases active-site	signature: 11364-11376	
Potential	Glycosylation office																			
Potential Potential Potential Physical Sites	Fnospnoryianou ones																			
çid	Kesidues																			
Incyte Amino A	Polypeptide ID																			
SEQ	A Š	cont cont																		

	its Analytical Methods and Databases		SPSCAN	ASE NEK1 BLAST_PRODOM 1 NSIS LL CYCLE 44030: M1-	HMMER_PFAM	HMMER_PFAM	BLIMPS_BLOCKS	2130, D132- BLIMPS_PRINTS
	Signature Sequences, Domains and Motifs		signal_cleavage: M1-S54	SERINE/THREONINE PROTEIN KINASE NEK1 EC 2.7.1. NIMA RELATED PROTEIN 1 TRANSFERASE ATP BINDING MITOSIS NUCLEAR PHOSPHORYLATION CELL CYCLE DIVISION TYROSINE PROTEIN PD144030: M1-1.394	Guanylate kinase: T281-Y385	PDZ domain: I3-V83	Guanylate kinase protein BL00856:	SH3 domain signature PR00452: A115-Q130, D132-BLIMPS_PRINTS
	Potential ation Sites Glycosylation Sites				N419		•	
	Potential Phosphorylation Sites		S54 S179 S260 S279 N159 N303 N401 S280 S327 S352 N540 N715 S370 S378 S440 S457 S525 S545 S580 S624 S664 S698 S708 S741 S747 T267 T354 T358 T403 T481 T490 T512 T634		S142 S200 S208 S242 S308 S374 S421 S450 T16 T280 T283 Y307 Y317 Y359			
	Incyte Amino Acid Polypepide ID Residues		830		455			
-	e epide D		7482905CD1		7483019CD1			_
			74825		74830		<u> </u>	
	SEQ 19	ö	7		∞			

Table 3

						1	<u>-</u>	\neg
Analytical Methods and Databases	BLAST_PRODOM	BLAST_PRODOM	BLAST_PRODOM	BLAST_DOMO	BLAST_DOMO	BLAST_DOMO	BLAST_DOMO	MOTIFS
Signature Sequences, Domains and Motifs	PROTEIN DOMAIN SH3 KINASE GUANYLATE BLAST_PRODOM TRANSFERASE ATP BINDING REPEAT GMP MEMBRANE PD001338: T280-Q373	PROTEIN MAGUK P55 SUBFAMILY MEMBER BLAST_PRODOM MPP3 DISCS LARGE HOMOLOG SH3 PD090357: P169-T280	PROTEIN MAGUK P55 SUBFAMILY MEMBER BLAST_PRODOM DISCS LARGE HOMOLOG SH3 DOMAIN	GUANYLATE KINASE DM00755 A57653 370- 570: P241-P444	GUANYLATE KINASE DM00755 P54936 769- 955: R246-K372, M388-P444	GUANYLATE KINASE DM00755 138757 709- 898: R246-P444	GUANYLATE KINASE DM00755 P31007 765- 954: R246-P444	Guanylate kinase signature: T280-V297
Potential Glycosylation Sites								
Potential Potential Phosphorylation Sites								
몆								
SEQ Incyte Amino Ac ID Polypeptide ID Residues								
SEQ ID	NO:							

Table 3

Analytical Methods and	Databases	NA COLO	HMMER
Signature Sequences, Domains and Motifs		Signal Peptide: M1-Soo	Signal Peptide: M31-S56
Potential		NIII) NII/4 NIZIS	
Potential	ylation Sites	S75 S82 S86 S113 S119 S140 S152 S175 S203 S402 S425 S430 S455 S697 S728 S733 S739 S747 S768 S776 S782 S796 S831 S836 S853 S106 S1022 S1117 S1127 S1136 S1147 S1127 S1152 S1178 S1194 S1254 S1259 S1340 S1347 S1351 S1369 S1381 S1413 S1525 S1426 S1463 S1572 S1579 S1582 S1593 S1620 S1639 S1593 T188 T428 T436 T487 T503 T737 T793 T838 T436 T487 T503 T1311 T1158 T1166 T1346 T1402 T1597	
Amino Acid		1.720	
Theyte	Polypeptide ID	5455490CD	
SFO		5	

Table 3

				<u> </u>				
Analytical Methods and Databases	HMMER_PFAM	HMMER_PFAM	TMAP	PROFILESCAN	BLIMPS_PRINTS	BLAST_PRODOM	BLAST_DOMO	MOTFS
Signature Sequences, Domains and Motifs	PDZ domain (or DHR, or GLGF): P1026-L1113	Enformatic protein kingse domain: F434-F707		Protein kinases signatures and profile: F501-I581	Tyrosine kinase catalytic domain sig. PR00109: M411-K524, Y547-IS65, V628-D650	MICROTUBULE ASSOCIATED TESTIS SPECIFIC SERINE/ITREONINE KINASE PD142315: H1235-T1720; PD182663: E785- H1061; PD135564: C83-Y242; PD041650: K243- D433	PROTEIN KINASE DOMAIN: DM00004 A54602 455-712: T436-G694; DM08046 P05986 1-397: S430-K580; DM00004 S42867 75-498: I437-T588; DM00004 S42864 41-325: E435-K580, T695	Serine/Threonine protein kinases active-site signature: 1553-1565
Potential Glycosylation Sites								
Potential Potential Potential Physical	Condon							
Amino Acid	Residuce	·						
Incyte Amino Ao	Folypeptide LD							
SEQ	a Ö	cont						

Analytical Methods and	Databases	HMMER_PFAM	MAP	ROFILESCAN	BLIMPS_PRINTS	BLAST_DOMO	MOTIFS	HMMER_PFAM	MAP	PROFILESCAN
Simanure Sequences Domains and Motifs A		Eukaryotic protein kinase domain: L146-F398 H	Transmembrane domains: S244-R267, D324-P341; TMAP N terminus is cytosolic.	Protein kinases signatures and profile: F248-A297 PROFILESCAN	Tyrosine kinase catalytic domain signature, PR00109: Y258-L276, G304-L314, A323-E345	PROTEIN KINASE DOMAIN: DM00004 A53300 64-305: L146-L386; DM08046 P06244 1-396: Q144-F435; DM00004 A57459 61-302: L146-L386; DM00004 S56639 153-391: I148-L386	Serine/Threonine protein kinases active-site M signature: 1264-L276	Bukaryotic protein kinase domain: Y12-L272 H	Transmembrane domain: V196-M224; N terminus TMAP is non-cytosolic.	Protein kinases signatures and profile: D111-S165 PI
Dotantial										
Dotontial	ylation Sites	S17 S45 S89 S107 S208 S244 S358 S425 T86 T167 T187 T337 T356						S31 S158 S258 S284 N240 S349 T48 T340 Y293		
-	Residues	1 449						01 358		
-	Incyte Polypeptide ID	5547067CD1						71675660CD1		
•		10						11		

Analytical Methods and Databases	BLIMPS_PRINTS	BLAST_PRODOM	BLAST_DOMO	MOTIFS	MOTIFS	HIMMER_PFAM	TMAP	PROFILESCAN	BLIMPS_PRINTS
Signature Sequences, Domains and Motifs	Tyrosine kinase catalytic domain signature: PR00109: M90-K103, Y126-L144, L241-l263	ONINE 029090: L272-	PROTEIN KINASE DOMAIN: DM00004 P27448 58-297: L18-L.253; DM00004 JC1446 20-261: V14-I263; DM00004 S24578 18-262: V14-I263;	Serine/Threonine protein kinases active-site signature: 1132-L.144	Protein kinases ATP-binding region signature: L18-MOTIFS K41	Eukaryotic protein kinase domain: Y12-L272	Transmembrane domain: V196-M224; N terminus TMAP is non-cytosolic.	Protein kinases signatures and profile: D111-S165 PROFILESCAN	Tyrosine kinase catalytic domain signature, PR00109: M90-K103, Y126-L144, G177-L187, Y197-S219, L241-I263
Potential Glycosylation Sites						N240			
Potential Potential Phosphorylation Sites						S31 S158 S258 S284 N240 S349 T48 T340	1,293		
Amino Acid Residues						358			
Incyte Polyneptide ID						71678683CD1			
SEQ T	ÖZ	cont				12			

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Analytical Methods and	Databases	BLAST_PRODOM	BLAST_DOMO	MOTIFS	MOTIFS	HMMER_PFAM	BLIMPS_PRINTS	TMAP
G: Company of the com	olgnature oequences, Donianis and Motto	TESTIS SPECIFIC SERINE/ THREONINE KINASE 2 PROTEIN KINASE, PD029090: L272- T358	PROTEIN KINASE DOMAIN: DM00004 P27448 58-297; L18-L253; DM00004 JC1446 20-261; V14-I263; DM00004 S24578 18-262; V14-I263; DM00004 I48609 55-294; L18-R260	Serine/Threonine protein kinases active-site signature: 1132-L144	Protein kinases ATP-binding region signature: L18-MOTIFS K41	N51 N187 N630 N726 Eukaryotic protein kinase domain: L159-F327, F32-HMMER_PFAM N768 N916 H106	Tyrosine kinase catalytic domain signature, PR00109: L168-L186, S247-V269, I296-A318	Transmembrane domain: E163-L183, N-terminus TMAP is non-cytosolic
	Potential Glycosylation Sites					N51 N187 N630 N726 N768 N916		N487
	Potential Phosphorylation Sites					S56 S85 S171 S207 S483 S660 S677 T53 T57 T245 T313 T401 T440 T555 T608 T658 T679 T712 T722 T737 T760 T765		S283 S289 S367 S417 T166 T191 T208 T214 Y328
	Incyte Amino Acid Polypeptide ID Residues					929		523
	tide ID					CD .		CD1
	Incyte Polypep					7474567CD1		3838946CD1
Г	S 日 S	12 cont				13		14

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Analytical Methods and	Databases	D BLAST_PRODOM	HMMER_PFAM	s TMAP	3 PROFILESCAN	BLAST_PRODOM	BLAST_DOMO	0- MOTIFS	MOTIFS
Signature Sequences, Domains and Motifs		HYDROXYPYRUVATE REDUCTASE PLASMID BLAST_PRODOM OXIDOREDUCTASE NADP PROTEIN GLYCERATE KINASE, PD014236: K131-T357, T357-L520	Eukaryotic protein kinase domain: F44-E276	Transmembrane domain: D133-I161 N-terminus is	cytosolic. Protein kinases signatures and profile: T140-E198	CASEIN KINASE I, GAMMA I ISOFORM EC 2.7.1. GAMMA TRANSFERASE SERINE/THREONINE ATP BINDING MULTIGENE FAMILY PHOSPHORYLATION; PD049080: M1-N43, PD015080: F315-W379	PROTEIN KINASE DOMAIN: DM00004 A56711 46-303: V46-Y304; DM00004 C56711 45-301: V46-Y304; DM00004 B56711 48-303: V46-Y304;	Protein kinases ATP-binding region signature: 150-	Serine/Threonine protein kinases active-site
Dotential	Glycosylation Sites		N370 N388						
	Potential Phosphorylation Sites Glycosylation Sites		124 S150 S229 T14 T137 T199 T258 T269 T355 T411	T454					
	Amino Acid Residues		459						
	Incyte Polypeptide ID		72001176CD1						
Г	SEQ D	NO: 14 cont	15						

		-	- 1			4	A 1 (1 1 1 1 1 1 1 1 1 1 1 1 1 1 1 1 1
SEQ	Incyte		Amino Acid	Potential	Potential	Signature Sequences, Domains and Motifs	Analytical Methods and
д ë	Polypeptide ID		Residues	Phosphorylation Sites Glycosylation Sites	Glycosylation Sites		Databases
16	55064363CD1		1360	S23 S56 S212 S253 S338 S382 S432 S486 S550 S609 S625 S632 S655 S677 S762 S843 S934 S991 S1025 S1031 S1040 S1041 S1056 S1084 T48 T205 T218 T428 T466 T545 T685 T796 T842 T887 T893 T945 T983 T1234 T1237 T1313	N381 N620	Eukaryotic protein kinase domain: V704-L955	НММЕК-РҒАМ
						Transmembrane domains:S445-T466, S1129-V1146: N-terminus is cytosolic	TMAP
		1				Protein kinases signature: T796-G848	ProfileScan
						gnature:L705-	MOTIFS
						Serine/Threonine protein kinases active-site signature:1816-V828	MOTIFS
						Tyrosine kinase catalytic domain signature PR00109:M773-R786, Y810-V828, G858- I868,A879-L901, L924-T946	BLIMPS-PRINTS
						Kinase, apoptosis, ASK1, MEK signal-regulating, mitogen-activated, MEKK5, MAP/ERK, MAPKKK5 PD018410:V75-N620	BLAST-PRODOM
		F					

Analytical Methods and Databases	BLAST-PRODOM	BLAST-PRODOM	BLAST-PRODOM	BLAST-DOMO	HMMER-PFAM	
Signature Sequences, Domains and Motifs	Kinase, apoptosis, ASK1, MEK signal-regulating, mitogen-activated, MEKK5, MAP/ERK, MAPKKK5 PD014104:P982-G1205	Kinase, apoptosis, ASK1, MEK signal-regulating, mitogen-activated, MEKK5, MAP/ERK, MAPKKK5 PD024456:E1215-R1348	Kinase, apoptosis, ASK1, MEK signal-regulating, mitogen-activated, MEKK5, MAP/ERK, MAPKKK5 PD012471:F621-D697	Protein kinase domains: DM00004 A48084 98-348: V704-R943; DM00004 Q01389 1176-1430: V704- T945; DM00004 Q10407 826-1084: V704-T945; DM00004 P41892 11-249: L705-T946	Eukaryotic protein kinase domain:L181-F439	
Potential Glycosylation Sites						
Potential Potential Phosphorylation Sites					S31 S35 S191 S250 S323 S338 S517 S600 S625 S1131 S1160 S1165 T67	T136 T154 T174 T203 T218 T268 T333 T396 T459 T492 T1161 T1201 T1231 T1251 T1273 T1294 Y428
Amino Acid Residues					1345	
Incyte Polypeptide ID					7482044CD1	
SEQ ID	NO: 16 cont				17	

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Analytical Methods and	Databases	TMAP		ProfileScan	MOTIFS		MOTIFS	BLAST-DOMO							
Signature Sequences Domains and Motifs		Transmembrane domain: A868-A890, N-terminus is TMAP	cytosolic	Protein kinases signature: L284-F339	Serine/Threonine protein kinases active-site	signature:1305-1317	Leucine zipper pattern: L826-L847	Protein kinase domains: DM00004 A48084 98-348: BLAST-DOMO	V704-R943; DM00004 Q01389 1176-1430: V704-	T945; DM00004 Q10407 826-1084: V704-T945;	DM00004 P41892 11-249: L705-	T946,DM00004 P51957 8-251: L187-R427,	DM00004 P41892 11-249: L187-V395,	DM00004 Q05609 553-797: E186-C419	
Potential	Glycosylation Sites														
	rylation Sites														
Amino Acid Potential	D Polypeptide ID Residues NO:														
	ptide ID														
Thouse	Polype														
QHO	d a g	17	cont			_									

Table (

0	Incute	Amino Acid	Potential	Potential	Signature Sequences, Domains and Motifs	Analytical Methods and	
日	ptide ID	Residues	Phosphorylation Sites Glycosylation Sites	Glycosylation Sites		Databases	
ÖN				OF LIVE COLLA SECTION SECTION	PFAM or GI GFI. 0555 HMMER PFAM	HIMMER PFAM	
18	7476595CD1	2038	S18 S28 S324 S329	NI6 N645 N/03 N/40 N1266 N1282 N1473	FDZ domain (Auso Known as Dink of Greek): \$555.		
			S333 S303 S407 S448 S536 S562	C 1 1 1 2 2 1 1 1 2 2 1 1 1 1 1 1 1 1 1			
			S647 S657 S666				
			S669 S674 S680				
			S707 S721 S728				
			S731 S780 S785				
			S871 S878 S882				
			S895 S903 S930				
			S938 S974 S1000				
			S1007 S1027 S1073				
			S1109 S1182 S1199				
			S1231 S1262 S1270				
			S1278 S1305 S1340				
			S1389 S1398 S1514				
			S1517 S1574 S1583				
			S1590 S1606 S1629				
			S1650 S1660 S1745				
			S1863 S1879 S1899				
			S1913 S1938 S1960				
			S2028 T32 T83 T99				
			T247 T333 T343				
	_		T349 T435 T465			•	
			T511 T569 T641				
			T695 T886 T1059				
			T1079 T1177 T1184				
			T1321 T1327 T1395				
			T1407 T1420 T1436				
			T1554 T1692 T1753				
			T1769 T1780 T1790				
			T1844 T1931 T1971				
			T2006 Y1794				٦.

OED)	1203.40		Amino Apid	Dotantial	Potential	Signature Sequences Domains and Motifs	Analytical Methods and
) (1)	Polypeptide ID			rylation Sites	Glycosylation Sites		Databases
ö				•			
18						Eukaryotic protein kinase domain: F30-F303	HMMER_PFAM
1103						TMAP: D225-F243; N-terminus is cytosolic	TMAP
						F97-	PROFILESCAN
						VI//	
						Tyrosine kinase catalytic domain signature	BLIMPS_PRINTS
		·				PR00109: M10/-K120, X 143-V 161, V 224-D246, P269-7291	
						MICROTUBILE ASSOCIATED TESTIS	BLAST PRODOM
						SPECIFIC SERINE/THREONINE PROTEIN	1
						KINASE 205KD TESTISSPECIFIC	
						SERINE/THREONINE PROTEIN KINASE	
						MAST205 KINASE, PD142315: H760-A1021,	
						P1578-P1716, P1498-P1609, PD069998: T639-	
						D734, PD182663: E499-N591	
						PROTEIN KINASE SERINE/THREONINE KIN4	BLAST_PRODOM
						MICROTUBULE ASSOCIATED TESTIS	
						SPECIFIC TESTISSPECIFIC MAST205,	
						PD040805: L306-N374	
						PROTEIN KINASE DOMAIN;	BLAST_DOMO
						DM00004 A54602 455-712: T32-G290;	
			:			DM00004 S42867 75-498: I33-K176, H190-F331;	
						DM08046 P05986 1-397: S28-K176, V203-D351;	
						DM08046 P06244 1-396: D29-K176, V203-F354	
						ATP/GTP-binding site motif A (P-loop): A1450-	MOTIFS
						T1457	

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Table 3

Analytical Methods and Databases	TFS	HMMER_PFAM	HMMER_PFAM	HMMER_PFAM	HMMER_PFAM	HMMER_PFAM
Anal Data	MOTIFS	THE STATE OF THE S	HI			
Signature Sequences, Domains and Motifs	Serine/Threonine protein kinases active-site signature: 1149-V161	CNH domain: K1266-K1550	Phorbol esters/diacylglycerol binding domain: H1051-C1100	PH domain: T1121-K1239	Eukaryotic protein kinase domain: F77-F343	Protein kinase C terminal domain: S344-D372
Potential Glycosylation Sites		N560 N792 N854 N1680 N1739 N1742				·
Potential Potential Phosphorylation Sites Glycosylation Sites		S167 S286 S344 S364 S369 S411 S459 S475 S507 S555 S616 S705 S750 S752 S781 S813 S877 S884 S917 S926 S940 S977 S997 S1013 S1193 S1322 S1334 S1357 S1457 S1568 S1583 S1658 S1673 S1694 S1702 S1731 S1751 T30 T64 T423 T591 T624 T691 T746 T780 T788 T959 T1011 T1032 T1050 T1121 T1223 T1293 T1543 T1763				
Amino Acid Residues		1770				
Incyte Amino A Polypeptide ID Residues		71824382CD1				
i	NO:	19				

				0.44.41.1	G. Daniel Commencer Daniel Detific	Analytical Mathode and
our Oes	cyte		Potential	rotential	orginalme oequerices, Domains and Mours	Aliai yticai Michigus and
& gÿ	Polypeptide ID	Residues	Phosphorylation Sites Glycosylation Sites	Glycosylation Sites		Databases
19 cont					Phorbol esters / diacylglycerol binding domain dag_pe_binding_domain: C1064-A1122	PROFILESCAN
					Tyrosine kinase catalytic domain signature PR00109: M154-S167, S191-M209, C263-E285	BLIMPS_PRINTS
-					Domain found in NIK1-lik	BLIMPS_PFAM
		-			PF00780B: I738-T780	
			-		PF00780F: T1050-A1096	
					PF00780G: K1195-H1238	
					PF00780I: M1485-N1514	
-					MYTONIC DYSTROPHY KINASE-RELATED	BLAST_PRODOM
					CDC42-BINDING KINASE PHORBOLESTER	
_					BINDING KIAA0451 PROTEIN PD143271:	
					R1643-P1770	
					MYTONIC DYSTROPHY KINASE-RELATED	BLAST_PRODOM
					CDC42-BINDING KINASE PHOKBOLESTEK	
					BLNDLING FD0/3023: E030-IN/13	
-					PHORBOLESTER BINDING KINASE	BLAST_PRODOM
			-		DYSTROPHY KINASE-RELATED CDC42-	
					BINDING SIMILAR SERINE/THREONINE	
					PROTEIN GENGHIS KHAN PD150840: W1518-	
					S1642	-
					PHORBOLESTER BINDING DYSTROPHY	BLAST_PRODOM
		-			KINASE-RELATED CDC42-BINDING KINASE	
					GENGHIS KHAN MYTONIC MYOTONIC POULL POULL BOOK 1959: D833-F967	
-					10011606: 2000 1000	

Table 3

is and				_																		SI	
Analytical Methods and Databases		BLAST_DOMO					MOTIFS		MOTIFS	MOTIFS		MOTIFS		MOTIFS		HMMER_PFAM			THAMED DEAM	TIVILVILLA LA	TMAP	BLIMPS_PRINTS	
Signature Sequences, Domains and Motifs			Q09013 83-336: 179-Q331; S42867 75-498: 179-	L226, V238-Y404, P1653-D1728; I38133 90-369:	E78-L226, V238-G330, P53894 353-658: L80-	G221, D205-Q331	Leucine zipper pattern L772-L793 L779-L800 L786-MOTIFS	L807	C-type lectin domain signature C1067-C1088	Phorbol esters / diacylglycerol binding domain	H1051-C1100	Protein kinases ATP-binding region signature 183-	K106	Serine/Threonine protein kinases active-site	signature Y197-M209	Ank repeat: E448-R480, D382-R414, V580-Q612,	E415-A447, N481-Q513, S349-E381, Q547-A579,	S613-K645, V646-G678	י ככת אויי	Eukaryotic protein kinase domain: \$150-F251	Transmembrane domain: S146-Y171	Tyrosine kinase catalytic domain signature	PR00109: M94-S107, L152-L174, E211-F233
Potential	horylation Sites Glycosylation Sites				•																		
Potential	Phosphorylation Sites															S91 S117 S146 S148	S264 S268 S299	S690 S697 T17 T166	T398 Y314				
ligi Sign	Residues															720	277	-					
Incyte	Polypeptide ID Residues															10000000	3300002						
SEO	, П	ÖN	19	cont												6	₹						

oide O:/ Sequence	Sequence Fragments
OCB1/5200	1-224, 1-277, 4-272, 14-161, 14-225, 42-679, 43-503, 43-609, 43-708, 43-714, 43-872, 48-688, 124-438, 178-4215, 199-420, 200-720, 240-549, 352-679, 355-637, 355-756, 371-754, 374-992, 446-992, 459-1093, 506-1102, 545-827, 564-824, 763-1296, 825-1296, 869-1296, 870-1296, 131-1857, 1049-1527, 1063-1697, 1098-1689, 1103-1774, 1133-1736, 1250-1743, 1250-1768, 1250-1840, 1312-1857, 1376-1857, 1416-1857, 1426-1857, 1426-1857, 1426-1857, 1426-1857, 1426-1857, 1496-2036, 1508-1998, 1515-2107, 1554-2211, 1635-2249, 1713-2241, 1716-2315, 1728-2380, 1775-2322, 1796-2438, 1809-2049, 2006-5055, 2020-2679, 2029-2385, 2056-2732, 2069-2702, 2107-2752, 2186-2443, 2196-2638, 2231-2698, 2271-2775, 2287-2580, 2302-2741, 2335-2806, 2407-2857, 2409-2669, 2432-2980, 2796-2997, 2810-3016, 2824-2994, 2950-3400, 3029-3604, 3029-3684, 3064-3648, 3100-3372, 3139-3684, 3186-3766, 3194-3457, 3212-3473, 3219-3456, 3228-3737, 3234-3704, 3236-3485, 3236-3719, 3245-3503, 3273-3887, 3295-3689, 3317-3583, 3317-3604, 3317-3939, 3341-3634, 3351-3979, 3357-3615, 3375-3621, 3396-3971, 3428-4081, 3454-4060,

3479-4086, 3488-4156, 3491-3759, 3511-3828, 3511-3977, 3540-3825, 3540-3985, 3540-4047, 3548-3834, 3550-4216, 3580-3916, 3590-3928, 3599-4202, 3611-4211, 3627-4351, 3629-4099, 3629-4339, 3630-3907, 3630-4382, 3634-4382, 3641-4215, 3645-3920, 3649-3932, 3649-3933, 3650-3889, 3651-3904, 3654-4181, 3654-4215, 3660-4212, 3662-4080, 3664-4226, 3667-4162, 3667-4210, 3672-4212, 3675-4215, 3683-4211, 3693-4230, 3704-4211, 3706-4173, 3712-4215, 3728-4215, 3729-4215, 3730-4214, 3735-4214, 3737-4112, 3748-4213, 3752-4575, 3755-4025, 3766-4216, 3770-4382, 3771-4382, 3774-4215,

Polynucleo SEQ ID No

21/461511

Incyte ID/ Length

Polynucleotide SEQ ID NO:/	Sequence Fragments
Incyte ID/ Sequence	
Length	222 1.00 2781 1216 2702 1215 2781 1215 2786-4015 3786-4216 3791-4211, 3795-4211, 3796-4215, 3796-4216,
	3/10-4194, 3/01-4210, 3/02-4213, 3/04-4215, 3809-4197, 3810-4144, 3817-4215, 3821-4112, 3821-4152, 3833-4162,
	3803-4050, 3603-4101, 3850-4145, 3852-4205, 3852-4215, 3854-4442, 3858-4165, 3863-4121, 3876-4442, 3884-4139,
	3885-4382, 3888-4216, 3905-4380, 3941-4382, 3947-4215, 4013-4562, 4081-4243, 4171-4645, 4178-4610, 4194-4692,
	4194-4697, 4194-4698, 4194-4699, 4194-4749, 4194-4780, 4194-4904, 4194-4933, 4207-4496, 4208-4470, 4208-4486,
	4208-4492, 4208-4493, 4208-4496, 4208-4525, 4208-4644, 4208-4680, 4208-4683, 4208-4687, 4208-4691, 4208-4694,
	4208.4702, 4208-4707, 4210-4526, 4211-4496, 4211-4680, 4215-4496, 4216-4496, 4217-4480, 4222-4496, 4241-4382,
	4241, 4257-4542, 4252-4612, 4257-4522, 4257-4534, 4257-4541, 4257-4542, 4257-4545, 4257-4562, 4291-4701,
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Table 5

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26	7198931CB1	SYNORAB01
27	7482905CB1	BMARTXE01
28	7483019CB1	BMARTXT02
29	5455490CB1	HNT2AGT01
30	5547067CB1	BRAIFEE05
31	71675660CB1	TESTNOT17
32	71678683CB1	TESTNOT17
33	7474567CB1	UCMCNOT02
34	3838946CB1	NOSEDIN01
35	72001176CB1	THP1NOT03
36	55064363CB1	BRAIFET02
37	7482044CB1	BRAUNOR01
39	71824382CB1	BRABDIR01
40	3566882CB1	LUNLTUE02

I :hromy	Vector	Library Description
TXE01		This 5' biased random primed library was constructed using RNA isolated from treated SH-SY3Y cells derived from a metastatic bone marrow neuroblastoma, removed from a 4-year-old Caucasian female (Schering AG). The medium was MEM/HAM'S F12 with 10% fetal calf serum. After reaching about 80% confluency cells were treated with 6-Hydroxydopamine (6-OHDA) at 100 microM for 8 hours.
BMARTXT02	pINCY	Library was constructed using RNA isolated from treated SH-SY5Y cell line derived from bone marrow neuroblastoma tumor cells removed from a 4-year-old Caucasian female. The cells were cultured in the presence of retinoic acid.
BRABDIR01	pINCY	Library was constructed using RNA isolated from diseased cerebellum tissue removed from the brain of a 2/-year-old Caucasian male, who died from a cerebrovascular accident. Patient history included Huntington's disease, emphysema, and tobacco abuse.
BRAIFEE05	PCDNA2.1	This 5' biased random primed library was constructed using KNA Isolated from the fetus who was stillborn with a hypoplastic left heart at 23 weeks' gestation.
BRAIFEN08	pINCY	This normalized fetal brain tissue library was constructed from 400 mousand morpolated four stillborn with a library. Starting RNA was made from brain tissue removed from a Caucasian male fetus who was stillborn with a hypoplastic left heart at 23 weeks' gestation. The library was normalized in 2 rounds using conditions adapted from Soares et al., PNAS (1994) 91:9228 and Bonaldo et al., Genome Research (1996) 6:791, except that a significantly longer (48 hours/round) reannealing hybridization was used.
BRAIFET02	pINCY	Library was constructed using RNA isolated from brain tissue removed from a Caucasian male fetus, who was stillborn with a hypoplastic left heart at 23 weeks' gestation.
BRAINON01	PSPORT1	Library was constructed and normalized from 4.88 million independent clones from the Dryan constructed and normalized from a 26-year-old Caucasian male during cranioplasty and excision of a cerebral made from brain tissue removed from a 26-year-old Caucasian male during cranioplasty and excision of a cerebral meningeal lesion. Pathology for the associated tumor tissue indicated a grade 4 oligoastrocytoma in the right fronto-parietal part of the brain.
BRAITUT03	PSPORT1	Library was constructed using RNA isolated from brain tunor ussue removed from the fibrillary giant and small-Caucasian female during excision of a cerebral meningeal lesion. Pathology indicated a grade 4 fibrillary giant and small-cell astrocytoma. Family history included benign hypertension and cerebrovascular disease.

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Library BRAUNOR01 BRAYIIIN03 BRAYIIIN03	NOR01	Vector Librar pINCY This re tissue at there at the from an infarct with a remain the from a strocy in the pippoc hemos cholan hypert hypotlan pINCY This mande for cerebrar rounds 6:791, and rec	Library Description This random printed library was constructed using RNA isolated from striatum, globus pallidus and posterior putamen itssue removed from an 81-year-old Caucasian female who died from a hemorrhage and ruptured thoracic aorta due to atherosclerosis. Pathology indicated moderate atherosclerosis involving the internal carotids, bilaterally; microscopic infarcts of the frontal cortex and hippocampus; and scattered diffuse amyloid plaques and ruptured thoracic aorta due to infarcts of the frontal cortex and hippocampus; and scattered diffuse amyloid plaques and neurofibrillary tangles, consistent with age. Grossly, the leptomeninges showed only mild thickening and hyalinization along the superior sagittal situs. The remainder of the leptomeninges sun temporal lobes, bilaterally. Microscopically, there were pairs of Alzheimer type II astrocytes within the deep layers of the neocortex. There was increased satellitosis around neurons in the deep gray matter the frontal poles and lobes, and temporal lobes, bilaterally. Microscopically, there were pairs of Alzheimer type II astrocytes within the deep layers of the neocortex. The aniygdala contained rare diffuse plaques and neurofibrillary tangles. The posterior hippocampus contained a microscopic area of cystic cavitation with hemosiderin laden macrophages surrounded by reactive gliosis. Patient history included sepsis, cholonagitis, post-operative atelectasis, pneumonia CAD, cardiomegaly due to left ventricular hypothyroidism, and peripheral vascular disease. This normalized library was constructed from 6.7 million independent clones from a brain tissue library. Starting RNA was made from RNA isolated from diseased hypothalamus tissue removed from a 57-year-old Caucasian male who died from a cerebrovascular accident. Patient history included Huntington's disease and emphysema. The library was normalized and recircularized to select for insert containing clones. Library was constructed using RNA isolated from aortic endothelial cell tissue from an ex
			male during a heart transplant.
HNT2AGT01	GT01	PBLUESCRIPT	PBLUESCRIPT Library was constructed at Stratagene (STR937233), using RNA isolated from the hNT2 cell line derived from a human teratocarcinoma that exhibited properties characteristic of a committed neuronal precursor. Cells were treated with retinoic acid for 5 weeks and with mitotic inhibitors for two weeks and allowed to mature for an additional 4 weeks in conditioned medium.

Table (

T : honery	Vector	Library Description
LUNITUE02	42.1	This 5' biased random primed library was constructed using RNA isolated from left upper lobe lung tumor tissue removed from a 56-year-old Caucasian male during complete pneumonectomy, pericardectomy and regional lymph node excision. Pathology indicated grade 3 squamous cell carcinoma forming a mass in the left upper lobe centrally. The tumor extended through pleura into adjacent pericardium. Patient history included hemoptysis and tobacco abuse. Family history included benign hypertension, cerebrovascular accident, atherosclerotic coronary artery disease in the mother; prostate cancer in the father; and type II diabetes in the sibling(s).
NOSEDINO1	pINCY	This normalized nasal polyp tissue library was constructed from 1.08 million independent clones from a pooled nasal polyp tissue library. Starting RNA was made from pooled cDNA from two donors. cDNA was generated using mRNA isolated from a nasal polyp removed from a 78-year-old Caucasian male during nasal polypectomy (donor A) and from nasal polyps from a nother donor (donor B). Pathology (A) indicated a nasal polyp and striking eosinophilia, especially deep in the epithelium. In many instances, eosinophils were undergoing frank necrosis with striking deposition of Charcot-Leyden crystals. Foci of eosinophil infiltration in small islands of cells were seen in certain areas, and those areas closer to the appearance surface were losing definition and evidently undergoing necrosis. Examination of respiratory epithelium showed loss of surface epithelium in many areas, and there was a tendency for cells to aggregate around the epithelium. This nasal polyp showed typical histology for polypoid change associated with allergic disease. Patient history included asthma, allergy tests (which were positive for histamine but negative for common substances), a pulmonary function test (PFT, which showed reduction in the forced expiratory volume (FEV), with increase after use of a bronchodilator), and nasal polyps. Patient history was not history (A) included asthma. Previous surgery (A) included a nasal polypectomy. The patient was not using glucocorticoids in treatment for asthma. The library was normalized in 1 round using conditions adapted from Soares et al., PNAS (1994) 91:9228-9232 and Bonaldo et al., Genome Research 6 (1996):791, except that a significantly longer (48 hours/round)
SYNORAB01	PBLUESCRIPT	Library was constructed using RNA isolated from the synovial membrane tissue of a 68-year-old Caucasian female with rheumatoid arthritis.

Library		Vector	Library Description
TESTN	IESTNOT17 pINCY	pINCY	Library was constructed from testis tissue removed from a 26-year-old Caucasian male who died from head trauma due to a motor vehicle accident. Serologies were negative. Patient history included a hernia at birth, tobacco use (1 1/2 ppd), marijuana use, and daily alcohol use (beer and hard liquor).
THP1N	THP1NOT03 pINCY	pINCY	Library was constructed using RNA isolated from untreated THP-1 cells. THP-1 is a human promonocyte line derived from the peripheral blood of a 1-year-old Caucasian male with acute monocytic leukemia (ref: Int. J. Cancer (1980) 26:171).
UCMC	UCMCNOT02 pINCY	pINCY	Library was constructed using RNA isolated from mononuclear cells obtained from the umbilical cord blood of nine individuals.

Program	Description	Reference	Parameter Threshold
ABIFACTURA	A program that removes vector sequences and masks ambiguous bases in nucleic acid sequences.	Applied Biosystems, Foster City, CA.	
ABIPARACEL FDF	A Fast Data Finder useful in comparing and annotating amino acid or nucleic acid sequences.	Applied Biosystems, Foster City, CA; Paracel Inc., Pasadena, CA.	Mismatch <50%
ABI AutoAssembler	A program that assembles nucleic acid sequences.	Applied Biosystems, Foster City, CA.	
BLAST	A Basic Local Alignment Search Tool useful in sequence similarity search for amino acid and nucleic acid sequences. BLAST includes five functions: blastp, blastn, blastx, tblastn, and tblastx.	Altschul, S.F. et al. (1990) J. Mol. Biol. 215:403-410; Altschul, S.F. et al. (1997) Nucleic Acids Res. 25:3389-3402.	ESTs: Probability value= 1.0E-8 or less Full Length sequences: Probability value= 1.0E-10 or less
FASTA	A Pearson and Lipman algorithm that searches for similarity between a query sequence and a group of sequences of the same type. FASTA comprises as least five functions: fasta, tfasta, fastx, tfastx, and ssearch.	Pearson, W.R. and D.J. Lipman (1988) Proc. Natl. Acad Sci. USA 85:2444-2448; Pearson, W.R. (1990) Methods Enzymol. 183:63-98; and Smith, T.F. and M.S. Waterman (1981) Adv. Appl. Math. 2:482-489.	ESTs: fasta E value=1.06E-6 Assembled ESTs: fasta Identity= 95% or greater and Match length=200 bases or greater; fastx E value=1.0E-8 or less Full Length sequences: fastx score=100 or greater
BLIMPS	A BLocks IMProved Searcher that matches a sequence against those in BLOCKS, PRINTS, DOMO, PRODOM, and PFAM databases to search for gene families, sequence homology, and structural fingerprint regions.	Henikoff, S. and J.G. Henikoff (1991) Nucleic Acids Res. 19:6565-6572; Henikoff, J.G. and S. Henikoff (1996) Methods Enzymol. 266:88-105; and Attwood, T.K. et al. (1997) J. Chem. Inf. Comput. Sci. 37:417-424.	Probability value= 1.0E-3 or less
HMMER	An algorithm for searching a query sequence against hidden Markov model (HMM)-based databases of protein family consensus sequences, such as PFAM.	Krogh, A. et al. (1994) J. Mol. Biol. 235:1501-1531; Sonnhammer, E.L.L. et al. (1988) Nucleic Acids Res. 26:320-322; Durbin, R. et al. (1998) Our World View, in a Nutshell, Cambridge Univ. Press, pp. 1-350.	PFAM hits: Probability value=1.0E-3 or less Signal peptide hits: Score=0 or greater

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	Table / (colle.)	(COIII.)	
Program	Description	Reference	Parameter Threshold
ProfileScan	An algorithm that searches for structural and sequence motifs in protein sequences that match sequence patterns defined in Prosite.	Gribskov, M. et al. (1988) CABIOS 4:61-66; Gribskov, M. et al. (1989) Methods Enzymol. 183:146-159; Bairoch, A. et al. (1997) Nucleic Acids Res. 25:217-221.	Normalized quality scores GCG-specified "HIGH" value for that particular Prosite motif. Generally, score=1.4-2.1.
Phred	A base-calling algorithm that examines automated sequencer traces with high sensitivity and probability.	Ewing, B. et al. (1998) Genome Res. 8:175-185; Ewing, B. and P. Green (1998) Genome Res. 8:186-194.	
Phrap	A Phils Revised Assembly Program including SWAT and CrossMatch, programs based on efficient implementation of the Smith-Waterman algorithm, useful in searching sequence homology and assembling DNA sequences.	Smith, T.F. and M.S. Waterman (1981) Adv. Appl. Math. 2:482-489; Smith, T.F. and M.S. Waterman (1981) J. Mol. Biol. 147:195-197; and Green, P., University of Washington, Seattle, WA.	Score= 120 or greater; Match length= 56 or greater
Consed	A graphical tool for viewing and editing Phrap assemblies.	Gordon, D. et al. (1998) Genome Res. 8:195-202.	72.
SPScan	A weight matrix analysis program that scans protein sequences for the presence of secretory signal peptides.	Nielson, H. et al. (1997) Protein Engineering 10:1-6; Claverie, J.M. and S. Audic (1997) CABIOS 12:431-439.	Score=3.5 or greater
TMAP	A program that uses weight matrices to delineate transmembrane segments on protein sequences and determine orientation.	Persson, B. and P. Argos (1994) J. Mol. Biol. 237:182-192; Persson, B. and P. Argos (1996) Protein Sci. 5:363-371.	
TMHMMER	A program that uses a hidden Markov model (HMM) to delineate transmembrane segments on protein sequences and determine orientation.	Sonnhammer, E.L. et al. (1998) Proc. Sixth Intl. Conf. on Intelligent Systems for Mol. Biol., Glasgow et al., eds., The Am. Assoc. for Artificial Intelligence Press, Menlo Park, CA, pp. 175-182.	2.
Motifs	A program that searches amino acid sequences for patterns that matched those defined in Prosite.	Bairoch, A. et al. (1997) Nucleic Acids Res. 25:217-221; Wisconsin Package Program Manual, version 9, page M51-59, Genetics Computer Group, Madison, WI.	:217-221; , page WI.

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What is claimed is:

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- 1. An isolated polypeptide selected from the group consisting of:
- a) a polypeptide comprising an amino acid sequence selected from the group consisting of SEQ ID NO:1-20,
- a polypeptide comprising a naturally occurring amino acid sequence at least 90% identical to an amino acid sequence selected from the group consisting of SEQ ID NO:1-20,
- c) a biologically active fragment of a polypeptide having an amino acid sequence selected from the group consisting of SEQ ID NO:1-20, and
- d) an immunogenic fragment of a polypeptide having an amino acid sequence selected from the group consisting of SEQ ID NO:1-20.
- An isolated polypeptide of claim 1 comprising an amino acid sequence selected from the group consisting of SEQ ID NO:1-20.
 - 3. An isolated polynucleotide encoding a polypeptide of claim 1.
 - 4. An isolated polynucleotide encoding a polypeptide of claim 2.
 - 5. An isolated polynucleotide of claim 4 comprising a polynucleotide sequence selected from the group consisting of SEQ ID NO:21-40.
- A recombinant polynucleotide comprising a promoter sequence operably linked to a polynucleotide of claim 3.
 - 7. A cell transformed with a recombinant polynucleotide of claim 6.
 - 8. A transgenic organism comprising a recombinant polynucleotide of claim 6.
 - 9. A method of producing a polypeptide of claim 1, the method comprising:
 - a) culturing a cell under conditions suitable for expression of the polypeptide, wherein said cell is transformed with a recombinant polynucleotide, and said recombinant polynucleotide comprises a promoter sequence operably linked to a polynucleotide encoding the polypeptide of claim 1, and

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- b) recovering the polypeptide so expressed.
- 10. A method of claim 9, wherein the polypeptide comprises an amino acid sequence selected from the group consisting of SEQ ID NO:1-20.

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- 11. An isolated antibody which specifically binds to a polypeptide of claim 1.
- 12. An isolated polynucleotide selected from the group consisting of:
- a) a polynucleotide comprising a polynucleotide sequence selected from the group consisting of SEQ ID NO:21-40,
- b) a polynucleotide comprising a naturally occurring polynucleotide sequence at least 90% identical to a polynucleotide sequence selected from the group consisting of SEQ ID NO:21-40,
- c) a polynucleotide complementary to a polynucleotide of a),
- d) a polynucleotide complementary to a polynucleotide of b), and
 - e) an RNA equivalent of a)-d).
 - 13. An isolated polynucleotide comprising at least 60 contiguous nucleotides of a polynucleotide of claim 12.

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- 14. A method of detecting a target polynucleotide in a sample, said target polynucleotide having a sequence of a polynucleotide of claim 12, the method comprising:
- a) hybridizing the sample with a probe comprising at least 20 contiguous nucleotides comprising a sequence complementary to said target polynucleotide in the sample, and which probe specifically hybridizes to said target polynucleotide, under conditions whereby a hybridization complex is formed between said probe and said target polynucleotide or fragments thereof, and
- b) detecting the presence or absence of said hybridization complex, and, optionally, if present, the amount thereof.

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- 15. A method of claim 14, wherein the probe comprises at least 60 contiguous nucleotides.
- 16. A method of detecting a target polynucleotide in a sample, said target polynucleotide having a sequence of a polynucleotide of claim 12, the method comprising:
- a) amplifying said target polynucleotide or fragment thereof using polymerase chain

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- reaction amplification, and
- b) detecting the presence or absence of said amplified target polynucleotide or fragment thereof, and, optionally, if present, the amount thereof.
- 5 17. A composition comprising a polypeptide of claim 1 and a pharmaceutically acceptable excipient.
 - 18. A composition of claim 17, wherein the polypeptide comprises an amino acid sequence selected from the group consisting of SEQ ID NO:1-20.
- 19. A method for treating a disease or condition associated with decreased expression of functional KAP, comprising administering to a patient in need of such treatment the composition of claim 17.
- 20. A method of screening a compound for effectiveness as an agonist of a polypeptide of claim 1, the method comprising:
 - exposing a sample comprising a polypeptide of claim 1 to a compound, and
 - b) detecting agonist activity in the sample.
- 20 21. A composition comprising an agonist compound identified by a method of claim 20 and a pharmaceutically acceptable excipient.
 - 22. A method for treating a disease or condition associated with decreased expression of functional KAP, comprising administering to a patient in need of such treatment a composition of claim 21.
 - 23. A method of screening a compound for effectiveness as an antagonist of a polypeptide of claim 1, the method comprising:
 - a) exposing a sample comprising a polypeptide of claim 1 to a compound, and
- 30 b) detecting antagonist activity in the sample.

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- 24. A composition comprising an antagonist compound identified by a method of claim 23 and a pharmaceutically acceptable excipient.
- 35 25. A method for treating a disease or condition associated with overexpression of

functional KAP, comprising administering to a patient in need of such treatment a composition of claim 24.

- 26. A method of screening for a compound that specifically binds to the polypeptide of claim 1, the method comprising:
 - a) combining the polypeptide of claim 1 with at least one test compound under suitable conditions, and
 - b) detecting binding of the polypeptide of claim 1 to the test compound, thereby identifying a compound that specifically binds to the polypeptide of claim 1.

27. A method of screening for a compound that modulates the activity of the polypeptide of claim 1, the method comprising:

- a) combining the polypeptide of claim 1 with at least one test compound under conditions permissive for the activity of the polypeptide of claim 1,
- b) assessing the activity of the polypeptide of claim 1 in the presence of the test compound, and
- comparing the activity of the polypeptide of claim 1 in the presence of the test compound with the activity of the polypeptide of claim 1 in the absence of the test compound, wherein a change in the activity of the polypeptide of claim 1 in the presence of the test compound is indicative of a compound that modulates the activity of the polypeptide of claim 1.
- 28. A method of screening a compound for effectiveness in altering expression of a target polynucleotide, wherein said target polynucleotide comprises a sequence of claim 5, the method comprising:
- exposing a sample comprising the target polynucleotide to a compound, under conditions suitable for the expression of the target polynucleotide,
- b) detecting altered expression of the target polynucleotide, and
- c) comparing the expression of the target polynucleotide in the presence of varying amounts of the compound and in the absence of the compound.
 - 29. A method of assessing toxicity of a test compound, the method comprising:
 - a) treating a biological sample containing nucleic acids with the test compound,
- b) hybridizing the nucleic acids of the treated biological sample with a probe comprising at least 20 contiguous nucleotides of a polynucleotide of claim 12 under

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conditions whereby a specific hybridization complex is formed between said probe
and a target polynucleotide in the biological sample, said target polynucleotide
comprising a polynucleotide sequence of a polynucleotide of claim 12 or fragment
thereof.

- 5 c) quantifying the amount of hybridization complex, and
 - d) comparing the amount of hybridization complex in the treated biological sample with the amount of hybridization complex in an untreated biological sample,
 wherein a difference in the amount of hybridization complex in the treated biological sample is indicative of toxicity of the test compound.

30. A diagnostic test for a condition or disease associated with the expression of KAP in a biological sample, the method comprising:

- a) combining the biological sample with an antibody of claim 11, under conditions suitable for the antibody to bind the polypeptide and form an antibody:polypeptide complex, and
- b) detecting the complex, wherein the presence of the complex correlates with the presence of the polypeptide in the biological sample.
- 31. The antibody of claim 11, wherein the antibody is:
- a) a chimeric antibody,

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- b) a single chain antibody,
- c) a Fab fragment,
- d) a F(ab')₂ fragment, or
- e) a humanized antibody.

- 32. A composition comprising an antibody of claim 11 and an acceptable excipient.
- 33. A method of diagnosing a condition or disease associated with the expression of KAP in a subject, comprising administering to said subject an effective amount of the composition of claim 32.
 - 34. A composition of claim 32, wherein the antibody is labeled.
- 35. A method of diagnosing a condition or disease associated with the expression of KAP in a subject, comprising administering to said subject an effective amount of the

composition of claim 34.

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36. A method of preparing a polyclonal antibody with the specificity of the antibody of claim 11, the method comprising:

- a) immunizing an animal with a polypeptide consisting of an amino acid sequence selected from the group consisting of SEQ ID NO:1-20, or an immunogenic fragment thereof, under conditions to elicit an antibody response,
- b) isolating antibodies from said animal, and
- c) screening the isolated antibodies with the polypeptide, thereby identifying a polyclonal antibody which binds specifically to a polypeptide comprising an amino acid sequence selected from the group consisting of SEQ ID NO:1-20.
 - 37. A polyclonal antibody produced by a method of claim 36.
- 15 38. A composition comprising the polyclonal antibody of claim 37 and a suitable carrier.
 - 39. A method of making a monoclonal antibody with the specificity of the antibody of claim 11, the method comprising:
 - a) immunizing an animal with a polypeptide consisting of an amino acid sequence selected from the group consisting of SEQ ID NO:1-20, or an immunogenic fragment thereof, under conditions to elicit an antibody response,
 - b) isolating antibody producing cells from the animal,
 - c) fusing the antibody producing cells with immortalized cells to form monoclonal antibody-producing hybridoma cells,
- 25 d) culturing the hybridoma cells, and
 - e) isolating from the culture monoclonal antibody which binds specifically to a polypeptide comprising an amino acid sequence selected from the group consisting of SEQ ID NO:1-20.
- 30 40. A monoclonal antibody produced by a method of claim 39.
 - 41. A composition comprising the monoclonal antibody of claim 40 and a suitable carrier.
 - 42. The antibody of claim 11, wherein the antibody is produced by screening a Fab expression library.

43. The antibody of claim 11, wherein the antibody is produced by screening a recombinant immunoglobulin library.

- 44. A method of detecting a polypeptide comprising an amino acid sequence selected from the group consisting of SEQ ID NO:1-20 in a sample, the method comprising:
 - a) incubating the antibody of claim 11 with a sample under conditions to allow specific binding of the antibody and the polypeptide, and
- b) detecting specific binding, wherein specific binding indicates the presence of a polypeptide comprising an amino acid sequence selected from the group consisting of SEQ ID NO:1-20 in the sample.
 - 45. A method of purifying a polypeptide comprising an amino acid sequence selected from the group consisting of SEQ ID NO:1-20 from a sample, the method comprising:
 - a) incubating the antibody of claim 11 with a sample under conditions to allow specific binding of the antibody and the polypeptide, and
 - separating the antibody from the sample and obtaining the purified polypeptide comprising an amino acid sequence selected from the group consisting of SEQ ID NO:1-20.
- 20 46. A microarray wherein at least one element of the microarray is a polynucleotide of claim 13.
 - 47. A method of generating an expression profile of a sample which contains polynucleotides, the method comprising:
 - a) labeling the polynucleotides of the sample,

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- b) contacting the elements of the microarray of claim 46 with the labeled polynucleotides of the sample under conditions suitable for the formation of a hybridization complex, and
- c) quantifying the expression of the polynucleotides in the sample.
- 48. An array comprising different nucleotide molecules affixed in distinct physical locations on a solid substrate, wherein at least one of said nucleotide molecules comprises a first oligonucleotide or polynucleotide sequence specifically hybridizable with at least 30 contiguous nucleotides of a target polynucleotide, and wherein said target polynucleotide is a polynucleotide of claim 12.

49. An array of claim 48, wherein said first oligonucleotide or polynucleotide sequence is completely complementary to at least 30 contiguous nucleotides of said target polynucleotide.

- 50. An array of claim 48, wherein said first oligonucleotide or polynucleotide sequence is completely complementary to at least 60 contiguous nucleotides of said target polynucleotide.
 - 51. An array of claim 48, wherein said first oligonucleotide or polynucleotide sequence is completely complementary to said target polynucleotide.
 - 52. An array of claim 48, which is a microarray.
 - 53. An array of claim 48, further comprising said target polynucleotide hybridized to a nucleotide molecule comprising said first oligonucleotide or polynucleotide sequence.
 - 54. An array of claim 48, wherein a linker joins at least one of said nucleotide molecules to said solid substrate.
- 55. An array of claim 48, wherein each distinct physical location on the substrate contains multiple nucleotide molecules, and the multiple nucleotide molecules at any single distinct physical location have the same sequence, and each distinct physical location on the substrate contains nucleotide molecules having a sequence which differs from the sequence of nucleotide molecules at another distinct physical location on the substrate.
 - 56. A polypeptide of claim 1, comprising the amino acid sequence of SEQ ID NO:1.
 - 57. A polypeptide of claim 1, comprising the amino acid sequence of SEQ ID NO:2.
- 30 58. A polypeptide of claim 1, comprising the amino acid sequence of SEQ ID NO:3.
 - 59. A polypeptide of claim 1, comprising the amino acid sequence of SEQ ID NO:4.
 - 60. A polypeptide of claim 1, comprising the amino acid sequence of SEQ ID NO:5.

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	61. A polypeptide of claim 1, comprising the amino acid sequence of SEQ ID NO:6.
	62. A polypeptide of claim 1, comprising the amino acid sequence of SEQ ID NO:7.
5	63. A polypeptide of claim 1, comprising the amino acid sequence of SEQ ID NO:8.
	64. A polypeptide of claim 1, comprising the amino acid sequence of SEQ ID NO:9.
10	65. A polypeptide of claim 1, comprising the amino acid sequence of SEQ ID NO:10.
10	66. A polypeptide of claim 1, comprising the amino acid sequence of SEQ ID NO:11.
	67. A polypeptide of claim 1, comprising the amino acid sequence of SEQ ID NO:12.
15	68. A polypeptide of claim 1, comprising the amino acid sequence of SEQ ID NO:13.
	69. A polypeptide of claim 1, comprising the amino acid sequence of SEQ ID NO:14.
20	70. A polypeptide of claim 1, comprising the amino acid sequence of SEQ ID NO:15.
20	71. A polypeptide of claim 1, comprising the amino acid sequence of SEQ ID NO:16.
	72. A polypeptide of claim 1, comprising the amino acid sequence of SEQ ID NO:17.
25	73. A polypeptide of claim 1, comprising the amino acid sequence of SEQ ID NO:18.
	74. A polypeptide of claim 1, comprising the amino acid sequence of SEQ ID NO:19.
30	75. A polypeptide of claim 1, comprising the amino acid sequence of SEQ ID NO:20.

76. A polynucleotide of claim 12, comprising the polynucleotide sequence of SEQ ID NO:21.

	77. A polynucleotide of claim 12, comprising the polynucleotide sequence of SEQ ID NO:22.
5	78. A polynucleotide of claim 12, comprising the polynucleotide sequence of SEQ ID NO:23.
	79. A polynucleotide of claim 12, comprising the polynucleotide sequence of SEQ ID NO:24.
10	80. A polynucleotide of claim 12, comprising the polynucleotide sequence of SEQ ID NO:25.
	81. A polynucleotide of claim 12, comprising the polynucleotide sequence of SEQ ID NO:26.
15	82. A polynucleotide of claim 12, comprising the polynucleotide sequence of SEQ ID NO:27.
20	83. A polynucleotide of claim 12, comprising the polynucleotide sequence of SEQ ID NO:28.
	84. A polynucleotide of claim 12, comprising the polynucleotide sequence of SEQ ID NO:29.
25	85. A polynucleotide of claim 12, comprising the polynucleotide sequence of SEQ ID NO:30.
,	86. A polynucleotide of claim 12, comprising the polynucleotide sequence of SEQ ID NO:31.
30	87. A polynucleotide of claim 12, comprising the polynucleotide sequence of SEQ ID
	NO:32.
35	88. A polynucleotide of claim 12, comprising the polynucleotide sequence of SEQ ID NO:33.

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89. A polynucleotide of claim 12, comprising the polynucleotide sequence of SEQ ID NO:34.

90. A polynucleotide of claim 12, comprising the polynucleotide sequence of SEQ ID NO:35.

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- 91. A polynucleotide of claim 12, comprising the polynucleotide sequence of SEQ ID NO:36.
- 92. A polynucleotide of claim 12, comprising the polynucleotide sequence of SEQ ID NO:37.
 - 93. A polynucleotide of claim 12, comprising the polynucleotide sequence of SEQ ID NO:38.
 - 94. A polynucleotide of claim 12, comprising the polynucleotide sequence of SEQ ID NO:39.
- 95. A polynucleotide of claim 12, comprising the polynucleotide sequence of SEQ ID NO:40.

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			12	65				1270)			e Trp Leu 1275 1 Val Ser
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			1 r Ile	460 Ser G				LS Le	u Pr			sp Ser Gln
			1 ıl Leu	475 Gly G				146	su .e S∈			1485 le Gln Ala 1500
Th	x Il	e Al	1 a Lys.	490 Leu S	er Il	e Ar	g P			y Gl	Ly Le	eu Glu Ser

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Leu Arg Ala Leu Arg Asp Gly Asn Lys Leu Ala Gln Met Glu Glu
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Glu Lys Ile Gly Ala Gln Ser His Gly Asp Asn Ser Cys Gly Ile
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Glu Ile Val Cys Lys Asp Met Arg Asn Leu Arg Leu Ala Tyr Lys
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Tyr Lys Glu Lys Phe Pro Ile Asn Gly Trp Lys Val Tyr Asp Pro
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Val Ser Glu Tyr Lys Arg Gln Gly Leu Pro Asn Glu Ser Trp Lys
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Tyr Leu Gln Thr Ile Met Asp Ala Asn Ala Gln Ser His Lys Leu
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Ile Ile Phe Asp Ala Arg Gln Asn Ser Val Ala Asp Thr Asn Lys
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Glu Trp Ile Ser Phe Gly His Arg Phe Ala Leu Arg Val Gly His
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Tyr Phe Arg His Leu Trp Asn Lys Ala Leu Leu Arg Ala Cys Ala
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Asp Gly Gly Ala Asn Arg Leu Tyr Asp Ile Thr Glu Gly Glu Arg
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Glu Ser Phe Leu Pro Glu Phe Ile Asn Gly Asp Phe Asp Ser Ile
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Ile Ser Thr Pro Asp Gln Asp His Thr Asp Phe Thr Lys Cys Leu
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Lys Met Leu Gln Lys Lys Ile Glu Glu Lys Asp Leu Lys Val Asp
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Val Ile Val Thr Leu Gly Gly Leu Ala Gly Arg Phe Asp Gln Ile
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Met Ala Ser Val Asn Thr Leu Phe Gln Ala Thr His Ile Thr Pro
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Phe Pro Ile Ile Ile Gln Glu Glu Ser Leu Ile Tyr Leu Leu
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Gln Pro Gly Lys His Arg Leu His Val Asp Thr Gly Met Glu Gly
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Asp Trp Cys Gly Leu Ile Pro Val Gly Gln Pro Cys Met Gln Val
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                                                         195
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 Thr Thr Thr Gly Leu Lys Trp Asn Leu Thr Asn Asp Val Leu Ala
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 Phe Gly Thr Leu Val Ser Thr Ser Asn Thr Tyr Asp Gly Ser Gly
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Cys	Met	Asp	Asp		Tyr	Ala	Ser	Gln		Leu	Glu	Glu	Leu	
Pro	Leu	Leu	Lys	Leu 80	Arg	His	Ala	His	Ile 85	Ser	Val	Tyr	Gln	
Leu	Phe	Ile	Thr	Trp 95	Asn	Gly	Glu	Ile	Ser 100	Ser	Leu	Tyr	Leu	Cys 105
Leu	Val	Met	Glu	Phe 110	Asn	Glu	Leu	Ser	Phe 115	Gln	Glu	Val	Ile	Glu 120
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				170		•	Leu		175					180
				185			Trp		190					195
				200			Ala		205					210
				215			Trp Met		220	-				225
				230			Ser	_	235					240
				245			Gln		250					255
				260			Leu		265					270
				275			His		280					285
				290			Leu		295				_	300
				305			Leu		310					315
				320			Thr		325					330
Leu	Leu	Ser	Met	335 Ala	Leu	Ala	Ser	Tyr	340 Cys	Leu	Val	Pro	Glu	345 Gly
Ser	Leu	Phe	Met		Leu	Ala	Leu	Leu		Met	His	Asp	Gln	360 Trp
Leu	Ser	Cys	Asp		Asp	Arg	Val	Pro		Lys	Arg	qzA	Phe	
Ser	Leu	Gly	Lys		Gly	Lys	Leu	Leu		Pro	Ile	Pro	Lys	
Leu	Pro	Trp	Pro		Glu	Leu	Val	Glu		Val	Val	Thr	Thr	
Glu	Leu	His	Asp	410 Arg 425	Val	Leu	Asp	Val	415 Gln 430	Leu	Cys	Ala	Cys	420 Ser 435
Leu	Leu	Leu	His		Leu	Gly	Gln	Gly		Ile	Val	Asn	Lys	
Pro	Leu	Glu	Lys		Pro	Asp	Leu	Ile		Gln	Val	Leu	Ala	
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				470					475		_	_		480
rrp	ьeu	ъeл	ser	Leu 485	тел	сτλ	Суѕ	тте	Lys 490		Gin	Gln	Phe	Glu 495

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Arg Ala Leu Leu Val Asn Asn Ala Tyr Arg Gly Leu Ala Ser Leu
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Val Lys Val Ser Glu Leu Ala Ala Phe Lys Val Val Val Gln Glu
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Glu Gly Gly Ser Gly Leu Ser Leu Ile Lys Glu Thr Tyr Gln Leu
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His Arg Asp Asp Pro Glu Val Val Glu Asn Val Gly Met Leu Leu
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Val His Leu Ala Ser Tyr Glu Glu Ile Leu Pro Glu Leu Val Ser
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Ser Ser Met Lys Ala Leu Leu Gln Glu Ile Lys Glu Arg Phe Thr
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Ser Ser Leu Glu Leu Val Ser Cys Ala Glu Lys Val Leu Leu Arg
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Leu	Tyr	Leu	Leu		Gln	Ile	Gly	Pro		Ser	Phe	Leu	Ile	
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Lys	Tyr	His	Ser		Arg	Ser	Ser	Arg		Lys	Ala	Pro	Ser	
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Lys	Asp	Glu	Glu		Gln	Met	Cys	Pro		Cys	Leu	Leu	Gly	
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Ser	Pro	Ser	Ser			Ala	Ala	Gln		Gln	Thr	Val	Gln	
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Cys	Leu	Phe	Ser		Asn	Trp	Asn	Val			Met	Ala	Leu	Arg 585
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Glu	Ser	Thr	Gly		Ser	Gly	Gly	Ser		Gly	Ser	Ser	Pro	Ser 615
Gly	G1y	Ala	Thr		Gly	Ser	Ser	Gln	Thr 625		Ile	Ser	Gly	Asp 630
Val	Val	Glu	Ala	Cys 635		Ser	Val	Leu	Ser 640		Val	Cys	Ala	Asp 645
Pro	Val	Туг	. Lys	Val		. Val	. Ala	Ala	Leu 655		Thr	Leu	Arg	Ala 660
Met	. Leu	Va]	Туг		Pro	Сув	His	Ser		Ala	Glu	Arg	Ile	Lys 675
Leu	Gln	Arc	, Leu	Let 680		Pro	Val	Val	Asp 685		Ile	Leu	Val	Lys 690
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Leu	Leu	ı Glı	ı Lev	суя	Lys	Gly	/ Glr	Ala	Gly	Glu	Leu	ı Ala	. Val	Gly

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				Val 830				•	835				040
				Ser 845					850				022
				Thr 860					865				070
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Lys Ala Lys Gln Pro Tyr Arg Glu Asp Thr Glu Trp Leu Lys Gly
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Asp Asn Glu Ala Lys Trp Lys Arg Glu Ile Tyr Gly Arg Gly Leu
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 Ile Met Phe Lys Val Val Pro Val Ser Asp Pro Pro Val Asn Ser
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1117

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                                     100
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Trp Gln Ala Arg Lys Ile Ser Asp Pro Ala Thr Cys Ala Gly Leu
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Val Pro Ser Asn His Leu Leu Lys Arg Lys Gln Arg Glu Phe Trp
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Trp Ser Gln Pro Tyr Gln Pro His Thr Cys Leu Lys Ser Thr Leu
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Tyr Lys Glu Glu Phe Val Gly Tyr Gly Gln Lys Phe Phe Ile Ala
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Gly Phe Arg Arg Ser Met Arg Leu Cys Arg Arg Lys Ser His Leu
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Ala Val Gly Ala Pro Tyr Glu Glu Val Val Arg Tyr Gln Arg Arg
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Pro Ser Asp Lys Tyr Arg Leu Ile Val Leu Met Gly Pro Ser Gly
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Val Gly Val Asn Glu Leu Arg Arg Gln Leu Ile Glu Phe Asn Pro
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 Ser Tyr Glu Thr Asn Gly Arg Glu Tyr His Tyr Val Ser Lys Glu
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Met	Phe	Ser	Pro	Thr 35	Ser	Ala	Pro	Ala		Phe	Leu	Thr	Lys	
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Phe	Leu	Asn	Pro	Arg 65	Ala	His	Ser	Ser	Pro 70	Gly	Thr	Pro	Cys	Ser 75
Ser	Arg	Pro	Leu	Pro 80	Trp	Ser	Cys	Arg	Thr 85	Ser	Asn	Arg	Lys	Ser 90
Leu	Ile	Val	Thr	Ser 95	Ser	Thr	Ser	Pro	Thr 100	Leu	Pro	Arg	Pro	His 105
Ser	Pro	Leu	His	Gly 110	His	Thr	Gly	Asn	Ser 115	Pro	Leu	qzA	Ser	Pro 120
Arg	Asn	Phe	Ser	Pro 125	Asn	Ala	Pro	Ala	His 130	Phe	Ser	Phe	Val	Pro 135
Ala	Arg	Ser	His	Ser 140	His	Arg	Ala	Asp	Arg 145	Thr	Asp	Gly	Arg	Arg 150
Trp	Ser	Leu	Ala	Ser 155	Leu	Pro	Ser	Ser	Gly 160	Tyr	Gly	Thr	Asn	Thr '
Pro	Ser	Ser	Thr	Val 170	Ser	Ser	Ser	Cys	Ser 175	Ser	Gln	Gļu	Lys	Leu 180
His	Gln	Leu	Pro	Phe 185	Gln	Pro	Thr	Ala	Asp 190	Glu	Leu	His	Phe	Leu 195
Thr	Lys	His	Phe	Ser 200	Thr	Glu	Ser	Val	Pro 205	Asp	Glu	Glu	Gly	Arg 210
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Arg	Ser	Pro	Val	Ser 230	Phe	Asp	Ser	Glu	Ile 235	Ile	Met	Met	Asn	His 240
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				260				Asn	265		_			270
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				335				Met	340					345
				350				Phe	355					360
				365				His	370	_				375
				380				Ile	385					390
				395				Ala	400			•		405
		_		410				Asp	415			_		420
				425				Pro	430					435
ייייי	<u>Ile</u>	TÀS	Leu			ASD	Li-LV	Ala		<i>y</i> L:	Ala	-Val	Pho	
Val	Arg	His	Lys	440 Ser 455	Thr	Arg	Gln	Arg	445 Phe 460	Ala	Met	Lys	Lys	450 Ile 465
Asn	Lys	Gln	Asn		Ile	Leu	Arg	Asn		Ile	Gln	Gln	Ala	
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Thr	Ala	Leu	Ser	Leu 980	Leu	Ile	Pro	Ser	Glu 985	His	His	Thr	Cys Ser 990
Pro	Leu	Ala	Ser	Pro 995	Met	Ser	Pro		Ser L000	Gln	Ser	Ser	Asn Pro 1005
Ser	Ser	Arg	_	Ser 1010	Ser	Pro	Ser		Asp L015	Phe	Leu	Pro	Ala Leu 1020
Gly	Ser	Met	_	Pro 1025	Pro	Ile	Ile		His LO30	Arg	Ala	Gly	Lys Lys 1035
Tyr	Gly	Phe		Leu 1040	Arg	Ala	Ile		Val 1045	Tyr	Met	Gly	Asp Ser 1050
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His	Val	Asn	_	Glu 1085	Pro	Val	His		Leu 1090	Val	His	Thr	Glu Val 1095
Val	Glu	Leu		Leu 1100	Lys	Ser	Gly		Lys 1105	Val	Ala	Ile	Ser Thr 1110
Thr	Pro	Leu		Asn 1115	Thr	Ser	Ile		Val 1120	Gly	Pro	Ala	Arg Lys 1125
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				1295					1300				Pro Pro 1305
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				1325					1330				Ala Ala 1335
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_				1370)				1375				Val Leu 1380
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Pro	Ser	Arg	Ala	1385 Leu 1400	Gly	Thr	Leu	Arg	1390 Gln 1405	Asp	Arg	Ala	1395 Glu Arg 1410
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Ala Pro Lys Gly Ala Gly Glu Ser Gly Glu Glu Asp Pro Phe Pro
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Ser Arg Asp Pro Arg Ser Leu Gly Pro Met Val Pro Ser Leu Leu
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Thr Gly Ile Thr Leu Gly Pro Pro Arg Met Glu Ser Pro Ser Gly
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Pro His Arg Arg Leu Gly Ser Pro Gln Ala Ile Glu Glu Ala Ala
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Ser Ser Ser Ser Ala Gly Pro Asn Leu Gly Gln Ser Gly Ala Thr
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                                    1585
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Ser Pro Ser Arg Lys Ala Thr Met Ala Gly Gly Leu Ala Asn Leu
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Pro Arg Glu Gln Gly Lys Thr Gln Pro Pro Ser Ala Pro Arg Leu
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                   80
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Pro Ile Arg Pro Ile Arg Gly Gln Gln Leu Lys Ile Leu Gly
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Leu Val Ala Lys Gly Ser Phe Gly Thr Val Leu Lys Val Leu Asp
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Cys Thr Gln Lys Ala Val Phe Ala Val Lys Val Val Pro Lys Val
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Lys Val Leu Gln Arg Asp Thr Val Arg Gln Cys Lys Glu Glu Val
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Ser Ile Gln Arg Gln Ile Asn His Pro Phe Val His Ser Leu Gly
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Asp Ser Trp Gln Gly Lys Arg His Leu Phe Ile Met Cys Ser Tyr
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Cys Ser Thr Asp Leu Tyr Ser Leu Trp Ser Ala Val Gly Cys Phe
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Asp Phe Gly Leu Ser Arg His Val Pro Gln Gly Ala Gln Ala Tyr
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Gly Gly Pro Tyr Asn His Ala Ala Asp Trp Trp Ser Leu Gly Val
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PCT/US01/47431 WO 02/46384

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Tyr Glu Ile Phe Glu Thr Ser Asp Gly Arg Ile Tyr Ile Ile Met
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Glu Leu Gly Val Gln Gly Asp Leu Leu Glu Phe Ile Lys Cys Gln
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Gly Ala Leu His Glu Asp Val Ala Arg Lys Met Phe Arg Gln Leu
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Ser Ser Ala Val Lys Tyr Cys His Asp Leu Asp Ile Val His Arg
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Asp Leu Lys Cys Glu Asn Leu Leu Asp Lys Asp Phe Asn Ile
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Lys Leu Ser Asp Phe Gly Phe Ser Lys Arg Cys Leu Arg Asp Ser
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Asn Gly Arg Ile Ile Leu Ser Lys Thr Phe Cys Gly Ser Ala Ala
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Tyr Ala Ala Pro Glu Val Leu Gln Ser Ile Pro Tyr Gln Pro Lys
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Val Tyr Asp Ile Trp Ser Leu Gly Val Ile Leu Tyr Ile Met Val
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 Cys Gly Ser Met Pro Tyr Asp Asp Ser Asp Ile Arg Lys Met Leu
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 Arg Ile Gln Lys Glu His Arg Val Asp Phe Pro Arg Ser Lys Asn
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                  230
 Leu Thr Cys Glu Cys Lys Asp Leu Ile Tyr Arg Met Leu Gln Pro
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 Asp Val Ser Gln Arg Leu His Ile Asp Glu Ile Leu Ser His Ser
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                                      265
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 Trp Leu Gln Pro Pro Lys Pro Lys Ala Met Ser Ser Ala Ser Phe
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 Lys Arg Glu Gly Glu Gly Lys Tyr Arg Ala Glu Cys Lys Leu Asp
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 Thr Lys Thr Gly Leu Arg Pro Asp His Arg Pro Asp His Lys Leu
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 Gly Ala Lys Thr Gln His Arg Leu Leu Val Val Pro Glu Asn Glu
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 Asn Arg Met Glu Asp Arg Leu Ala Glu Thr Ser Arg Ala Lys Asp
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  Lys Lys Thr Pro Thr Asp Phe Val Glu Arg Phe Leu Pro Arg Glu
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Met Asp Ile Leu Ala Thr Val Asn His Gly Ser Ile Ile Lys Thr
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Tyr Glu Ile Phe Glu Thr Ser Asp Gly Arg Ile Tyr Ile Ile Met
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Glu Leu Gly Val Gln Gly Asp Leu Leu Glu Phe Ile Lys Cys Gln
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Gly Ala Leu His Glu Asp Val Ala Arg Lys Met Phe Arg Gln Leu
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Ser Ser Ala Val Lys Tyr Cys His Asp Leu Asp Ile Val His Arg
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Asp Leu Lys Cys Glu Asn Leu Leu Leu Asp Lys Asp Phe Asn Ile
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Lys Leu Ser Asp Phe Gly Phe Ser Lys Arg Cys Leu Arg Asp Ser
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Asn Gly Arg Ile Ile Leu Ser Lys Thr Phe Cys Gly Ser Ala Ala
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                                     175
                                                          180
Tyr Ala Ala Pro Glu Val Leu Gln Ser Ile Pro Tyr Gln Pro Lys
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                                     190
                                                          195
Val Tyr Asp Ile Trp Ser Leu Gly Val Ile Leu Tyr Ile Met Val
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                                     205
Cys Gly Ser Met Pro Tyr Asp Asp Ser Asp Ile Arg Lys Met Leu
                215
                                     220
Arg Ile Gln Lys Glu His Arg Val Asp Phe Pro Arg Ser Lys Asn
                230
                                     235
Leu Thr Cys Glu Cys Lys Asp Leu Ile Tyr Arg Met Leu Gln Pro
                245
                                     250
Asp Val Ser Gln Arg Leu His Ile Asp Glu Ile Leu Ser His Ser
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                                                          270
                                     265
Trp Leu Gln Pro Pro Lys Pro Lys Ala Thr Ser Ser Ala Ser Phe
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                                     280
                                                          285
Lys Arg Glu Gly Gly Lys Tyr Arg Ala Glu Cys Lys Leu Asp
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                                     295
Thr Lys Thr Gly Leu Arg Pro Asp His Arg Pro Asp His Lys Leu
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Gly Ala Lys Thr Gln His Arg Leu Leu Val Val Pro Glu Asn Glu
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                                       40
                 Dbo
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 Val Ala Val Phe Val Phe Asp Lys Lys Leu Ile Asp Lys Tyr Gln
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                                      70
 Lys Phe Glu Lys Asp Gln Ile Ile Asp Ser Leu Lys Arg Gly Val
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                                       85
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Gln Gln Leu Thr Arg Leu Arg His Pro Arg Leu Leu Thr Val Gln
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His Pro Leu Glu Glu Ser Arg Asp Cys Leu Ala Phe Cys Thr Glu
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Pro Val Phe Ala Ser Leu Ala Asn Val Leu Gly Asn Trp Glu Asn
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                                     130
                 125
Leu Pro Ser Pro Ile Ser Pro Asp Ile Lys Asp Tyr Lys Leu Tyr
                                                          150
                                     145
                 140
Asp Val Glu Thr Lys Tyr Gly Leu Leu Gln Val Ser Glu Gly Leu
                                                          165
                                     160
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Ser Phe Leu His Ser Ser Val Lys Met Val His Gly Asn Ile Thr
                                     175
                 170
Pro Glu Asn Ile Ile Leu Asn Lys Ser Gly Ala Trp Lys Ile Met
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                 185
Gly Phe Asp Phe Cys Val Ser Ser Thr Asn Pro Ser Glu Gln Glu
                                                          210
                                      205
                 200
Pro Lys Phe Pro Cys Lys Glu Trp Asp Pro Asn Leu Pro Ser Leu
                                      220
                 215
Cys Leu Pro Asn Pro Glu Tyr Leu Ala Pro Glu Tyr Ile Leu Ser
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Val Ser Cys Glu Thr Ala Ser Asp Met Tyr Ser Leu Gly Thr Val
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Met Tyr Ala Val Phe Asn Lys Gly Lys Pro Ile Phe Glu Val Asn
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 Lys Gln Asp Ile Tyr Lys Ser Phe Ser Arg Gln Leu Asp Gln Leu
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 Ser Arg Leu Gly Ser Ser Ser Leu Thr Asn Ile Pro Glu Glu Val
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 Arg Glu His Val Lys Leu Leu Leu Asn Val Thr Pro Thr Val Arg
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 Pro Asp Ala Asp Gln Met Thr Lys Ile Pro Phe Phe Asp Asp Val
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 Gly Ala Val Thr Leu Gln Tyr Phe Asp Thr Leu Phe Gln Arg Asp
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 Asn Leu Gln Lys Ser Gln Phe Phe Lys Gly Leu Pro Lys Val Leu
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                                      355
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 Pro Lys Leu Pro Lys Arg Val Ile Val Gln Arg Ile Leu Pro Cys
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 Leu Thr Ser Glu Phe Val Asn Pro Asp Met Val Pro Phe Val Leu
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 Pro Asn Val Leu Leu Ile Ala Glu Glu Cys Thr Lys Glu Glu Tyr
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 Val Lys Leu Ile Leu Pro Glu Leu Gly Pro Val Phe Lys Gln Gln
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 Glu Pro Ile Gln Ile Leu Leu Ile Phe Leu Gln Lys Met Asp Leu
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 Leu Leu Thr Lys Thr Pro Pro Asp Glu Ile Lys Asn Ser Val Leu
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  Pro Met Val Tyr Arg Ala Leu Glu Ala Pro Ser Ile Gln Ile Gln
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  Glu Leu Cys Leu Asn Ile Ile Pro Thr Phe Ala Asn Leu Ile Asp
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  Tyr Pro Ser Met Lys Asn Ala Leu Ile Pro Arg Ile Lys Asn Ala
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  Cys Leu Gln Thr Ser Ser Leu Ala Val Arg Val Asn Ser Leu Val
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  Cys Leu Gly Lys Ile Leu Glu Tyr Leu Asp Lys Trp Phe Val Leu
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  Asp Asp Ile Leu Pro Phe Leu Gln Gln Ile Pro Ser Lys Glu Pro
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                   530
  Ala Val Leu Met Gly Ile Leu Gly Ile Tyr Lys Cys Thr Phe Thr
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                   545
  His Lys Lys Leu Gly Ile Thr Lys Glu Gln Leu Ala Gly Lys Val
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565
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Leu Pro His Leu Ile Pro Leu Ser Ile Glu Asn Asn Leu Asn Leu
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Asn Gln Phe Asn Ser Phe Ile Ser Val Ile Lys Glu Met Leu Asn
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                                    595
Arg Leu Glu Ser Glu His Lys Thr Lys Leu Glu Gln Leu His Ile
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                                    610
Met Gln Glu Gln Gln Lys Ser Leu Asp Ile Gly Asn Gln Met Asn
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                620
Val Ser Glu Glu Met Lys Val Thr Asn Ile Gly Asn Gln Gln Ile
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Asp Lys Val Phe Asn Asn Ile Gly Ala Asp Leu Leu Thr Gly Ser
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Glu Ser Glu Asn Lys Glu Asp Gly Leu Gln Asn Lys His Lys Arg
                                     670
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Ala Ser Leu Thr Leu Glu Glu Lys Gln Lys Leu Ala Lys Glu Gln
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Glu Gln Ala Gln Lys Leu Lys Ser Gln Gln Pro Leu Lys Pro Gln
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Val His Thr Pro Val Ala Thr Val Lys Gln Thr Lys Asp Leu Thr
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Asp Thr Leu Met Asp Asn Met Ser Ser Leu Thr Ser Leu Ser Val
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Ser Thr Pro Lys Ser Ser Ala Ser Ser Thr Phe Thr Ser Val Pro
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Ser Met Gly Ile Gly Met Met Phe Ser Thr Pro Thr Asp Asn Thr
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Lys Arg Asn Leu Thr Asn Gly Leu Asn Ala Asn Met Gly Phe Gln
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Thr Ser Gly Phe Asn Met Pro Val Asn Thr Asn Gln Asn Phe Tyr
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Ser Ser Pro Ser Thr Val Gly Val Thr Lys Met Thr Leu Gly Thr
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Pro Pro Thr Leu Pro Asn Phe Asn Ala Leu Ser Val Pro Pro Ala
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Gly Ala Lys Gln Thr Gln Gln Arg Pro Thr Asp Met Ser Ala Leu
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Asn Asn Leu Phe Gly Pro Gln Lys Pro Lys Val Ser Met Asn Gln
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Leu Ser Gln Gln Lys Pro Asn Gln Trp Leu Asn Gln Phe Val Pro
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Pro Gln Gly Ser Pro Thr Met Gly Ser Ser Val Met Gly Thr Gln
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Leu Asp Pro Gly Gly Arg Gln Leu Lys Val Arg Asp Arg Asn Phe
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                                      70
                  65
Gln Leu Arg Gln Asn Leu Tyr Leu Val Gly Phe Gly Lys Ala Val
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                  80
Leu Gly Met Ala Ala Ala Glu Glu Leu Leu Gly Gln His Leu
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                                     100
                  95
Val Gln Gly Val Ile Ser Val Pro Lys Gly Ile Arg Ala Ala Met
                                      115
                 110
Glu Arg Ala Gly Lys Gln Glu Met Leu Leu Lys Pro His Ser Arg
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                                                          135
                 125
Val Gln Val Phe Glu Gly Ala Glu Asp Asn Leu Pro Asp Arg Asp
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                                      145
                 140
Ala Leu Arg Ala Ala Leu Ala Ile Gln Gln Leu Ala Glu Gly Leu
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                 155
Thr Ala Asp Asp Leu Leu Leu Val Leu Ile Ser Gly Gly Ser
                                      175
                 170
·Ala Leu Leu Pro Ala Pro Ile Pro Pro Val Thr Leu Glu Glu Lys
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                                      190
                 185
Gln Thr Leu Thr Arg Leu Leu Ala Ala Arg Gly Ala Thr Ile Gln
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                 200
Glu Leu Asn Thr Ile Arg Lys Ala Leu Ser Gln Leu Lys Gly Gly
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 Gly Leu Ala Gln Ala Ala Tyr Pro Ala Gln Val Val Ser Leu Ile
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 Leu Ser Asp Val Val Gly Asp Pro Val Glu Val Ile Ala Ser Gly
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                 245
 Pro Thr Val Ala Ser Ser His Asn Val Gln Asp Cys Leu His Ile
                                                           270
                                      265
                  260
 Leu Asn Arg Tyr Gly Leu Arg Ala Ala Leu Pro Arg Ser Val Lys
                                      280
                 275
 Thr Val Leu Ser Arg Ala Asp Ser Asp Pro His Gly Pro His Thr
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                  290
 Cys Gly His Val Leu Asn Val Ile Ile Gly Ser Asn Val Leu Ala
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 Leu Ala Glu Ala Gln Arg Gln Ala Glu Ala Leu Gly Tyr Gln Ala
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 Val Val Leu Ser Ala Ala Met Gln Gly Asp Val Lys Ser Met Ala
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 Gln Phe Tyr Gly Leu Leu Ala His Val Ala Arg Thr Arg Leu Thr
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 Pro Ser Met Ala Gly Ala Ser Val Glu Glu Asp Ala Gln Leu His
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                  365
 Glu Leu Ala Ala Glu Leu Gln Ile Pro Asp Leu Gln Leu Glu Glu
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 Ala Leu Glu Thr Met Ala Trp Gly Arg Gly Pro Val Cys Leu Leu
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  Ala Gly Gly Glu Pro Thr Val Gln Leu Gln Gly Ser Gly Arg Gly
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                                       415
                  410
  Gly Arg Asn Gln Glu Leu Ala Leu Arg Val Gly Ala Glu Leu Arg
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                                       430
                  425
  Arg Trp Pro Leu Gly Pro Ile Asp Val Leu Phe Leu Ser Gly Gly
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                                       445
                   440
  Thr Asp Gly Gln Asp Gly Pro Thr Glu Ala Ala Gly Ala Trp Val
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                   455
  Thr Pro Glu Leu Ala Ser Gln Ala Ala Glu Gly Leu Asp Ile
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  Ala Thr Phe Leu Ala His Asn Asp Ser His Thr Phe Phe Cys Cys
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                   485
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Asn Val Met Asp Thr His Leu Leu Phe Leu Arg Pro Arg
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Val Gly Lys Lys Ile Gly Cys Gly Asn Phe Gly Glu Leu Arg Leu
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                                      55
Gly Lys Asn Leu Tyr Thr Asn Glu Tyr Val Ala Ile Lys Leu Glu
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Pro Ile Lys Ser Arg Ala Leu Gln Leu His Leu Glu Tyr Arg Phe
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                                      85
Tyr Lys Gln Leu Gly Ser Ala Gly Glu Gly Leu Pro Gln Val Tyr
                 95
                                    100
Tyr Phe Gly Pro Cys Gly Lys Tyr Asn Ala Met Val Leu Glu Leu
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                110
Leu Gly Pro Ser Leu Glu Asp Leu Phe Asp Leu Cys Asp Arg Thr
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Phe Thr Leu Lys Thr Val Leu Met Ile Ala Ile Gln Leu Leu Ser
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                                     145
Arg Met Glu Tyr Val His Ser Lys Asn Leu Ile Tyr Arg Asp Val
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                                                         165
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Lys Pro Glu Asn Phe Leu Ile Gly Arg Gln Gly Asn Lys Lys Glu
                                     175
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His Val Ile His Ile Ile Asp Phe Gly Leu Ala Lys Glu Tyr Ile
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                                     190
Asp Pro Glu Thr Lys Lys His Ile Pro Tyr Arg Glu His Lys Ser
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                                                         210
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Leu Thr Gly Thr Ala Arg Tyr Met Ser Ile Asn Thr His Leu Gly
                                     220
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Lys Glu Gln Ser Arg Arg Asp Asp Leu Glu Ala Leu Gly His Met
Phe Met Tyr Phe Leu Arg Gly Ser Leu Pro Trp Gln Gly Leu Lys
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Ala Asp Thr Leu Lys Glu Arg Tyr Gln Lys Ile Gly Asp Thr Lys
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Arg Asn Thr Pro Ile Glu Ala Leu Cys Glu Asn Phe Pro Glu Glu
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                                     280
Met Ala Thr Tyr Leu Arg Tyr Val Arg Arg Leu Asp Phe Phe Glu
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Lys Pro Asp Tyr Glu Tyr Leu Arg Thr Leu Phe Thr Asp Leu Phe
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                 305
                                                          315
Clu bys bys Gly Tyr Thr Phe Asp Tyr Alu
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Arg Pro Ile Pro Thr Pro Val Gly Ser Val His Val Asp Ser Gly
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                                     340
Ala Ser Ala Ile Thr Arg Glu Ser His Thr His Arg Asp Arg Pro
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Arg Gly Glu Trp Glu Ile Gln Pro Ser Arg Gln Thr Asn Thr Ser
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                                     385
                380
Tyr Leu Thr Ser His Leu Ala Ala Asp Arg His Gly Gly Ser Val
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                                     400
                395
Gln Val Val Ser Ser Thr Asn Gly Glu Leu Asn Val Asp Asp Pro
                                                          420
                                     415
                 410
Thr Gly Ala His Ser Asn Ala Pro Ile Thr Ala His Ala Glu Val
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 Ala Gly Thr Trp Arg Ala Ala Val Glu Cys Ser Gly Arg Gly Leu
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 Gly Ala Ala Ser Glu Ser Pro Gln Cys Pro Pro Pro Gly Val
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 Glu Gly Ala Ala Gly Pro Ala Glu Pro Asp Gly Ala Ala Glu Gly
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                                       85
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 Ala Ala Gly Gly Ser Gly Glu Gly Glu Ser Gly Gly Pro Arg
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 Arg Ala Leu Arg Ala Val Tyr Val Arg Ser Glu Ser Ser Gln Gly
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 Gly Ala Ala Gly Gly Pro Glu Ala Gly Ala Arg Gln Cys Leu Leu
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                                      130
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 Arg Ala Cys Glu Ala Glu Gly Ala His Leu Thr Ser Val Pro Phe
                                      145
                  140
 Gly Glu Leu Asp Phe Gly Glu Thr Ala Val Leu Asp Ala Phe Tyr
                                                           165
                                      160
                  155
 Asp Ala Asp Val Ala Val Val Asp Met Ser Asp Val Ser Arg Gin
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                  170
 Pro Ser Leu Phe Tyr His Leu Gly Val Arg Glu Ser Phe Asp Met
                                      190
                  185
 Ala Asn Asn Val Ile Leu Tyr His Asp Thr Asp Ala Asp Thr Ala
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                  200
  Leu Ser Leu Lys Asp Met Val Thr Gln Lys Asn Thr Ala Ser Ser
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                  215
  Gly Asn Tyr Tyr Phe Ile Pro Tyr Ile Val Thr Pro Cys Thr Asp
                                                           240
                                       235
                  230
  Tyr Phe Cys Cys Glu Ser Asp Ala Gln Arg Arg Ala Ser Glu Tyr
                                                            255
                                       250
                  245
  Met Gln Pro Asn Trp Asp Asn Ile Leu Gly Pro Leu Cys Met Pro
                                                            270
                                       265
                  260
  Leu Val Asp Arg Phe Ile Ser Leu Leu Lys Asp Ile His Val Thr
                                                            285
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Car	Cys	Va l	ጥ ን ድንድ	ሙሙ	Lare	Glu	ሞኮኍ	T.O.I	T.OU	Acn	Acn	Tle	Δrα	Lare
	_		_	290	_				295		_			300
Ala	Arg	Glu	ГÀЗ	Tyr 305	Gln	Gly	Glu	Glu	Leu 310	Ala	Lys	Glu	Leu	Ala 315
Arg	Ile	Lys	Leu	Arg 320	Met	Asp	Asn	Thr	Glu 325	Val	Leu	Thr	Ser	Asp 330
Ile	Ile	Ile	Asn		Leu	Leu	Ser	Tyr	_	Asp	Ile	Gln	Asp	
Asp	Ala	Met	Val	Lys	Leu	Val	Glu	Thr	Leu	Glu	Met	Leu	Pro	Thr
Cys	Asp	Leu	Ala		Gln	His	Asn	Ile		Phe	His	Tyr	Ala	
Ala	Leu	Asn	Arg	_	Asn	Ser	Thr	Gly	_	Arg	Glu	Lys	Ala	
Gln	Ile	Met	Leu	380 Gln	Val	Leu	Gln	Ser	385 Cys	Asp	His	Pro	Gly	390 Pro
Asp	Met	Phe	Cys	395 Leu	Cys	Gly	Arg	Ile	400 Tyr	Lys	Asp	Ile	Phe	405 Leu
Δen	Ser	Δen	Cve	410	Asn	Asn	Thr	Ser	415	Δen	Ser	Δla	Tle	420
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Trp	Tyr	Arg	гуѕ	440	Pne	GIU	ren	GIn	445	ser	ьeu	туr	ser	450
Ile	Asn	Leu	Ala	Val 455	Leu	Leu	Ile	Val	Ala 460	Gly	Gln	Gln	Phe	Glu 465
Thr	Ser	Leu	Glu	Leu 470	Arg	Lys	Ile	Gly	Val 475	Arg	Leu	Asn	Ser	Leu 480
Leu	Gly	Arg	Lys	Gly 485	Ser	Leu	Glu	Lys	Met 490	Asn	Asn	Tyr	Trp	Asp 495
Val	Gly	Gln	Phe		Ser	Val	Ser	Met		Ala	His	Asp	Val	
Lys	Ala	Val	Gln		Ala	Glu	Arg	Leu		Lys	Leu	Lys	Pro	
Val	Trp	Tyr	Leu		Ser	Leu	Val	Gln		Leu	Leu	Leu	Ile	
Arg	Phe	Lys	Lys	Thr	Ile	Ile	Glu	His	Ser	Pro	Arg	Gln	Glu	Arg
Leu	Asn	Phe	Trp		Asp	Ile	Ile	Phe	_	Ala	Thr	Asn	Glu	
Thr	Asn	Gly	Leu		Phe	Pro	Val	Leu		Ile	Glu	Pro	Thr	
Va1	Tyr	Gln	Pro		Tyr	Val	Ser	Ile		Asn	Glu	Ala	Glu	
Arg	Thir	Val	Ser	590 Leu	Trp	His	Val	Ser	595 Pro	Thr	Glu	Met	Lys	600 Gln
Met	: His	Glu	Tro	605 Asn	Phe	Thr	Ala	Ser	610 Ser	Ile	Lvs	Glv	Ile	615 Ser
			_	620					625		_	_		630
	Ser			635					640					645
	Ser			650					655					660
Ser	Arg	Phe	Phe	Ser 665	Leu	Val	Lys	Glu	Met 670		Thr	Asn	Thr	Ala 675
Gly	/ Ser	Thr	Val	Glu 680	Leu	Glu	Gly	Glu	Thr 685	_	Gly	Asp	Thr	Leu 690
Glu	ı Tyr	Glu	Tyr	Asp 695	His	Asp	Ala	Asn	Gly 700		Arg	Val	Val	Leu 705
Gly	/ Lys	Gly	Thr		Gly	Ile	Val	Tyr		Gly	Arg	Asp	Leu	Ser 720
Asr	ı Gln	Val	Arg	Ile	Ala	Ile	Lys	Glu	Ile	Pro	Glu	Arg	Asp	Ser
Arg	y Tyr	Ser	Gln		Leu	His	Glu	Glu		Ala	Leu	His	Lys	<u> </u>
Let	ı Lys	His	Arg	740 Asn	Ile	Val	Gln	Tyr	745 Leu		Ser	Val	Ser	750 Glu

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760
Asn Gly Tyr Ile Lys Ile Phe Met Glu Gln Val Pro Gly Gly Ser
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Leu Ser Ala Leu Leu Arg Ser Lys Trp Gly Pro Met Lys Glu Pro
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Thr Ile Lys Phe Tyr Thr Lys Gln Ile Leu Glu Gly Leu Lys Tyr
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Leu His Glu Asn Gln Ile Val His Arg Asp Ile Lys Gly Asp Asn
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Val Leu Val Asn Thr Tyr Ser Gly Val Val Lys Ile Ser Asp Phe
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Gly Thr Ser Lys Arg Leu Ala Gly Val Asn Pro Cys Thr Glu Thr
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Phe Thr Gly Thr Leu Gln Tyr Met Ala Pro Glu Ile Ile Asp Gln
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Gly Pro Arg Gly Tyr Gly Ala Pro Ala Asp Ile Trp Ser Leu Gly
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Cys Thr Ile Ile Glu Met Ala Thr Ser Lys Pro Pro Phe His Glu
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Leu Gly Glu Pro Gln Ala Ala Met Phe Lys Val Gly Met Phe Lys
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Ile His Pro Glu Ile Pro Glu Ala Leu Ser Ala Glu Ala Arg Ala
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Phe Ile Leu Ser Cys Phe Glu Pro Asp Pro His Lys Arg Ala Thr
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Thr Ala Glu Leu Leu Arg Glu Gly Phe Leu Arg Gln Val Asn Lys
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 Gly Lys Lys Asn Arg Ile Ala Phe Lys Pro Ser Glu Gly Pro Arg
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 Gly Val Val Leu Ala Leu Pro Thr Gln Gly Glu Pro Met Ala Thr
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 Leu Trp Glu Glu Gln Asn Gln Val Ala Ser Asn Leu Gln Glu Cys
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 Val Ala Gln Ser Ser Glu Glu Leu His Leu Ser Val Gly His Ile
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 Lys Gln Ile Ile Gly
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Lys	Met	Val			Gln	Leu	Glu	Thr		Asn	His	Lys	Met	Val 1200

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				350					355					360
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				560				Lys	565					570
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Leu	Leu	Asp	Ile	Leu 35	Ile	Cys	Leu	Tyr	Asp 40	Glu	Cys	Asn	Asn	
Pro	Leu	Arg	Arg	Glu 50	Lys	Asn	Ile	Leu	Glu 55	Tyr	Leu	Glu	Trp	
Lys	Pro	Phe	Thr	Ser 65	Lys	Val	Lys	Gln	Met 70	Arg	Leu	His	Arg	Glu 75
Asp	Phe	Glu	Ile	Leu 80	Lys	Val	Ile	Gly	Arg 85	Gly	Ala	Phe	Gly	Glu 90
Val	Ala	Val	Val	Lys 95	Leu	Lys	Asn	Ala	Asp 100	Lys	Val	Phe	Ala	Met 105
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	Phe			125					130					135
	Ile			140					145					150
	Leu			155					160					165
	Ser			170					175					180
	Leu			185					190					195
	Tyr			200					205					210
	Asn			215					220					225
	Met			230					235				_	240
	Asp			245					250				_	255
	Gly Met			260					265					270
	Leu			275					280			_		285
	Gln			290					295					300
	Leu			305					310					315
	Asn			320					325			_		330
	Asp			335					340					345
	Val			350					355			_		360
	Cys	•		365					370					375
	Phe			380					385					390
Ser	Ser	Cys	Val	395 Leu	Ser	Asp	Arg	Ser	400 Cys	Leu	Arg	Val	Thr	405 Ala
	Pro			410 Leu					415 Asn					420 Leu
7	7 ~~	7	T	425	m1	61		m	430	•	_		_	435
rsp.	yez	11.37	-1-213	440	-1111111		-∧ <u>-</u> 3	un.	445	A T G	Arg	-I-le	Lyo	Arg
Leu	Glu	Gln	Glu		Leu	Glu	Leu	Ser		Lys	Leu	Gln	Glu	450 Ser 465
Thr	Gln	Thr	Val		Ala	Leu	Gln	Tyr		Thr	Val	Asp	Gly	
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Ile Glu Lys Leu Arg Lys Gln Val Thr Glu Ser Ser His Leu Glu
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 Asp Ser Glu Lys Lys Asn Leu Glu Leu Leu Ser Glu Ile Glu Gln
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                  920
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Ser	Ser	Leu	Ile	Asp 980	Phe	Leu	Asp	Asp	Thr 985	Asp	Pro	Val	Glu Asn 990
Thr	Tyr	Val	Trp		Pro	Ser	Val	_		His	Ile	Gln	Ser Arg 1005
Ser	Thr	Ser		Ser 1010	Thr	Ser	Ser		Ala L015	Glu	Pro	Val	Lys Thr 1020
Val	Asp	Ser	Thr		Leu	Ser	Val	His		Pro	Thr	Leu	Arg Lys 1035
Lys	Gly	Сув	Pro		Ser	Thr	Gly	Phe		Pro	Lys	Arg	Lys Thr 1050
His	G1n	Phe	Phe	-	Lys	Ser	Phe	Thr		Pro	Thr	Lys	Cys His
Gln	Cys	Thr	Ser		Met	Val	Gly	Leu		Arg	Gln	Gly	Cys Ser 1080
Cys	Glu	Val	Cys		Phe	Ser	Суз	His		Thr	Cys	Val	Asn Lys 1095
Ala	Pro	Thr	Thr		Pro	Val	Pro	Pro		Gln	Thr	Lys	Gly Pro 1110
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His	Val	Arg	Ile		Lys	Pro	Ala	Gly		Lys	Lys	Gly	Trp Gln 1140
Arg	Ala	Leu	Ala		Val	Сув	Asp	Phe		Leu	Phe	Leu	Tyr Asp 1155
Ile	Ala	Glu	Gly	_	Ala	Ser	Gln	Pro		Val	Val	Ile	Ser Gln 1170
Val	Ile	Asp	Met		Asp	Glu	Glu	Phe		·Val	Ser	Ser	Val Leu 1185
Ala	Ser	Asp	Val		His	Ala	Ser	Arg		Asp	Ile	Pro	Cys Ile 1200
Phe	Arg	Val	Thr		Ser	Gln	Leu	Ser		Ser	Asn	Asn	Lys Cys 1215
Ser	Ile	Leu	Met		Ala	Asp	Thr	Glu		Glu	Lys	Asn	Lys Trp 1230
Val	Gly	Val	Leu		Glu	Leu	His	Lys		Leu	Lys	Lys	Asn Lys 1245
Phe	Arg	Asp	Arg		Val	Tyr	Val	Pro		G1u	Ala	Tyr	Asp Ser 1260
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Ile	His	Gln	Ile		Leu	Ile	Pro	Asn		Gln	Leu	Val	Ala Val 1320
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Leu	Asp	Gly	Arg		Thr	Asp	Phe	Tyr		Leu	Ser	Glu	Thr Lys
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Thr	Cys	Leu			Ala	Met	Lys			Val	Leu	Суş	Tyr Glu 1380
Leu	Phe	_Gln	_Ser			Ara	His	_Ara	•		Lvs	<u>Glu</u>	Ile Gln
				1385					1390				1395
Val	Pro	Tyr	Asn			Trp	Met	Ala			Ser	Glu	Gln Leu
رحرو	\ \(= \)	G1.,	Dho	1400		G1 v	Phe	יים, ז	1405		Dro	Len	1410 Asn Gly
Cys	val	GTĀ	FIIC	1415		GTĀ	7 116	ne a	1420		FIO	Dea	1425

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Tyr Thr Asp Cys Gln Gly Arg Arg Ser Arg Gln Gln Glu Leu Met
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Leu	Ala	Lys	Ile		Asp	Phe	Gly	Leu		Lys	Trp	Met	Glu	
Ser	Thr	Arg	Met		Tyr	Ile	Glu	Arg		Ala	Leu	Arg	Gly	
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•	-	-		455	- 23	-3 .	.	0 7	460	7	33-	03 -	~ 3	465
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Gln Asp Gly Val Ser Cys Thr Pro Leu Gln Leu Ala Leu Arg Ser
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Arg Lys Gln Gly Ile Met Ser Phe Leu Glu Gly Lys Glu Pro Ser
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For two-letter codes and other abbreviations, refer to the "Guidance Notes on Codes and Abbreviations" appearing at the beginning of each regular issue of the PCT Gazette.

(54) Title: KINASES AND PHOSPHATASES SEQUENCES, AND USE THEREOF

(57) Abstract: The invention provides human kinases and phosphatases (KAP) and polnucleotides which identify and encode KAP. The invention also provides expresson vectors, host cells, antibodies, agonists, and antagonists. The invention also provides methods for diagnosing, treating, or preventing disorders associated with aberrant expression of KAP.



/046384

INTERNATIONAL SEARCH REPORT

PCT/US 01/47431

A. CLASSIFICATION OF SUBJECT MATTER IPC 7 C12N15/54 C12N $\cdot C12N9/16$ C07K16/40 C12N15/55 C12N9/12 C12N15/11 According to International Patent Classification (IPC) or to both national classification and IPC B. FIELDS SEARCHED Minimum documentation searched (classification system followed by classification symbols) C12N IPC 7 Documentation searched other than minimum documentation to the extent that such documents are included in the fields searched Electronic data base consulted during the international search (name of data base and, where practical, search terms used) BIOSIS, EPO-Internal, CHEM ABS Data, EMBASE C. DOCUMENTS CONSIDERED TO BE RELEVANT Relevant to claim No. Citation of document, with indication, where appropriate, of the relevant passages Category ° 1 - 10TOYOOKA SHIN-ICHI ET AL: "HD-PTP: A novel χ protein tyrosine phosphatase gene on human chromosome 3p21.3." BIOCHEMICAL AND BIOPHYSICAL RESEARCH COMMUNICATIONS, 'Online! vol. 278, no. 3 30 November 2000 (2000-11-30), pages 671-678, XP002216615 ISSN: 0006-291X see Fig. 1 sequence of the full-length HD-PTP which shows 99.94% identity with SEQ ID N 1 of the present application in 1636 amino-acid overlap. & DATABASE SWALL 'Online! P,X 1 March 2001 (2001-03-01) "Protein tyrosine phosphatase HD-PTP" retrieved from EMBL, accession no. Q9H3S7 Database accession no. Q9H3S7 the whole document Patent family members are listed in annex. Further documents are listed in the continuation of box C. Special categories of cited documents : "T" later document published after the international filing date or priority date and not in conflict with the application but cited to understand the principle or theory underlying the "A" document defining the general state of the art which is not considered to be of particular relevance invention "X" document of particular relevance; the daimed invention cannot be considered novel or cannot be considered to involve an inventive step when the document is taken alone "E" earlier document but published on or after the international "L" document which may throw doubts on priority claim(s) or which is cited to establish the publication date of another citation or other special reason (as 'specified) document of particular relevance; the claimed invention cannot be considered to involve an inventive step when the document is combined with one or more other such documents, such combination being obvious to a person skilled in the art. "O" document referring to an oral disclosure, use, exhibition or other means Document published prior to the international filing date but later than the priority date claimed *&" document member of the same patent family Date of mailing of the international search report Date of the actual completion of the international search 1 2,03,03 16 October 2002 Authorized officer Name and mailing address of the ISA European Patent Office, P.B. 5818 Patentiaan 2 NL – 2280 HV Rijswijk Tel. (+31-70) 340–2040, Tx. 31 651 epo ni, Fax: (+31-70) 340–3016 Vix, 0

INTERNATIONAL SEARCH REPORT

Intel nal Application No
PCT/US 01/47431

		PC1/US 01/4/431
C.(Continua Category °	cliation of document, with indication, where appropriate, of the relevant passages	Relevant to claim No.
X	WO 00 63392 A (SHIMIZU KENJI ;KYOWA HAKKO KOGYO KK (JP)) 26 October 2000 (2000-10-26) SEQ ID N 1 (claim 4, p58-69) shows 99.9% identity with SEQ ID N 21 of the present application in 1958 nucleot. overlap; SEQ ID N 2 (claim1) shows 99.8% identity with SEQ ID N 1 in 1636 aa overlap	1-13,56
Α	GOEKJIAN P G ET AL: "PROTEIN KINASE C IN THE TREATMENT OF DISEASE: SIGNAL TRANSDUCTION PATHWAYS, INHIBITORS, AND AGENTS IN DEVELOPMENT" CURRENT MEDICINAL CHEMISTRY, BENTHAM SCIENCE PUBLISHERS BV, BE, vol. 6, no. 9, 1999, pages 877-903, XP001009989 ISSN: 0929-8673 the whole document	1-20,22, 23,25-95
Α	WO 96 13592 A (MANDELKOW ECKHARD; MANDELKOW EVA MARIA (DE); MAX PLANCK GESELLSCHA) 9 May 1996 (1996-05-09) the whole document	1-20,22, 23,25-95
Α .	COHEN PHILIP: "The development and therapeutic potential of protein kinase inhibitors." CURRENT OPINION IN CHEMICAL BIOLOGY, vol. 3, no. 4, August 1999 (1999-08), pages 459-465, XP002216616 ISSN: 1367-5931 the whole document	19,20, 22,23, 25-30
	SARMIENTO MAURO ET AL: "Structure-based discovery of small molecule inhibitors targeted to protein tyrosine phosphatase 1B." JOURNAL OF MEDICINAL CHEMISTRY, vol. 43, no. 2, 27 January 2000 (2000-01-27), pages 146-155, XP002216617 ISSN: 0022-2623 the whole document	19,20, 22,23, 25-30

national application No. PCT/US 01/47431

INTERNATIONAL SEARCH REPORT

Box I Observations where certain claims were found unsearchable (Continuation of item 1 of first sheet)
This International Search Report has not been established in respect of certain claims under Article 17(2)(a) for the following reasons:
Claims Nos.: because they relate to subject matter not required to be searched by this Authority, namely:
2. X Claims Nos.: 21, 24 because they relate to parts of the International Application that do not comply with the prescribed requirements to such an extent that no meaningful International Search can be carried out, specifically:
see FURTHER INFORMATION sheet PCT/ISA/210
3. Claims Nos.: because they are dependent claims and are not drafted in accordance with the second and third sentences of Rule 6.4(a).
Box II Observations where unity of invention is lacking (Continuation of item 2 of first sheet)
This International Searching Authority found multiple inventions in this international application, as follows:
see additional sheet
As all required additional search fees were timely paid by the applicant, this International Search Report covers all searchable claims.
2. As all searchable claims could be searched without effort justifying an additional fee, this Authority did not invite payment of any additional fee.
3. As only some of the required additional search fees were timely paid by the applicant, this International Search Report covers only those claims for which fees were paid, specifically claims Nos.:
4. No required additional search fees were timely paid by the applicant. Consequently, this International Search Report is restricted to the invention first mentioned in the claims; it is covered by claims Nos.: 1-20, 22-23, 25-55 (all partially) and 56, 76 (complete)
, 20, 22 20, 20 20 20 20 20 20 20 20 20 20 20 20 20
Remark on Protest The additional search fees were accompanied by the applicant's protest.
No protest accompanied the payment of additional search fees.

Continuation of Box I.2

Claims Nos.: 21, 24

Present claims 21 and 24 relate to a composition defined by reference to a desirable characteristic or property , namely agonist or antagonist of a polypeptide. The claims cover all compounds having this characteristic or property, whereas the application provides support within the meaning of Article 84 EPC and / or disclosure within the meaning of Article 83 EPC for none such products. In the present case, the claims so lack support, and the application so lacks disclosure, that a meaningful search over the whole of the claimed scope is impossible. A meaningful search could not be carried out for the subject-matter of claims 21 and 24 because it is not possible to determine if any of the presently known substances is falling under the terms of these claims. Besides it is noted, that the compounds of claims 21 and 24 are not rendered novel just because of the

that they have been identified by the method of claims 20 or 23, e.g. such

compounds can already exist. (Apart from this, it is also not possible to establish the scope of these claims without testing all known substances, clearly an undue burden).

The applicant's attention is drawn to the fact that claims, or parts of claims, relating to inventions in respect of which no international search report has been established need not be the subject of an international preliminary examination (Rule 66.1(e) PCT). The applicant is advised that the EPO policy when acting as an International Preliminary Examining Authority is normally not to carry out a preliminary examination on matter which has not been searched. This is the case irrespective of whether or not the claims are amended following receipt of the search report or during any Chapter II procedure.

This International Searching Authority found multiple (groups of) inventions in this international application, as follows:

1. Claims: 1-20, 22-23, 25-55 (all partially) and 56, 76 (complete)

The invention 1 relates to a phosphatase polynucleotide sequence (SEQ ID N 20), its encoded polypeptide (SEQ ID N 1), and their use in the diagnosis, treatment and prevention of diseases and disorders.

2. Claims: 1-20, 22-23, 25-55 (all partially) and 57, 77 (complete)

The invention 1 relates to a phosphatase or Kinase polynucleotide sequence (SEQ ID N 21), its encoded polypeptide (SEQ ID N 2), and their use in the diagnosis, treatment and prevention of diseases and disorders.

3. Claims: 1-20, 22-23, 25-55 (all partially) and 58, 78 (complete)

The invention I relates to a phosphatase or kinase polynucleotide sequence (SEQ ID N 23), its encoded polypeptide (SEQ ID N 3), and their use in the diagnosis, treatment and prevention of diseases and disorders.

 Claims: 1-20, 22-23, 25-55 (all partially) and 59, 79 (complete)

The invention I relates to a phosphatase or kinase polynucleotide sequence (SEQ ID N 24), its encoded polypeptide (SEQ ID N 4), and their use in the diagnosis, treatment and prevention of diseases and disorders.

5. Claims: 1-20, 22-23, 25-55 (all partially) and 60, 80 (complete)

The invention 1 relates to a phosphatase or kinase polynucleotide sequence (SEQ ID N 25), its encoded polypeptide (SEQ ID N 5), and their use in the diagnosis, treatment and prevention of diseases and disorders.

6. Claims: 1-20, 22-23, 25-55 (all partially) and 61, 81 (complete)

The invention 1 relates to a phosphatase or kinase polynucleotide sequence (SEQ ID N 26), its encoded polypeptide (SEQ ID N 6), and their use in the diagnosis, treatment and prevention of diseases and disorders.

7. Claims: 1-20, 22-23, 25-55 (all partially) and 62, 82 (complete)

The invention 1 relates to a phosphatase or kinase polynucleotide sequence (SEQ ID N 27), its encoded polypeptide (SEQ ID N 7), and their use in the diagnosis, treatment and prevention of diseases and disorders.

8. Claims: 1-20, 22-23, 25-55 (all partially) and 63, 83 (complete)

The invention 1 relates to a phosphatase or kinase polynucleotide sequence (SEQ ID N 28), its encoded polypeptide (SEQ ID N 8), and their use in the diagnosis, treatment and prevention of diseases and disorders.

9. Claims: 1-20, 22-23, 25-55 (all partially) and 64, 85 (complete)

The invention 1 relates to a phosphatase or kinase polynucleotide sequence (SEQ ID N 29), its encoded polypeptide (SEQ ID N 9), and their use in the diagnosis, treatment and prevention of diseases and disorders.

10. Claims: 1-20, 22-23, 25-55 (all partially) and 65, 85 (complete)

The invention 1 relates to a phosphatase or kinase polynucleotide sequence (SEQ ID N 30), its encoded polypeptide (SEQ ID N 10), and their use in the diagnosis, treatment and prevention of diseases and disorders.

ll. Claims: 1-20, 22-23, 25-55 (all partially) and 66, 86 (complete)

The invention 1 relates to a phosphatase or kinase polynucleotide sequence (SEQ ID N 31), its encoded polypeptide (SEQ ID N 11), and their use in the diagnosis,

treatment and prevention of diseases and disorders.

12. Claims: 1-20, 22-23, 25-55 (all partially) and 67, 87 (complete)

The invention 1 relates to a phosphatase or kinase polynucleotide sequence (SEQ ID N 32), its encoded polypeptide (SEQ ID N 12), and their use in the diagnosis, treatment and prevention of diseases and disorders.

13. Claims: 1-20, 22-23, 25-55 (all partially) and 68, 88 (complete)

The invention 1 relates to a phosphatase or kinase polynucleotide sequence (SEQ ID N 33), its encoded polypeptide (SEQ ID N 13), and their use in the diagnosis, treatment and prevention of diseases and disorders.

14. Claims: 1-20, 22-23, 25-55 (all partially) and 69, 89 (complete)

The invention 1 relates to a phosphatase or kinase polynucleotide sequence (SEQ ID N 34), its encoded polypeptide (SEQ ID N 14), and their use in the diagnosis, treatment and prevention of diseases and disorders.

15. Claims: 1-20, 22-23, 25-55 (all partially) and 70, 90 (complete)

The invention 1 relates to a phosphatase or kinase polynucleotide sequence (SEQ ID N 35), its encoded polypeptide (SEQ ID N 15), and their use in the diagnosis, treatment and prevention of diseases and disorders.

16. Claims: 1-20, 22-23, 25-55 (all partially) and 71, 91 (complete)

The invention I relates to a phosphatase or kinase polynucleotide sequence (SEQ ID N 36), its encoded polypeptide (SEQ ID N 16), and their use in the diagnosis, treatment and prevention of diseases and disorders.

17. Claims: 1-20, 22-23, 25-55 (all partially) and 72, 92 (complete)

The invention 1 relates to a phosphatase or kinase polynucleotide sequence (SEQ ID N 37), its encoded polypeptide (SEQ ID N 17), and their use in the diagnosis, treatment and prevention of diseases and disorders.

18. Claims: 1-20, 22-23, 25-55 (all partially) and 73, 93 (complete)

The invention 1 relates to a phosphatase or kinase polynucleotide sequence (SEQ ID N 38), its encoded polypeptide (SEQ ID N 18), and their use in the diagnosis, treatment and prevention of diseases and disorders.

19. Claims: 1-20, 22-23, 25-55 (all partially) and 74, 94 (complete)

The invention 1 relates to a phosphatase or kinase polynucleotide sequence (SEQ ID N 39), its encoded polypeptide (SEQ ID N 19), and their use in the diagnosis, treatment and prevention of diseases and disorders.

20. Claims: 1-20, 22-23, 25-55 (all partially) and 75, 95 (complete)

The invention 1 relates to a phosphatase or kinase polynucleotide sequence (SEQ ID N 40), its encoded polypeptide (SEQ ID N 20), and their use in the diagnosis, treatment and prevention of diseases and disorders.

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